Designing Functional Genomics Experiments for Successful Analysis

Mark Fernandes CRUK-CI (Original deck: RORY STARK 18/9/2017)

Agenda

- WHY PERFORM EXPERIMENTS?
- WHY THINK ABOUT EXPERIMENTAL DESIGN? WHAT MAKES FOR A WELL DESIGNED EXPERIMENT? KEY ASPECTS OF EXPERIMENTAL DESIGN
 - Experimental variables
 - Power: variance and replicates
- Bias: confounding factors, randomisation, and controls
 DESIGN PARAMETERS FOR FUNCTIONAL SEQUENCING
 EXPERIMENTS

EXPERIMENTAL DESIGN PROCESS ATCRUK-CI





Why Perform Experiments?

Why Think About Experimental Design?





Reproducible Research





Crisis in Reproducible Research <u>http://rpubs.com/neilfws/6577_68</u>

Retraction notices per 100 000 publications by year of Entrez record creation



Do not end up here! <u>https://retractionwatch.com</u>

47 of 53 high-profile cancer studies were not reproducible!



NATURE | COMMENT

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Drug development: Raise standards for preclinical cancer research

C. Glenn Begley & Lee M. Ellis

Affiliations | Corresponding author

Nature **483**, 531–533 (29 March 2012) | doi:10.1038/483531a Published online 28 March 2012





Need for Good Design





Consequences of Poor Experimental Design...

- Cost of experimentation. We have a responsibility to CRUK donors!
- Limited & Precious material, esp. clinical samples.
- Immortalization of data sets in public databases and methods in the literature. Our bad science begets more bad science.
- Ethical concerns of experimentation: animals and clinical samples.





A Well-Designed Experiment: Should have

CLEAR OBJECTIVES

FOCUS AND SIMPLICITY

SUFFICIENT POWER (i.e. detect expected degree of change)

RANDOMISED COMPARISONS

And be

PRECISE UNBIASED AMENABLE TO STATISTICAL ANALYSIS REPRODUCIBLE





Obligatory Statistician quote: Ronald A. Fisher(1890-1962)

"TO CONSULT THE STATISTICIAN AFTER AN EXPERIMENT IS FINISHED IS OFTEN MERELY TOASK HIM TO CONDUCT A POST MORTEM EXAMINATION. HE CAN PERHAPS SAY WHAT THE EXPERIMENT DIED OF." (1938)



Obligatory Statistician quote: Karl Pearson (1857-1936)

"Statistics is the grammar of science."







Aspects of Experimental Design

EXPERIMENTAL FACTORS

VARIABILITY

- Sources of Variance
- Replicates

BIAS

- Confounding factors
- Randomisation wherever a decision is to be made
 - Controls for both measured and unmeasured factors
- Controls





Experimental Factors





Experimental Factors

FACTORS: ASPECTS OF EXPERIMENT THAT CHANGE AND INFLUENCE THE OUTCOME OF THE EXPERIMENT

- e.g. time, weight, drug, gender, ethnicity, country, plate, cage etc.

VARIABLE TYPE DEPENDS ON TYPE OF MEASUREMENT:

- Categorical (nominal), e.g. gender
- Categorical with ordering (ordinal), e.g. tumour grade
- Discrete, e.g. shoe size, number of cells
- Continuous, e.g. body weight in kg, height in cm

INDEPENDENT AND DEPENDENT VARIABLES

- Independent variable (IV): what you change
- Dependent variable (DV): what changes due to IV
- "If (independent variable), then (dependent variable)"



Capturing Variance





Sources of Variation

BIOLOGICAL "NOISE"

- Biological processes are inherently stochastic
- Single cells, cell populations, individuals, organs, species....
- Timepoints, cell cycle, synchronized vs. unsynchronized

TECHNICAL NOISE

- Reagents, antibodies, temperatures, pollution
- Platforms, runs, operators

CONSIDER IN ADVANCE AND CONTROL

REPLICATION REQUIRED TO CAPTURE VARIANCE



Types of Replication

BIOLOGICAL REPLICATION:

- In vivo:
 - Patients
 - Mice
- In vitro:
 - Different cell lines
 - Re-growing cells (passages)



TECHNICAL REPLICATION:

- Experimental protocol
- Measurement platform (i.e. sequencer)



How many samples? WHY DO YOU NEED REPLICATES? CALCULATING APPROPRIATE SAMPLE SIZES

- Power calculations
- Planning for precision
- Resource equation



- Power: the probability of detecting an effect of a specified size if present.
 - Identify and control the sources of variability
 - Biological variability
 - Technical variability
 - Using appropriate numbers of samples (sample size/replicates)
 - Power calculations estimate sample size required to detect an effect *if degree* of variability is known
 - Depends on δ , n, sd, α , H_A
 - If adding samples increases variability, that alone won't add power!

R has tools for this but example of non-R (GUI) tool for power analysis is G*Power <u>http://www.gpower.hhu.de</u>



Confounding Factors and Bias





(according to Gary Larson's Far Side)

Accurate

Biased





Imprecise



(according to Gary Larson's Far Side)

Accurate

Biased



Imprecise





(according to Gary Larson's Far Side) **Biased**

Accurate













(according to Gary Larson's Far Side) **Biased**

Accurate













CAMBRIDGE INSTITUTE



ALSO KNOWN AS EXTRANEOUS, HIDDEN, LURKING OR MASKING FACTORS, OR THE THIRD VARIABLE OR MEDIATOR VARIABLE.

MAY MASK AN ACTUAL ASSOCIATION OR FALSELY DEMONSTRATE AN APPARENT ASSOCIATION BETWEEN THE INDEPENDENT & DEPENDENT VARIABLES.



Confounding Factors

OTHER EXAMPLES:

- Democrats were less satisfied with their sex lives than Republicans. (ABC poll report).
- Slightly overweight people live longer than thin people (US Centre for Disease Control).
 (https://www.nhs.uk/news/obesity/overweight-people-live
 - longer-study-claims/) Use of BMI, existing medical observation, quality of life

INADEQUATE MANAGEMENTAND MONITORING OF CONFOUNDING FACTORS

- one of the most common causes of researchers wrongly assuming that a correlation leads to a causality.

IF A STUDY DOES NOT CONSIDER CONFOUNDING FACTORS, DON'T BELIEVE IT!



Sciencexpress

Report

Genetic Signatures of Exceptional Longevity in Humans

Paola Sebastiani,¹* Nadia Solovieff,¹ Annibale Puca,² Stephen W. Hartley,¹ Efthymia Melista,³ Stacy Andersen,⁴ Daniel A. Dworkis,³ Jemma B. Wilk,⁵ Richard H. Myers,⁵ Martin H. Steinberg,⁶ Monty Montano,³ Clinton T. Baldwin,^{6,7} Thomas T. Perls⁴*

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•GWAS STUDY: 800 CENTENARIANS VS. CONTROLS

•FOUND 150 SNPS PREDICTING CENTENARIANS WITH 77 % ACCURACY

• PROBLEM: THEY USED DIFFERENT SNP CHIPSFOR CENTENARIANS AND CONTROLS

• RETRACTED IN 2011 FOLLOWING INDEPENDENT REVIEWAND QC OF DATA

http://www.the-scientist.com/blog/display/57558/



Technical Confounding Factors: Batch Effects



Control

Treatment 1

Treatment 2

The difference between Control, Treatment 1 and Treatment 2 is confounded by day and plate.



Solutions

RANDOMISATION

- Statistical analysis assume randomised comparisons
- May not see issues caused by non-randomised comparisons
- Make every decision random not arbitrary

BLINDING (removing the influence of condition knowledge)

- Especially important where subjective measurements are taken
- Every experiment should reach its potential degree of blinding



Randomised Block Design

Blocking is the arranging of *experimental units* in groups (blocks) that are similar to one another.



RBD across plates so that each plate contains spatially randomised **equal proportions** of:

- Control
- Treatment 1
- Treatment 2

controlling plate effects.



Randomised Block Design

Good design example: Alzheimer's study from GlaxoSmithKline

Plate effects by *plate*

Left PCA plot show *large plate effects*. Each colour corresponds to a different plate

Plate effects by *case/control*

Right PCA plot shows each plate cluster contains *equal proportions* of cases (blue) and controls (green).



Despite a plate effect being in existence, each plate has the same proportions of cases & controls

Experimental Controls





Experimental Controls

CONTROLLING ERRORS

- Type I: False Positives (reject true H₀)
 - Use Negative controls: A group that should have minimal or no effect
- Type II: False Negative (fail to reject a false H₀)
 - Use Positive controls: A group where known response expected

TECHNICAL CONTROLS

- Detect/correct technical biases
- Normalise measurements (quantification) e.g. RNA spike-in





Examples of Experimental Controls

WILD-TYPE ORGANISM (KNOCKOUTS) INACTIVE SIRNA (SILENCING) VEHICLE (TREATMENTS) INPUT: FRAGMENTED CHROMATIN (CHIP) SPIKE-INS (QUANTIFICATION/NORMALISATION) "GOLD STANDARD" DATAPOINTS MULTI-LEVEL CONTROLS

– e.g. contrast Vehicle/Input vs. Treatment/Input



Design Parameters for Sequencing Experiments





Design Issues: Sequencing Experiments

PLATFORMS LIBRARY PREPS MULTIPLEXING AND POOLING STRATEGIES SINGLE-END VS PAIRED END SEQUENCING DEPTH

- Coverage
- Lanes

VALIDATION

- Knock-downs
- Pull-downs



Experimental Design process at CRUK-CI





Establishing an experimental design process

- Students required to take (this) Experimental Design class
- All sequencing and proteomics experiments require experimental design review meeting
 - Simple form: EDM Form.docx
 - Attended by Scientists, Genomics/Proteomics Core, Bioinformatics Core, Statistician
 - Project opened in LIMS afterwards
- Randomisation and Layouts
 - Checkpoint for experiment
 - Project cleared for sample submission
- Keys:
 - Form and meeting not onerous
 - (Currently) not chargeable
 - Scientists agree process improves experiments!



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CRI Experimental Design Meetings

TUESDAY 30 MIN SLOTS (2:00-3:00PM) WITH BIOINFORMATICS & GENOMICS/PROTEOMICS CORES

DISCUSSION:

- Planning, time-scale, cost, aims, scope, questions
- Choosing the correct technology
- Technical issues e.g. what sequencing depth?
- Sample collection and processing methods
- Sample information (meta-data) collection
- Randomisation, Blocking and Replication issues
- Analyst?
- Pilot study?
- Effect size & Sample-size calculation?



Practical: Investigation into the effect of RARα on transcription in breast cancer tissue treated with estrogen

- RARα is a transcription factor that appears to interact with estrogen (E2) in ER+ breast cancer.
- We are interested in characterising this interaction by looking at how gene expression changes in breast cancer cells treated with estrogen when RARα is not present (using a siRNA in cultured cells).
- We wish to identify which estrogen- induced and estrogenrepressed genes are impacted by the presence or absence of RARα, and to analyse the key pathways involved.



Experimental Design Practical Questions I

- 1. What are your objectives?
- 2. What are you measuring?
- 3. What are your primary sample groups of interest?
- 4. What controls will you use each type of sample group?
- 5. What constitutes a replicate in this experiment? Are they biological or technical? How many samples/replicates should be collected?
- 6. Sketch out the design as a matrix, with sample numbers
- 7. What sample group comparisons (contrasts) will you make with the data? Which gene set(s) will you use for pathway analysis?
- 8. What are possible confounding factors and sources of bias?



Experimental Design Practical Questions I

- 9. How will you confirm effective silencing?
- 10. What information about your experiment should be recorded to help identify any problems should there beany?
- 11. Will you be multiplexing samples? How will you assign barcodes? Will you use pooled libraries? How many pools? How will samples be assigned to pools?
- 12. What are the sequencing parameters you need to be aware of (e.g. sequencing type and depth)?
- 13. What other types of data might be useful to assay, and how might the sequencing parameters need to change to accommodate this?
- 14. Can you think of any other design related issues that could/should be addressed?

