

# Designing Functional Genomics Experiments for Successful Analysis

RORY STARK  
18 SEPTEMBER 2017

# Agenda

**WHY PERFORM EXPERIMENTS?**

**WHY THINK ABOUT EXPERIMENTAL DESIGN?**

**WHAT MAKES FOR A WELL DESIGNED EXPERIMENT?**

**KEY ASPECTS OF EXPERIMENTAL DESIGN**

- Experimental variables
- Power: variance and replicates
- Bias: confounding factors, randomisation, and controls

**DESIGN PARAMETERS FOR FUNCTIONAL SEQUENCING  
EXPERIMENTS**

**EXPERIMENTAL DESIGN PROCESS AT CRUK-CI**

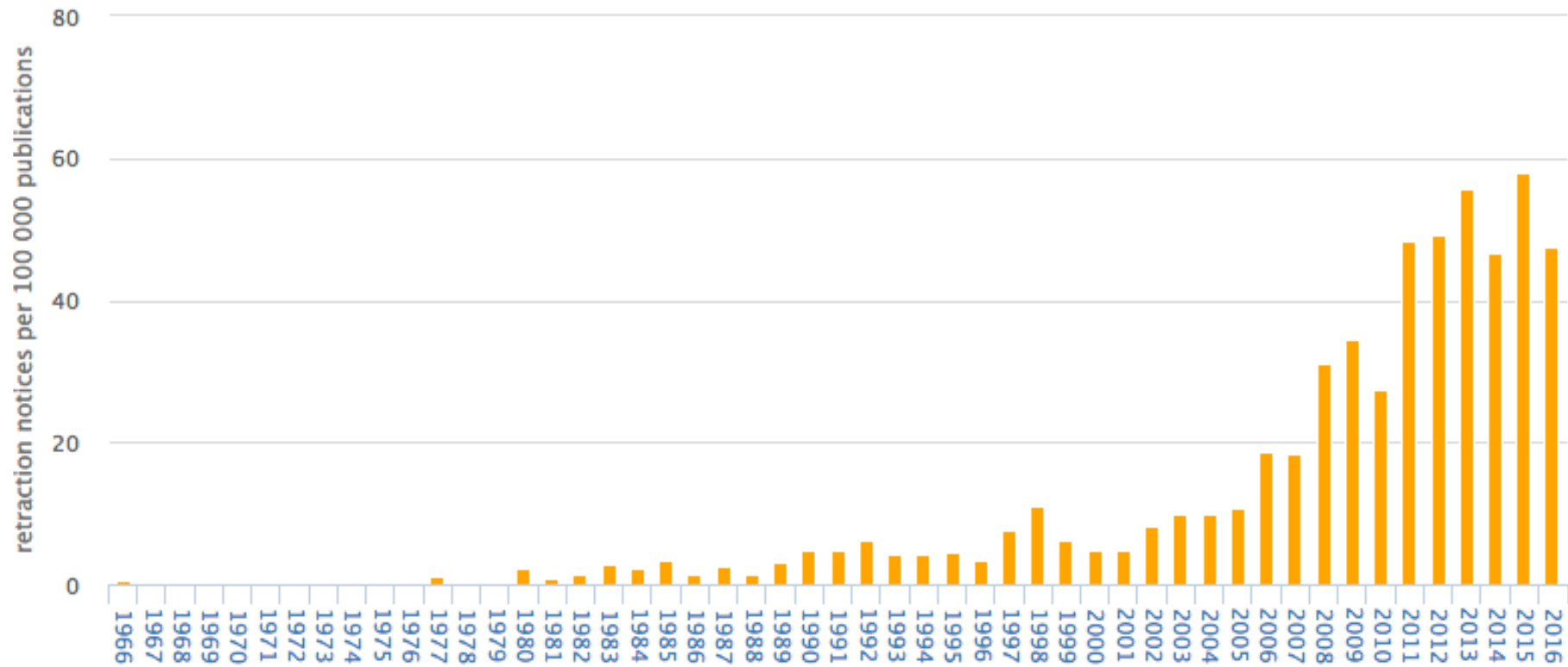
# Why Perform Experiments?

# Why Think About Experimental Design?

# Reproducible Research

# Crisis in Reproducible Research

Retraction notices per 100 000 publications by year of Entrez record creation



# 47 of 53 high-profile cancer studies were not reproducible!



NATURE | COMMENT



## Drug development: Raise standards for preclinical cancer research

**C. Glenn Begley & Lee M. Ellis**

**Affiliations | Corresponding author**

*Nature* **483**, 531–533 (29 March 2012) | doi:10.1038/483531a

Published online 28 March 2012

# Need for Good Design





# Consequences of Poor Experimental Design...

- **Cost** of experimentation. We have a responsibility to CRUK donors!
- **Limited & Precious** material, esp. clinical samples.
- **Immortalization** of data sets in public databases and methods in the literature. Our bad science begets more bad science.
- **Ethical concerns** of experimentation: animals and clinical samples.

# A Well-Designed Experiment:

Should have

CLEAR OBJECTIVES

FOCUS AND SIMPLICITY

SUFFICIENT POWER

RANDOMISED COMPARISONS

And be

PRECISE

UNBIASED

AMENABLE TO STATISTICAL ANALYSIS

REPRODUCIBLE

# Ronald A. Fisher(1890-1962)



*“TO CONSULT THE STATISTICIAN AFTER AN EXPERIMENT IS FINISHED IS OFTEN MERELY TO ASK HIM TO CONDUCT A POST MORTEM EXAMINATION. HE CAN PERHAPS SAY WHAT THE EXPERIMENT DIED OF.” (1938)*

# Aspects of Experimental Design

## EXPERIMENTAL FACTORS

### VARIABILITY

- Sources of Variance
- Replicates

### BIAS

- Confounding factors
- Randomisation wherever a decision is to be made
  - Controls for both measured and unmeasured factors
- Controls

# Experimental Factors



# Experimental Factors

## FACTORS: ASPECTS OF EXPERIMENT THAT CHANGE AND INFLUENCE THE OUTCOME OF THE EXPERIMENT

- e.g. time, weight, drug, gender, ethnicity, country, plate, cage etc.

## VARIABLE TYPE DEPENDS ON TYPE OF MEASUREMENT:

- Categorical (**nominal**) , e.g. gender
- Categorical with ordering (**ordinal**), e.g. tumour grade
- **Discrete**, e.g. shoe size, number of cells
- **Continuous**, e.g. body weight in kg, height in cm

## INDEPENDENT AND DEPENDENT VARIABLES

- Independent variable (IV): what you change
- Dependent variable (DV): what changes due to IV
- “If (**independent** variable), then (**dependent** variable)”



# Capturing Variance

# Sources of Variation

## BIOLOGICAL “NOISE”

- Biological processes are inherently stochastic
- Single cells, cell populations, individuals, organs, species....
- Timepoints, cell cycle, synchronized vs. unsynchronized

## TECHNICAL NOISE

- Reagents, antibodies, temperatures, pollution
- Platforms, runs, operators

**CONSIDER IN ADVANCE AND CONTROL**

***REPLICATION REQUIRED TO CAPTURE VARIANCE***



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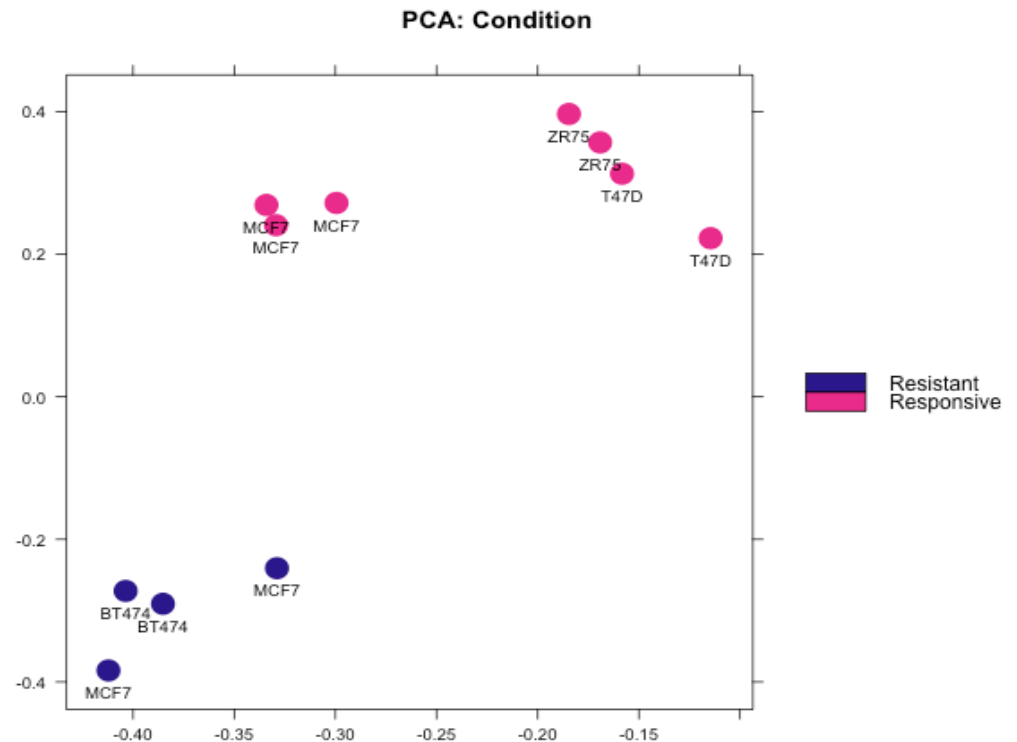
# Types of Replication

## BIOLOGICAL REPLICATION:

- *In vivo*:
  - Patients
  - Mice
- *In vitro*:
  - Different cell lines
  - Re-growing cells (passages)

## TECHNICAL REPLICATION:

- Experimental protocol
- Measurement platform (i.e. sequencer)



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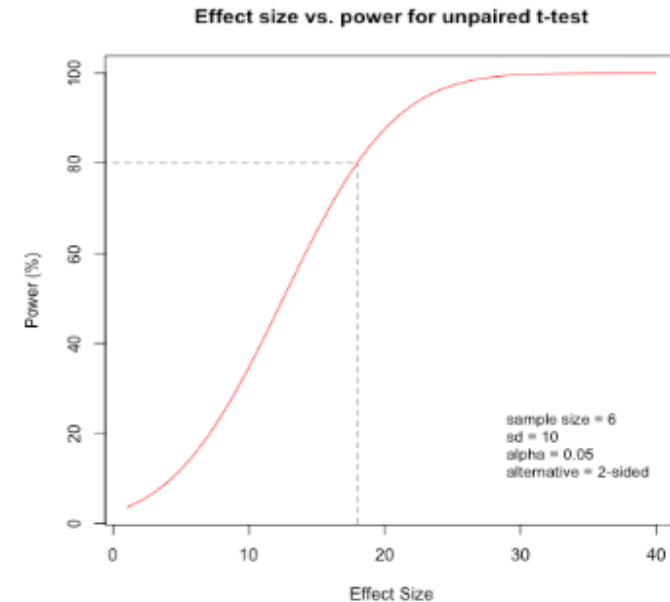
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# How many samples?

## WHY DO YOU NEED REPLICATES?

## CALCULATING APPROPRIATE SAMPLE SIZES

- Power calculations
- Planning for precision
- Resource equation



- Power: the **probability** of detecting an **effect** of a specified size if present.
  - Identify and control the **sources of variability**
    - Biological variability
    - Technical variability
  - Using **appropriate numbers** of samples (sample size/replicates)
  - Power calculations estimate sample size required to detect an effect *if degree of variability is known*
    - Depends on  $\delta$ ,  $n$ ,  $sd$ ,  $\alpha$ ,  $H_A$
  - If adding samples increases variability, that alone won't add power!

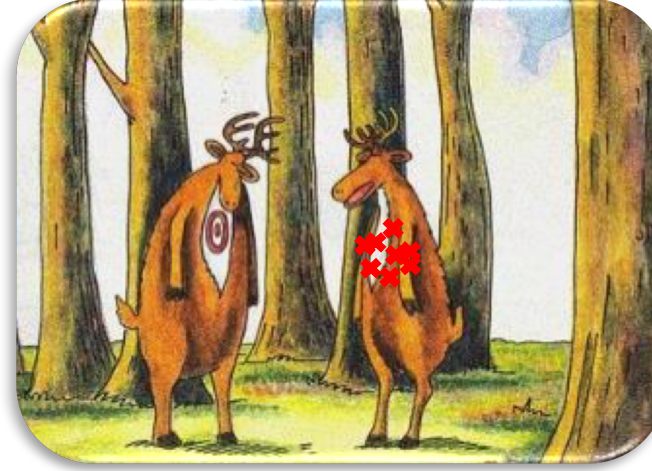
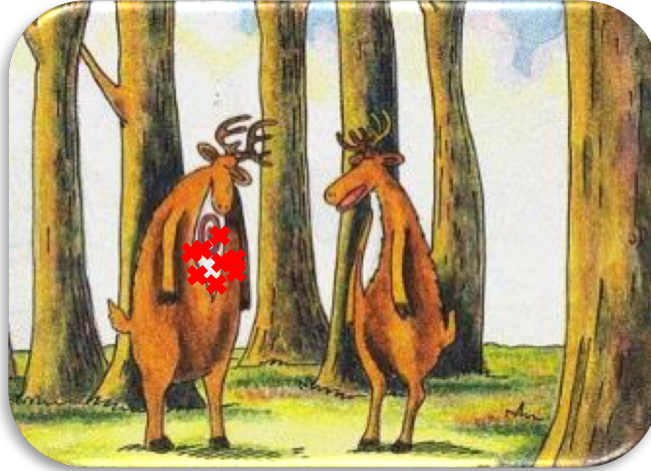
# Confounding Factors and Bias

# Precision, Accuracy & Bias

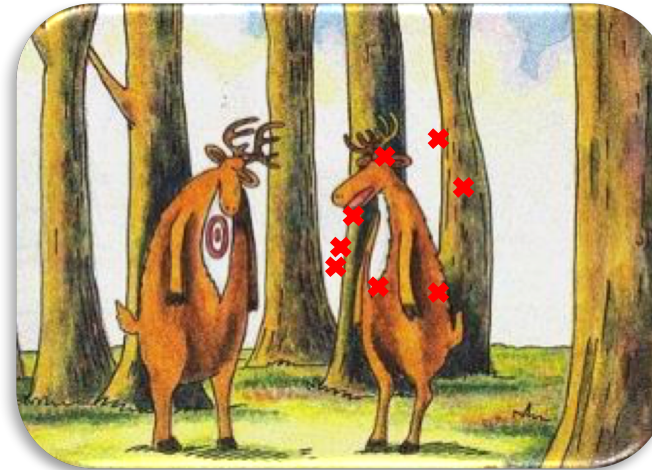
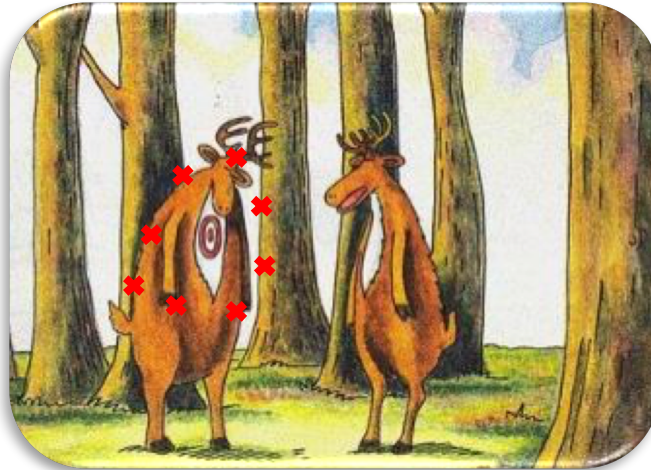
Accurate

Biased

Precise



Imprecise



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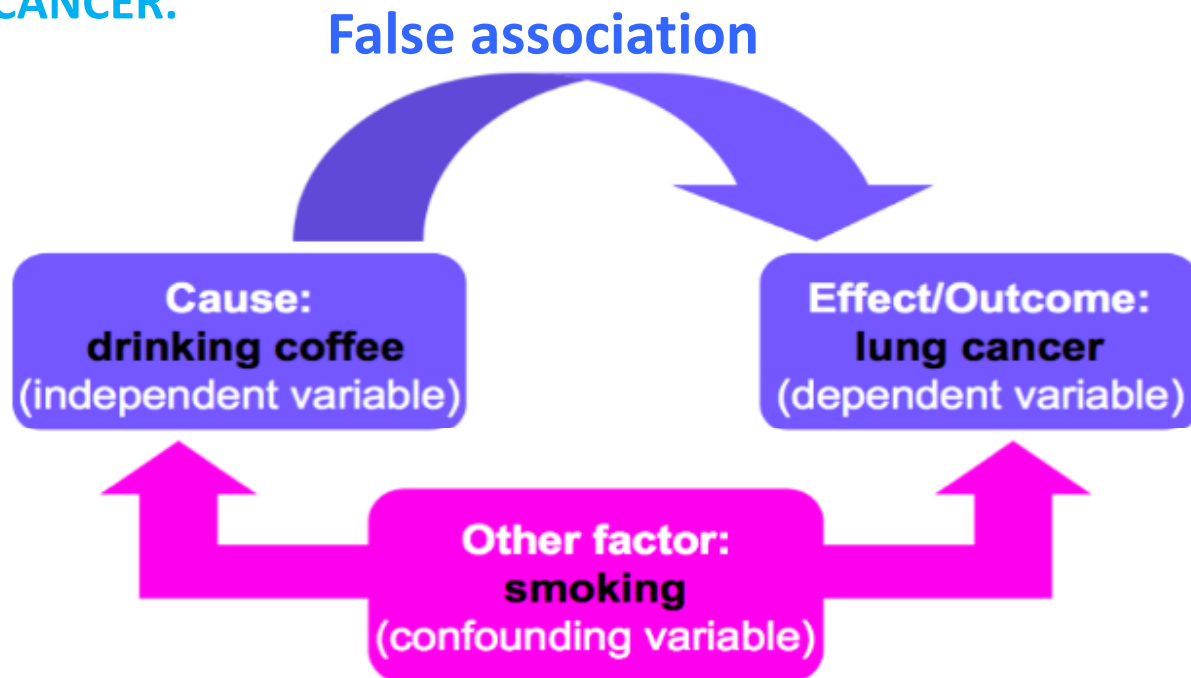
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# Confounding Factors

ALSO KNOWN AS EXTRANEOUS, HIDDEN, LURKING OR MASKING FACTORS,  
OR THE THIRD VARIABLE OR MEDIATOR VARIABLE.

MAY MASK AN ACTUAL ASSOCIATION OR FALSELY DEMONSTRATE AN APPARENT  
ASSOCIATION BETWEEN THE INDEPENDENT & DEPENDENT VARIABLES.

HYPOTHETICAL EXAMPLE WOULD BE A STUDY OF COFFEE DRINKING AND LUNG  
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# Confounding Factors

## OTHER EXAMPLES:

- Democrats were less satisfied with their sex lives than Republicans. (ABC poll report).
- Slightly overweight people live longer than thin people (US Centre for Disease Control).

## INADEQUATE MANAGEMENT AND MONITORING OF CONFOUNDING FACTORS

- one of the most common causes of researchers wrongly assuming that a correlation leads to a causality.

**IF A STUDY DOES NOT CONSIDER CONFOUNDING FACTORS,  
DON'T BELIEVE IT!**



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AN EXPRESSION OF CONCERN HAS BEEN PUBLISHED ON THIS PAPER; SEE LAST PAGE

Scienceexpress

Report

## Genetic Signatures of Exceptional Longevity in Humans

Paola Sebastiani,<sup>1\*</sup> Nadia Solovieff,<sup>1</sup> Annibale Puca,<sup>2</sup> Stephen W. Hartley,<sup>1</sup> Efthymia Melista,<sup>3</sup> Stacy Andersen,<sup>4</sup> Daniel A. Dworkis,<sup>3</sup> Jemma B. Wilk,<sup>5</sup> Richard H. Myers,<sup>5</sup> Martin H. Steinberg,<sup>6</sup> Monty Montano,<sup>3</sup> Clinton T. Baldwin,<sup>6,7</sup> Thomas T. Perls<sup>4\*</sup>

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- **GWAS STUDY: 800 CENTENARIANS VS. CONTROLS**
- **FOUND 150 SNPS PREDICTING CENTENARIANS WITH 77 % ACCURACY**
- **PROBLEM: THEY USED DIFFERENT SNP CHIPS FOR CENTENARIANS AND CONTROLS**
- **RETRACTED IN 2011 FOLLOWING INDEPENDENT REVIEW AND QC OF DATA**



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<http://www.the-scientist.com/blog/display/57558/>

# Solutions

## RANDOMISATION

- Statistical analysis assume randomised comparisons
- May not see issues caused by non-randomised comparisons
- Make every decision random not arbitrary

## BLINDING

- Especially important where subjective measurements are taken
- Every experiment should reach its potential degree of blinding

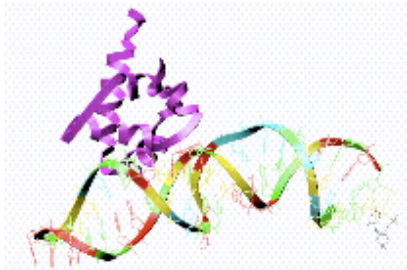


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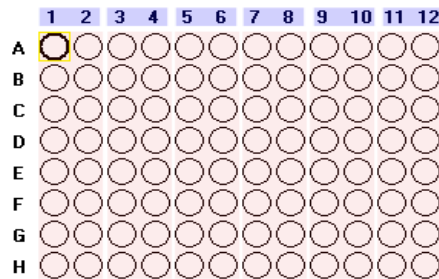


# Technical Confounding Factors: Batch Effects



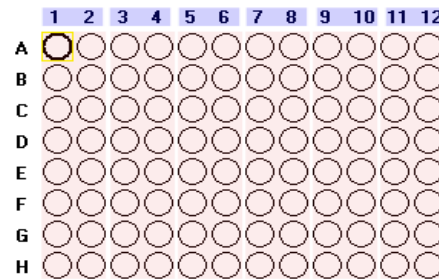
RNA Extraction

Day1, Plate 1



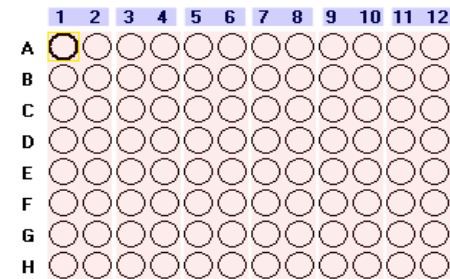
Control

Day2, Plate 2



Treatment 1

Day3, Plate 3



Treatment 2

The difference between Control, Treatment 1 and Treatment 2 is confounded by **day** and **plate**.

# Solutions

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## BLINDING

- Especially important where subjective measurements are taken
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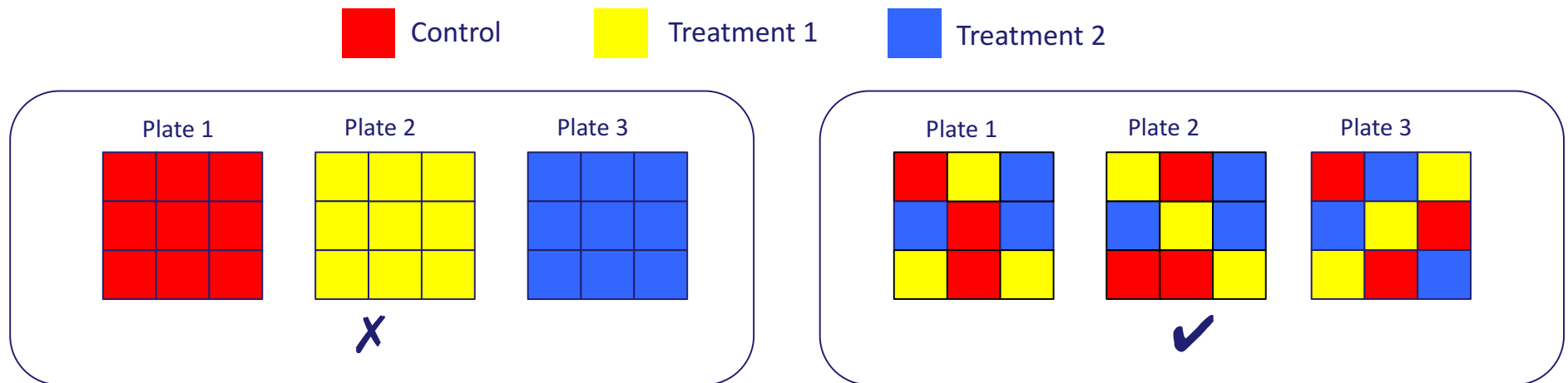


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# Randomised Block Design

**Blocking** is the arranging of *experimental units* in groups (blocks) that are similar to one another.



RBD across plates so that each plate contains spatially randomised **equal proportions** of:

- Control
- Treatment 1
- Treatment 2

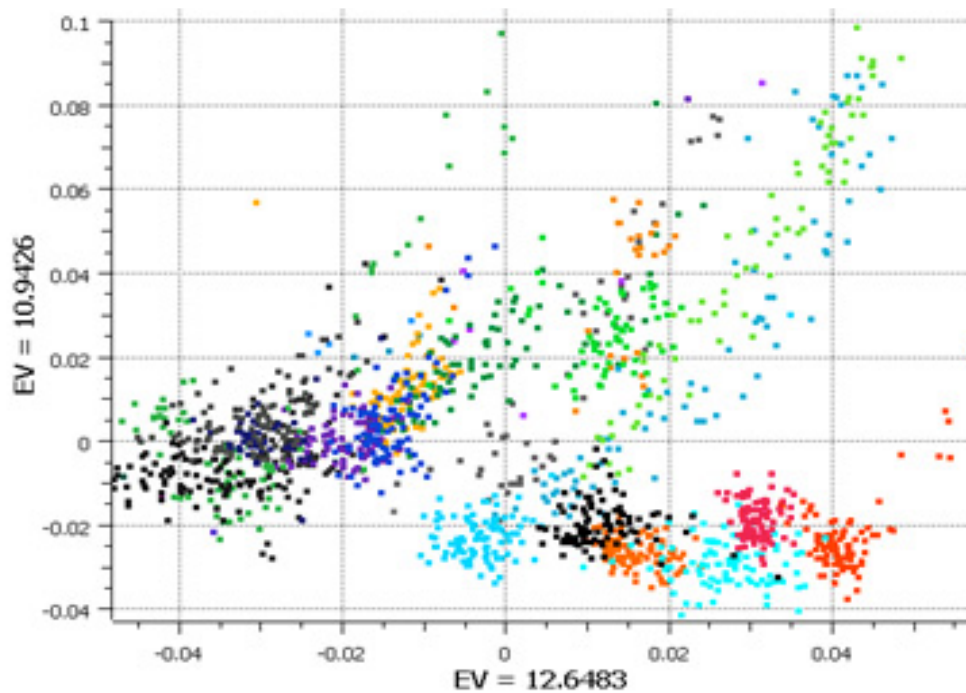
controlling plate effects.

# Randomised Block Design

**Good** design example: Alzheimer's study from GlaxoSmithKline

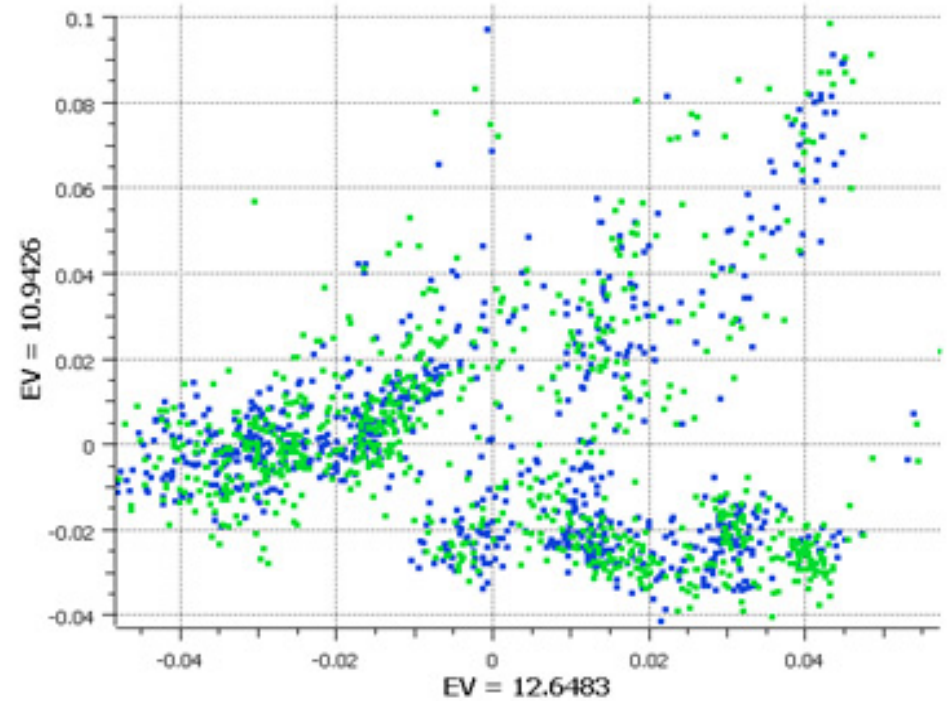
## Plate effects by plate

Left PCA plot show *large plate effects*.  
Each colour corresponds to a different plate



## Plate effects by case/control

Right PCA plot shows each plate cluster contains *equal proportions* of cases (blue) and controls (green).



# Experimental Controls

# Experimental Controls

## CONTROLLING ERRORS

- Type I: FP
  - Negative controls: should have minimal or no effect
- Type II: FN
  - Positive controls: known effect

## TECHNICAL CONTROLS

- Detect/correct technical biases
- Normalise measurements (quantification)



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# Examples of Experimental Controls

WILD-TYPE ORGANISM (KNOCKOUTS)

INACTIVE SIRNA (SILENCING)

VEHICLE (TREATMENTS)

INPUT: FRAGMENTED CHROMATIN (CHIP)

SPIKE-INS (QUANTIFICATION/NORMALISATION)

“GOLD STANDARD” DATAPOINTS

MULTI-LEVEL CONTROLS

- e.g. contrast Vehicle/Input vs. Treatment/Input



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# Design Parameters for Sequencing Experiments





# Design Issues: Sequencing Experiments

## PLATFORMS

## LIBRARY PREPS

## MULTIPLEXING AND POOLING STRATEGIES

## SINGLE-END VS PAIRED END

## SEQUENCING DEPTH

- Coverage
- Lanes

## VALIDATION

- Knock-downs
- Pull-downs



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# Experimental Design process at CRUK-CI



# Establishing an experimental design process

- Students required to take (this) Experimental Design class
- All sequencing and proteomics experiments require experimental design review meeting
  - Simple form: [EDM Form.docx](#)
  - Attended by Scientists, Genomics/Proteomics Core, Bioinformatics Core, Statistician
  - Project opened in LIMS afterwards
- Randomisation and Layouts
  - Checkpoint for experiment
  - Project cleared for sample submission
- Keys:
  - Form and meeting not onerous
  - (Currently) not chargeable
  - Scientists agree process improves experiments!



# Acknowledgements

Others who contributed to  
these lecture notes:

**Sarah Vowler**

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**James Hadfield**

**Jing Su**



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# CRI Experimental Design Meetings

**TUESDAY 30 MIN SLOTS (2:00-3:00PM) WITH BIOINFORMATICS & GENOMICS/PROTEOMICS CORES**

## **DISCUSSION:**

- Planning, time-scale, cost, aims, scope, questions
- Choosing the correct technology
- Technical issues e.g. what sequencing depth?
- Sample collection and processing methods
- Sample information (meta-data) collection
- Randomisation, Blocking and Replication issues
- Analyst?
- Pilot study?
- Effect size & Sample-size calculation?



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# Practical: Investigation into the effect of RAR $\alpha$ on transcription in breast cancer tissue treated with estrogen

- RAR $\alpha$  is a transcription factor that appears to interact with estrogen (E2) in ER+ breast cancer.
- We are interested in characterising this interaction by looking at how gene expression changes in breast cancer cells treated with estrogen when RAR $\alpha$  is not present (using a siRNA in cultured cells).
- We wish to identify which estrogen- induced and estrogen-repressed genes are impacted by the presence or absence of RAR $\alpha$ , and to analyse the key pathways involved.



# Experimental Design Practical Questions I

1. What are your objectives?
2. What are you measuring?
3. What are your primary sample groups of interest?
4. What controls will you use each type of sample group?
5. What constitutes a replicate in this experiment? Are they biological or technical? How many samples/replicates should be collected?
6. Sketch out the design as a matrix, with sample numbers
7. What sample group comparisons (contrasts) will you make with the data? Which gene set(s) will you use for pathway analysis?
8. What are possible confounding factors and sources of bias?



# Experimental Design Practical Questions I

9. How will you confirm effective silencing?
10. What information about your experiment should be recorded to help identify any problems should there be any?
11. Will you be multiplexing samples? How will you assign barcodes? Will you use pooled libraries? How many pools? How will samples be assigned to pools?
12. What are the sequencing parameters you need to be aware of (e.g. sequencing type and depth)?
13. What other types of data might be useful to assay, and how might the sequencing parameters need to change to accommodate this?
14. Can you think of any other design related issues that could/should be addressed?

