

INTRODUCTION TO SINGLE CELL RNA-SEQ

Analysis of single cell RNA-seq data

Katarzyna Kania

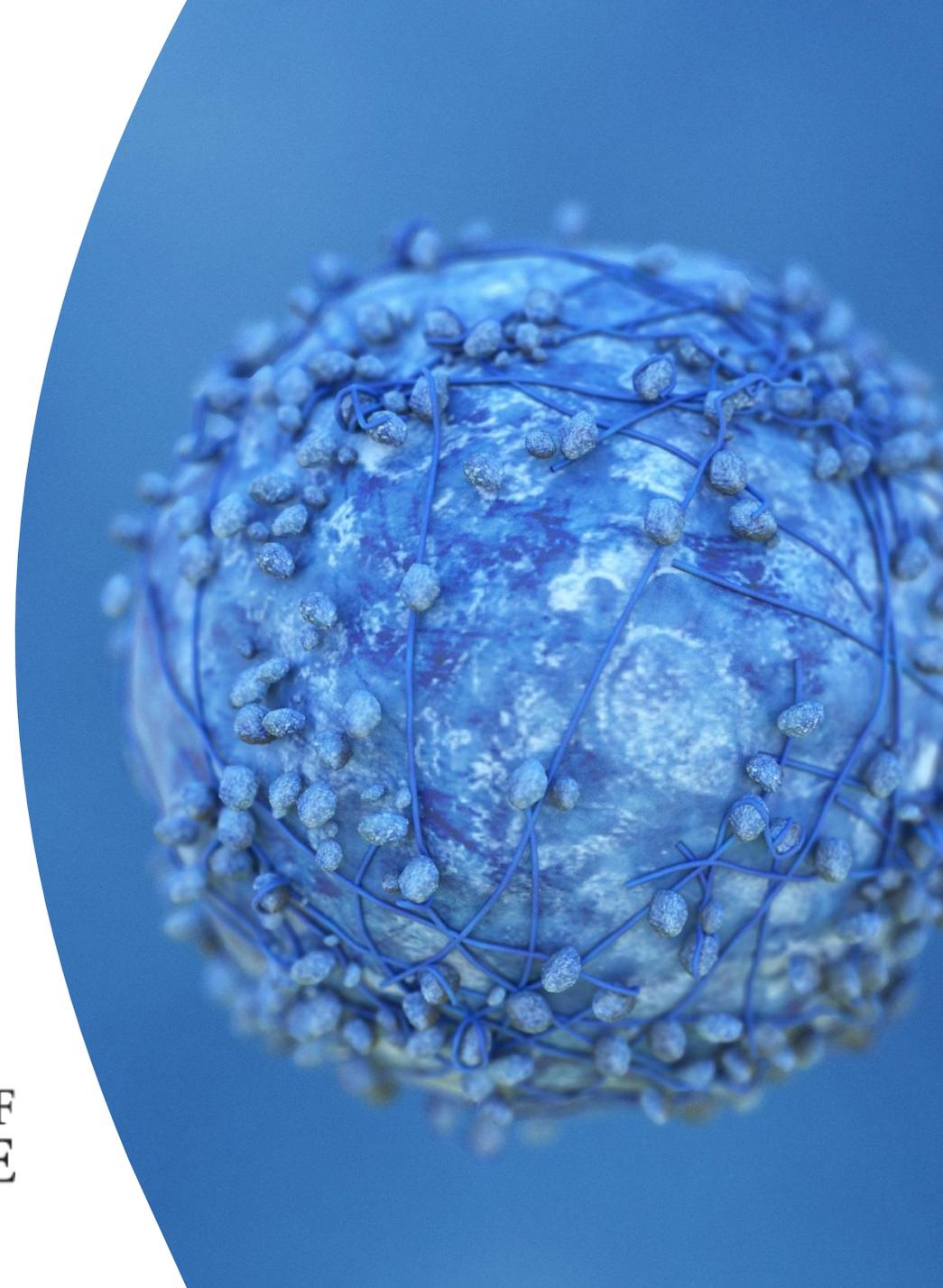
9th February 2026



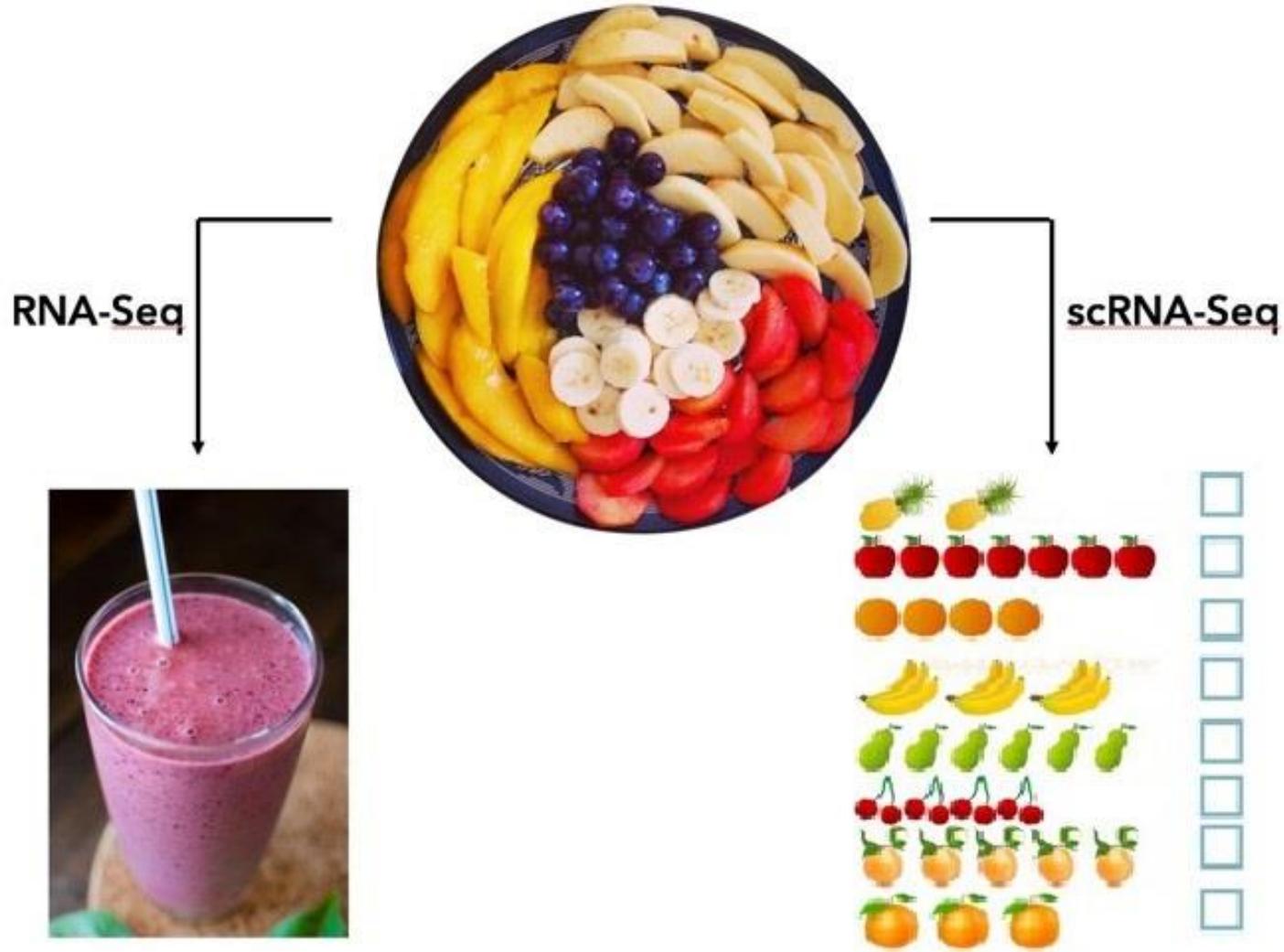
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BULK VS SINGLE CELL RNA-SEQ



RNA-Seq

scRNA-Seq

- Average expression level
- Comparative transcriptomics
 - Disease biomarker
 - Homogenous systems

- Separate populations
- Define heterogeneity
 - Identify rare cell populations
 - Cell population dynamics



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BULK VS SINGLE CELL RNA-SEQ

1. mRNA: TruSeq RNA-Seq (Gold Standard)

- ~20,000 transcripts
 - More when consider splice variants / isoforms
- Observe 80-95% of transcripts depending on sequencing depth

2. Low input methods ~3000 cells / well

- 4000-6000 transcripts per sample
 - Limiting to transcripts observed across all samples
- Observe 20-60% of the transcriptome

3. Single Cell Methods

- 200 -10,000 transcripts per cell
- Observe 10-50% of the transcriptome
- Many transcripts will show up with zero counts in every cell. (even GAPDH)
- If you only looked at transcripts observed in all cells numbers drop dramatically.

BULK VS SINGLE CELL RNA-SEQ

	Deep RNA-seq	Sort-seq	Low input	scRNA-seq
Transcriptome Coverage	High	High	Moderate	Low
Throughput	Moderate	Low	High	Low
Cell Subtype Information	None	Moderate	None	High
Sequencing Depth	Moderate	Moderate	Low	High
Cost per Sample	Moderate	Moderate	Low	High

Disadvantages of scRNA-seq

- Dropouts and noisy data
- Lowly expressed genes might be undetected
- Samples will contain doublets
- Replicates without batch effect are unlikely
- Expensive

Source: Sarah Boswell, Harvard Medical School, September 2020

APPLICATIONS

naturemedicine

Letter | Published: 08 June 2020

A single-cell atlas of the peripheral immune response in patients with severe COVID-19

Aaron J. Wilk, Arjun Rustagi, Nancy Q. Zhao, Jonasel Roque, Giovanni J. Martínez-Colón, Julia L. McKechnie, Geoffrey T. Iverson, Thanmayi Ranganath, Rosemary Vergara,

LETTER

<https://doi.org/10.1038/s41596-018-0394-6>

A single-cell atlas of the airway epithelium reveals the CFTR-rich pulmonary ionocyte

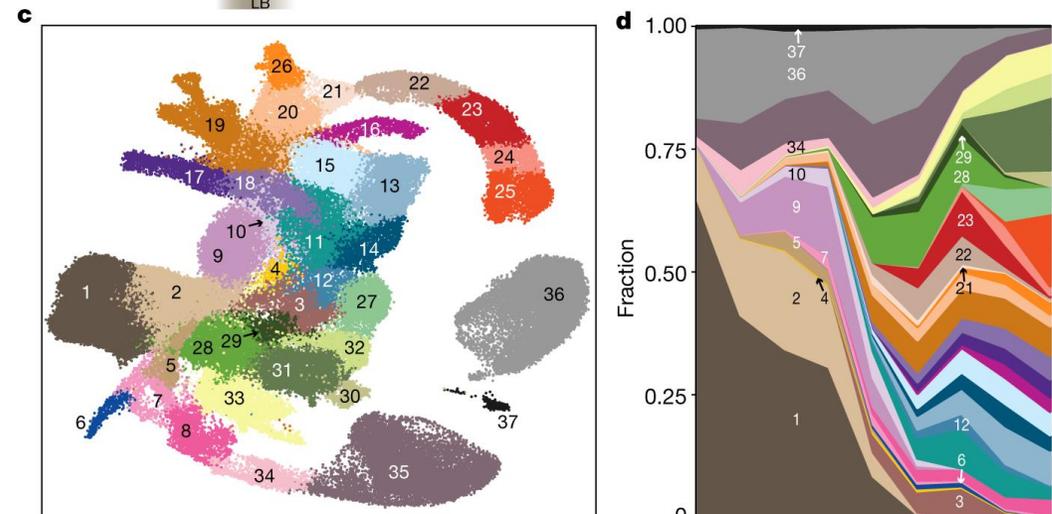
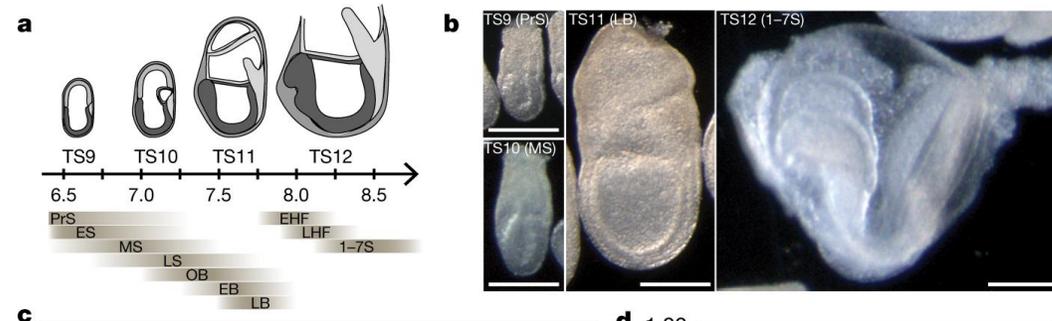
Lindsey W. Plasschaert^{1,3,7}, Rapolas Zilionis^{2,3,7}, Rayman Choo-Wing^{1,5}, Virginia Savova^{2,6}, Judith Knehr⁴, Guglielmo Romà⁴, Allon M. Klein^{2*} & Aron B. Jaffe^{1,5*}

nature

Article | Published: 20 February 2019

A single-cell molecular map of mouse gastrulation and early organogenesis

Blanca Pijuan-Sala, Jonathan A. Griffiths, Carolina Guibentif, Tom W. Hiscock, Wajid Jawaid, Fernando J. Calero-Nieto, Carla Mulas, Ximena Ibarra-Soria, Richard C. V.



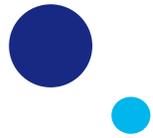
- 1 Epiblast
- 2 Primitive streak
- 3 Caudal epiblast
- 4 Primordial germ cells
- 5 Anterior primitive streak
- 6 Notochord
- 7 Def. endoderm
- 8 Gut
- 9 Nascent mesoderm
- 10 Mixed mesoderm
- 11 Intermediate mesoderm
- 12 Caudal mesoderm
- 13 Paraxial mesoderm
- 14 Somitic mesoderm
- 15 Pharyngeal mesoderm
- 16 Cardiomyocytes
- 17 Allantois
- 18 ExE mesoderm
- 19 Mesenchyme
- 20 Haemato-endothelial prog.
- 21 Blood progenitors 1
- 22 Blood progenitors 2
- 23 Erythroid 1
- 24 Erythroid 2
- 25 Erythroid 3
- 26 Endothelium
- 27 Neuromesodermal progenitors
- 28 Rostral neuroectoderm
- 29 Caudal neuroectoderm
- 30 Neural crest
- 31 Forebrain/midbrain/hindbrain
- 32 Spinal cord
- 33 Surface ectoderm
- 34 Visceral endoderm
- 35 ExE endoderm
- 36 ExE ectoderm
- 37 Parietal endoderm

Source: Pijuan-Sala et al. Nature 566, 490–495 (2019)



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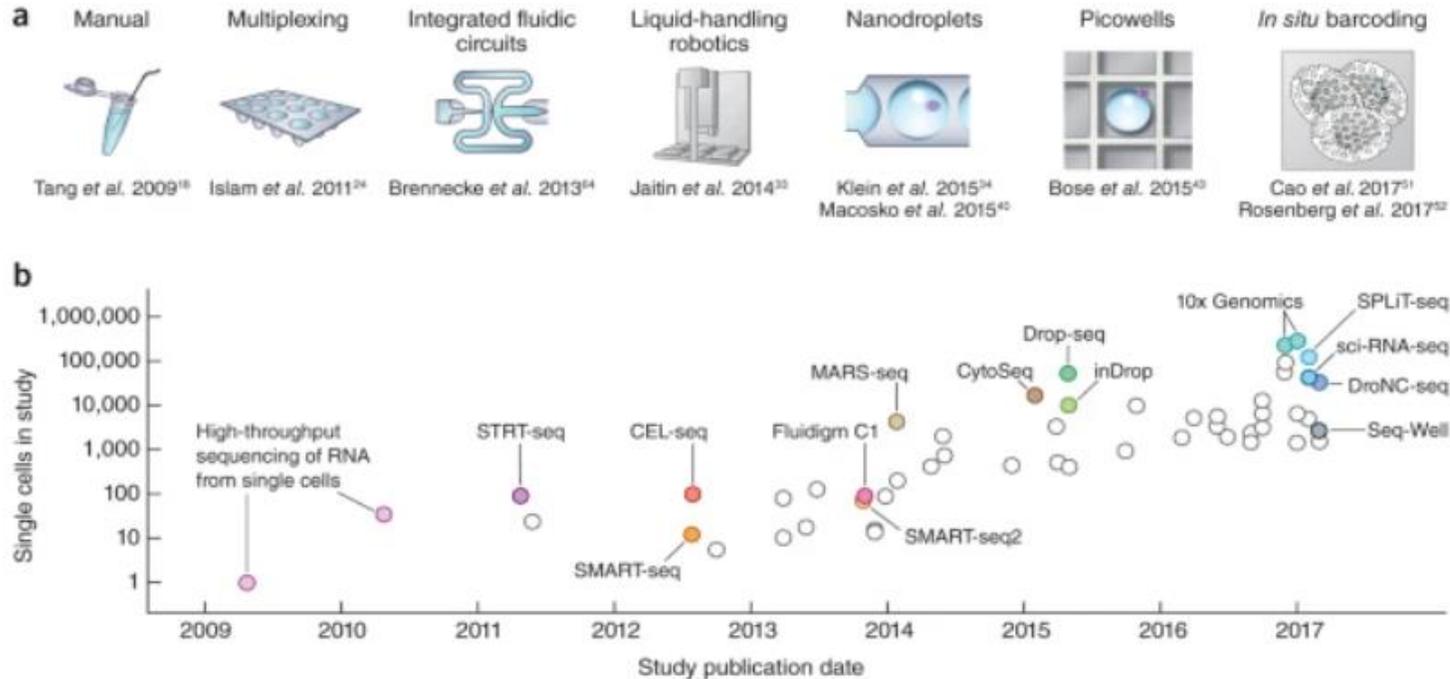
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TECHNOLOGIES



Figure 1: Scaling of scRNA-seq experiments.



Source: Svensson et al. *Nat Protoc* 13, 599–604 (2018)



Parse WT Penta –
5mln cells in one
experiment

Chromium X



Making 1 million cell experiments routine



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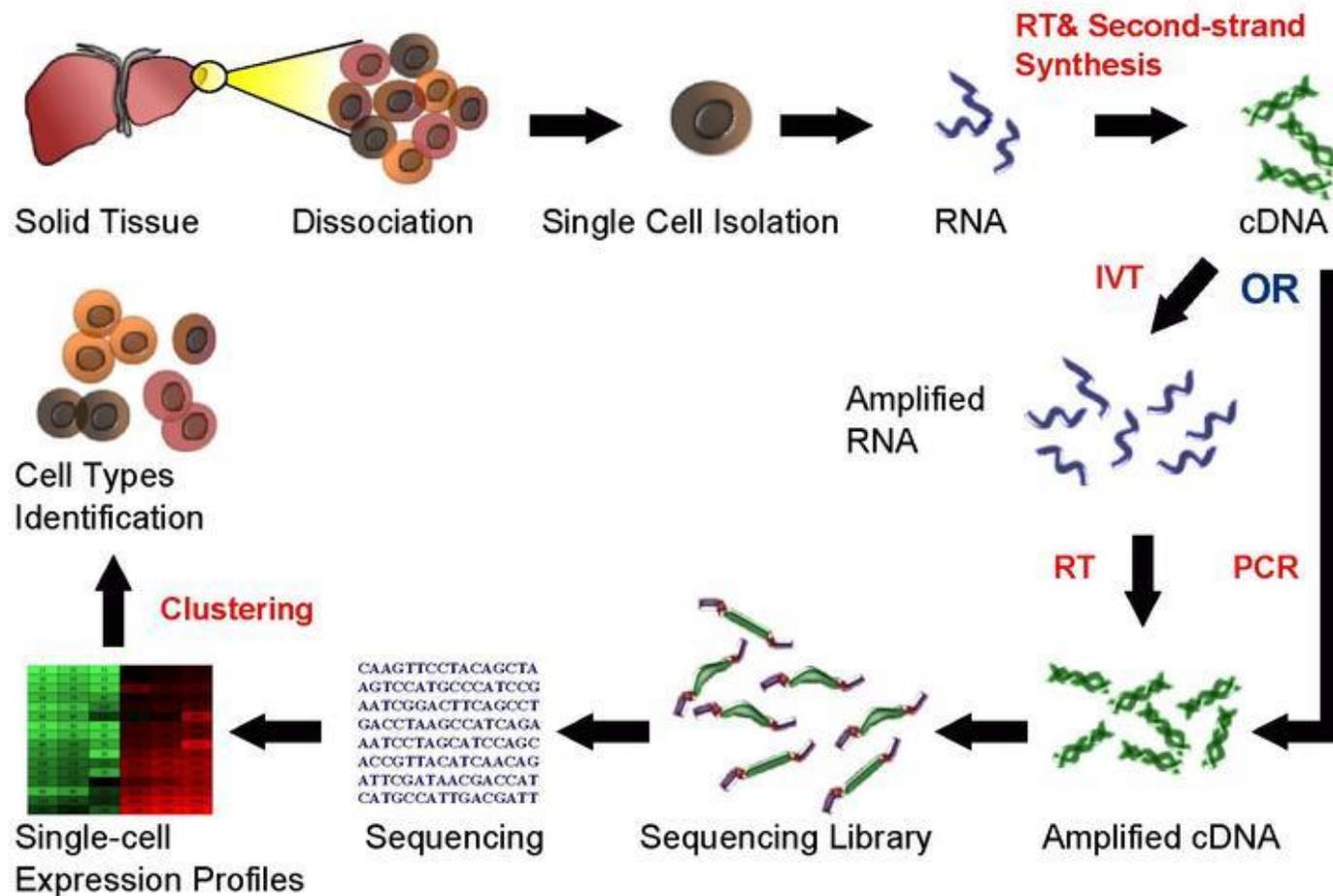
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WORKFLOW

Single Cell RNA Sequencing Workflow

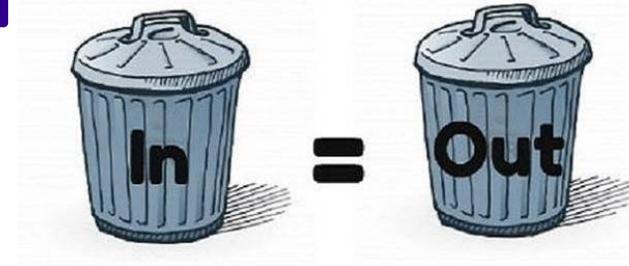


Good sample preparation is key to success!



Source: https://en.wikipedia.org/wiki/Single_cell_sequencing

SAMPLE PREPARATION



- Understand well the nature of the sample (sampling conditions, preparation, purity)
- Identify the source of technical difficulties in order to resolve them first
- Practice your sample preparation, optimise the protocol well, do not rush to the final experiment
- A well planned pilot experiment is essential for evaluating sample preparation and for understanding the required number of cells.
- You need your cells to be highly viable (>90-95%), have no clumps and no debris. Cell membrane integrity is a must!
- Free-floating RNA will make analysis more challenging
- Be cautious about FACS (especially with more fragile cells). If FACS necessary for enrichment, remember that time is crucial factor
- Count with haemocytometer or cell counter (Countess II Automated Cell Counter) – do not trust sorter counts
- Fixation and cryopreservation are not compatible with many techniques



METHODS

1) Cells in wells, traps and valves (nanowell, Flow sorting, CellenOne, Fluidigm C1, SmartSeq, plexWell/seqWell)

- Screen for and retrieve single cells of interest
- Enrich for rare cells with decided properties
- Control the cellular microenvironment
- Monitor and control cell-cell interactions
- Precise/extensive manipulation of single cells



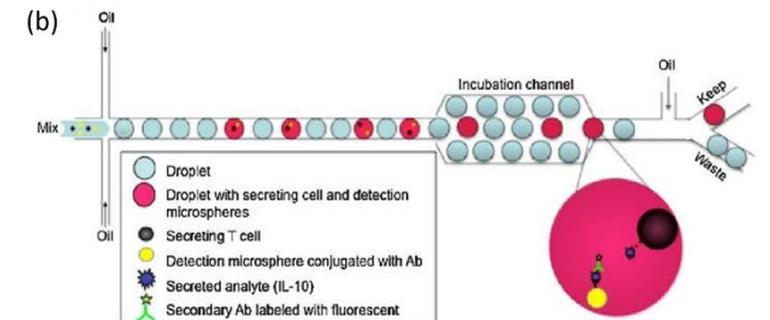
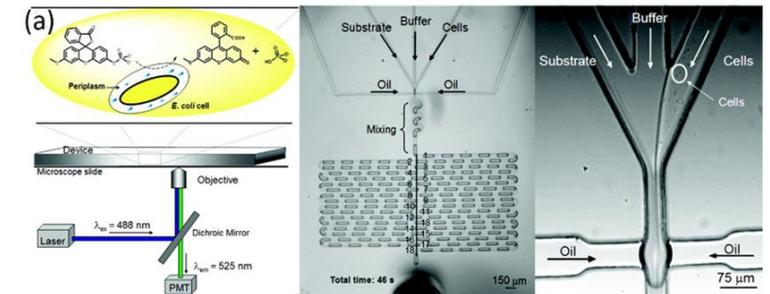
Passive wells



Active pumps and valves

2) Droplets (Drop-seq, 10x Genomics)

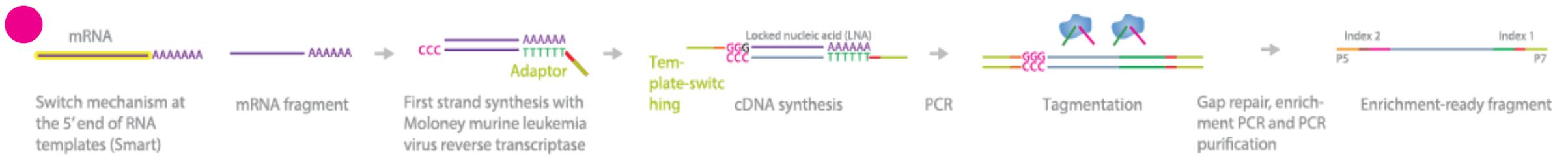
- Introduce distinct 'packets' of reagents to single cell (e.g. barcodes)
- Perform amplification on individual cells
- Sort large population of single cells



3) Combinatorial indexing (SCI-seq, SPLiT-seq)

- Economic use of reagents for cell separation
- Efficiency of handling larger population than Drop-seq
- Maintain complexities of population without bias from droplet or well

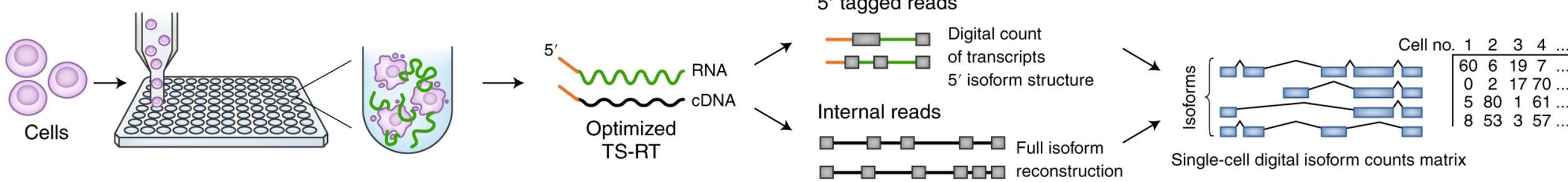
SMART-SEQ2/3/4 OVERVIEW



Developed for single cell but can be performed using total RNA.

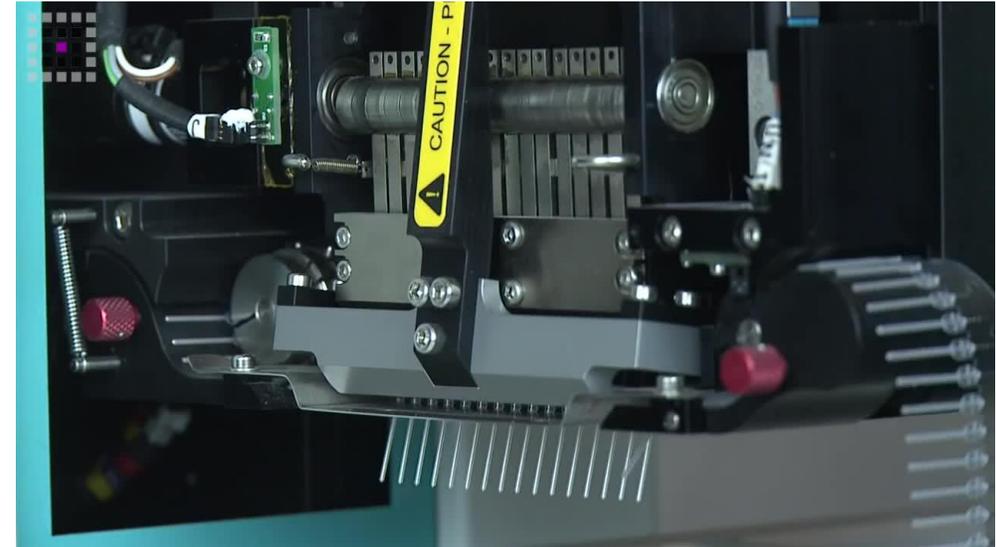
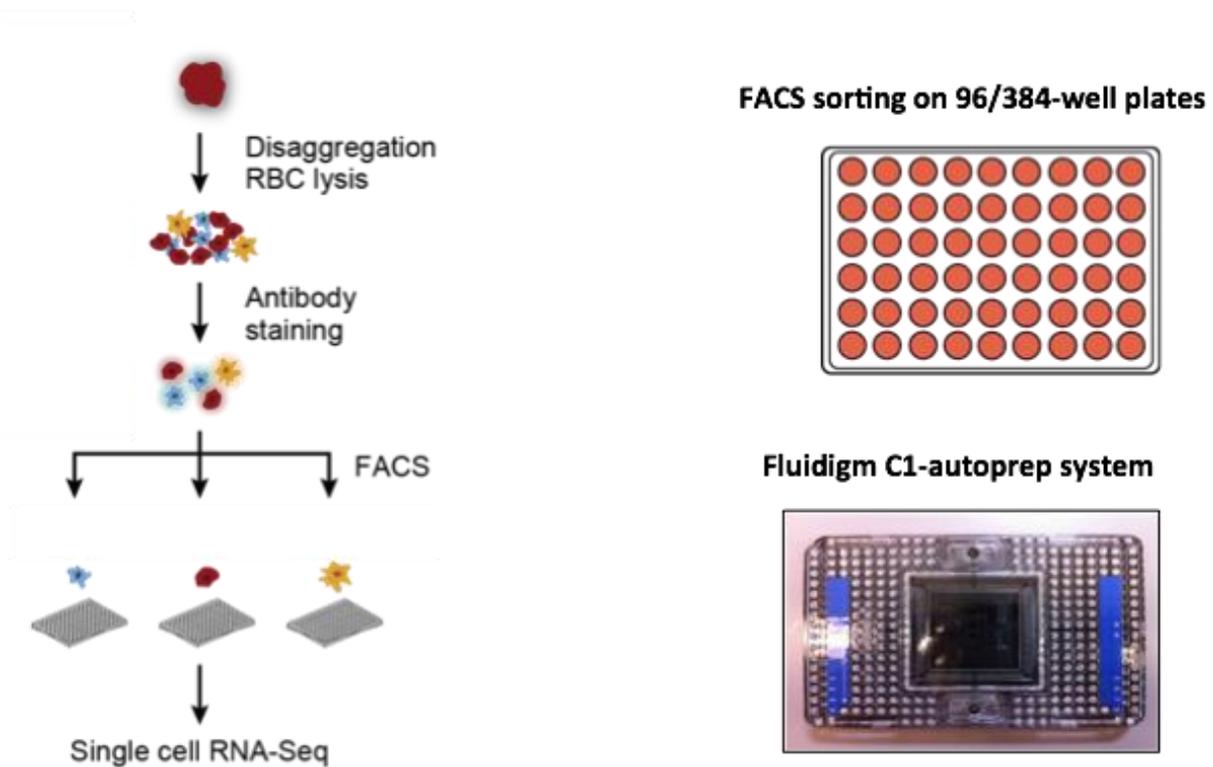
- Selects for poly-A tail.
- Full transcript assay.
- Uses template switching for 5' end capture.
- Standard Illumina sequencing.
- Plate-based solution so labour intensive, slow and costly (~\$12/cell)

a

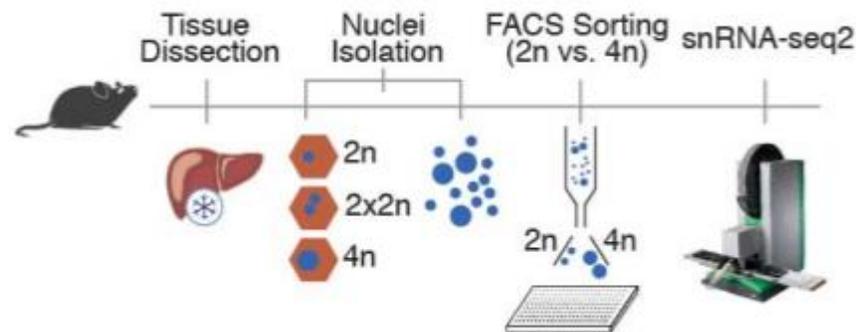


Source: Macosko, Nat Biotechnol 38 (2020).

SMART-SEQ2/3/4 + MOSQUITO LV



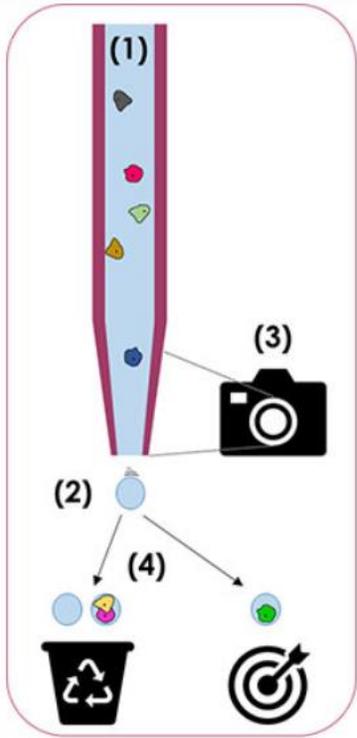
Source: SPT Labtech



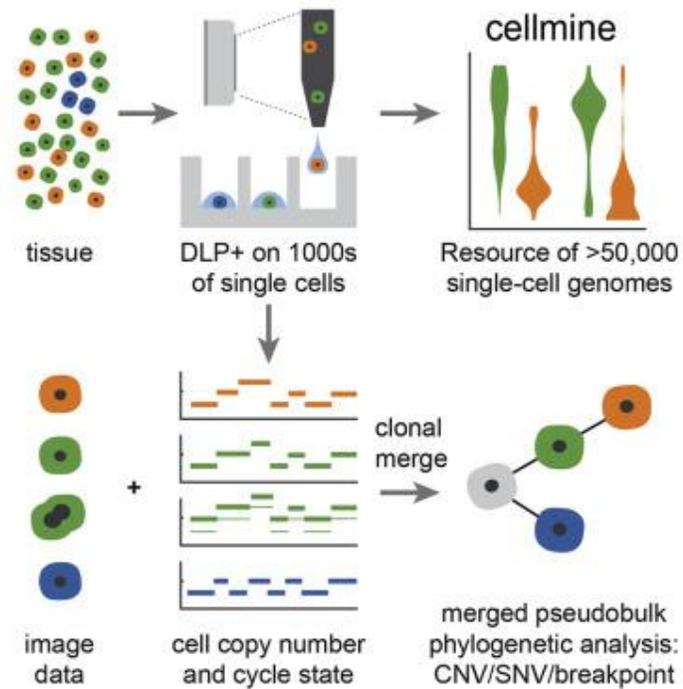
- Mosquito LV makes assay miniaturisation simple, leading to significant savings on precious reagents and time.
- Mosquito LV offers highly accurate and precise multichannel pipetting from 25 nL to 1.2 μ L.
- SmartSeq2 cost reduced from \$12 to \$4 per cell

CELLENONE

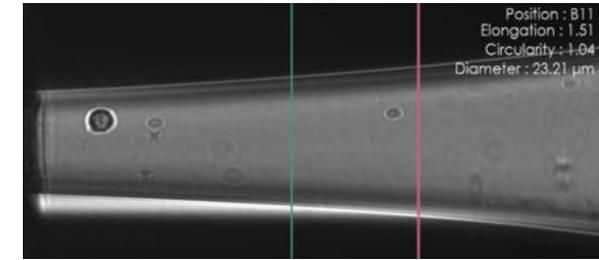
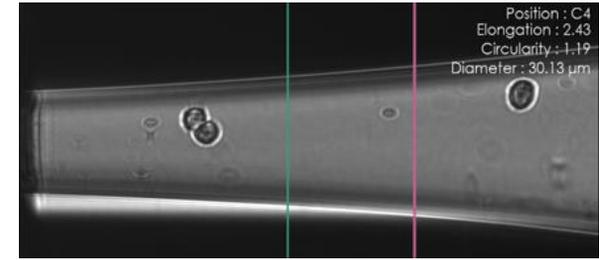
- 1) Cell suspension is aspirated into a glass capillary
- 2) Generation of drops on demand, in air
- 3) Thanks to automated imaging, cellenONE tracks cells and determines if upcoming drops will contain or not a single cell
- 4) Drops containing single cells are dispensed into selected targets, drops without cells or with more than one cells are dispensed into recycling tube



Source: Cellenion



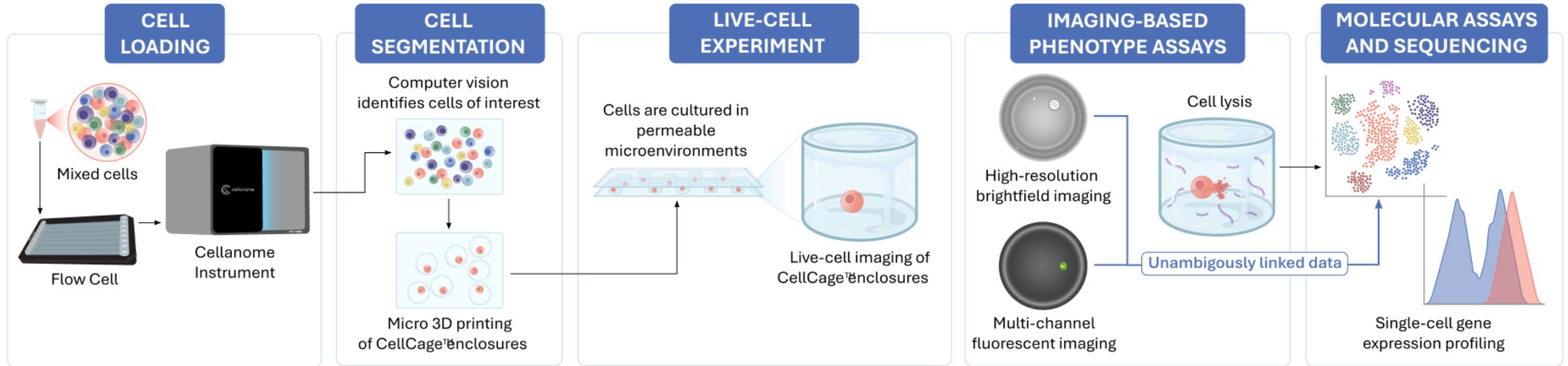
Source: Laks et al. *Cell*. 179(5):1207-1221.e22. (2019)



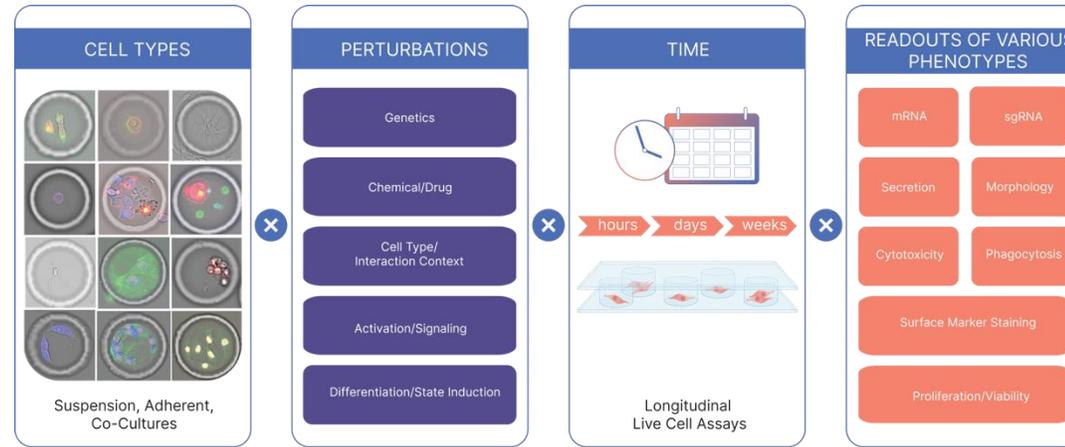
Source: Cellenion

CELLANOME

The Cellanome Platform reveal cellular behavior, interactions and molecular programs in ways that static snapshots and fragmented approaches cannot.



Multi-modal data integration and analysis



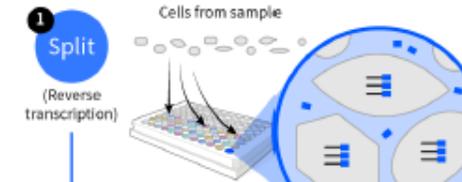
SPLIT-SEQ OVERVIEW (PARSE BIOSCIENCES, SCALE BIO)



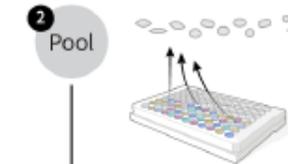
Video available at:
<https://www.youtube.com/watch?v=WqaeZe7mKUc>

- Time flexibility – single experiment for samples collected on different dates (up to 6months storage)
- No instrument required for experiment. Computational pipeline available
- Up to 48 samples / 100k cells or 96 samples / 1M cells – kit has to be used at once
- Doublet rate of 0.27% per 1000 cells (3.4% per library)
- Random hexamers method on top of PolyA capture
- Works with any species, any sizes of cells/nuclei & results in lower background noise
- CRISPR, TCR, BCR profiling compatibility

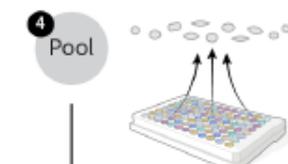
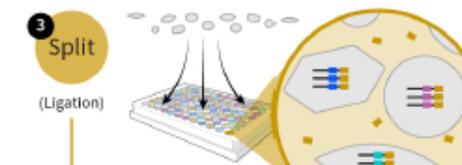
In the first round of barcoding, fixed cell samples are distributed into 48 wells, and cDNA is generated with an in-cell reverse transcription (RT) reaction using well-specific barcoded primers.



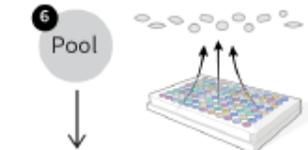
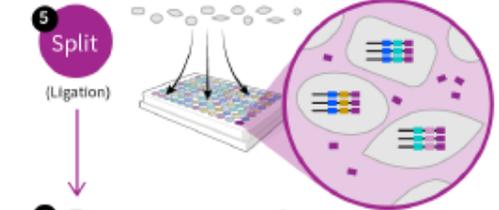
Cells from each well are pooled back together.



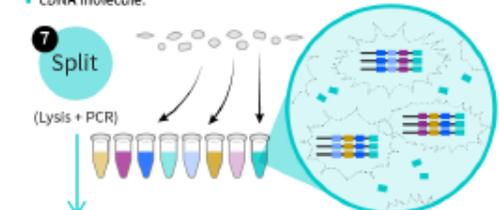
Cells are then distributed into 96 wells, and an in-cell ligation reaction appends a second well-specific barcode to the cDNA.



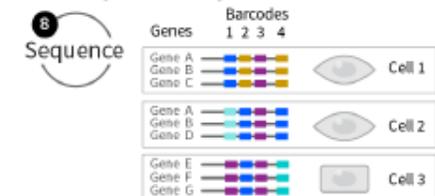
The third-round barcode is appended with another round of in-cell ligation.



After three rounds of barcoding, the cells are pooled and split into 8 distinct populations we term sublibraries. The user can choose the number of cells in each sublibrary to control the depth of sequencing. Cells will not be pooled again after this step. After this final split cells are lysed and the barcoded cDNA is isolated. A fourth sublibrary-specific barcode is introduced by PCR to each cDNA molecule.

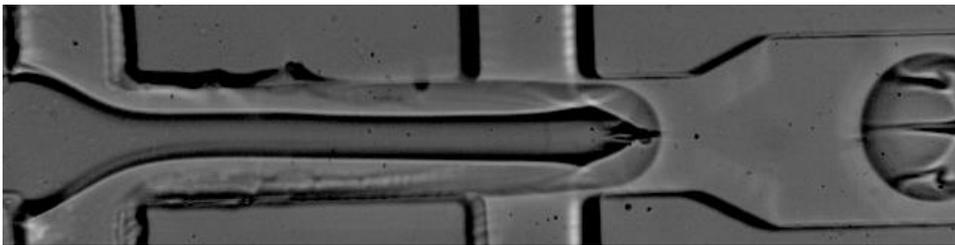


After sequencing, each single cell transcriptome is assembled by combining reads containing the same four-barcode combination.



DROP-SEQ OVERVIEW

- Moved throughput from hundreds to thousands.
- Droplet-based processing using microfluidics
- Nanoliter scale aqueous drops in oil.
- 3' End
- Bead based (STAMPs).
- Single-cell transcriptomes attached to microparticles.
- Cell barcodes use split-pool synthesis.
- Uses UMI (Unique Molecular Identifier)
- Chance to have two cells within one droplet

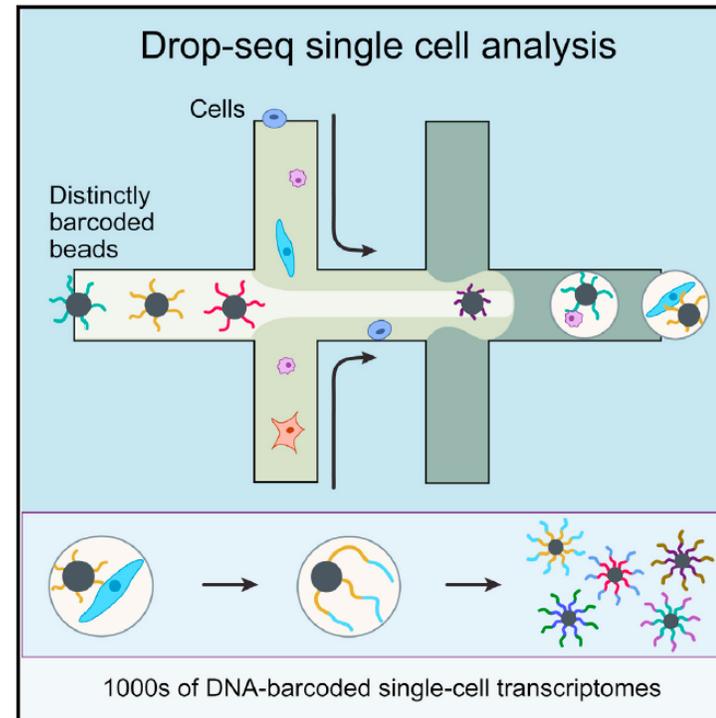


Resource

Cell

Highly Parallel Genome-wide Expression Profiling of Individual Cells Using Nanoliter Droplets

Graphical Abstract



Authors

Evan Z. Macosko, Anindita Basu, ...,
Aviv Regev, Steven A. McCarroll

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(E.Z.M.),
mccarroll@genetics.med.harvard.edu
(S.A.M.)

In Brief

Capturing single cells along with sets of uniquely barcoded primer beads together in tiny droplets enables large-scale, highly parallel single-cell transcriptomics. Applying this analysis to cells in mouse retinal tissue revealed transcriptionally distinct cell populations along with molecular markers of each type.

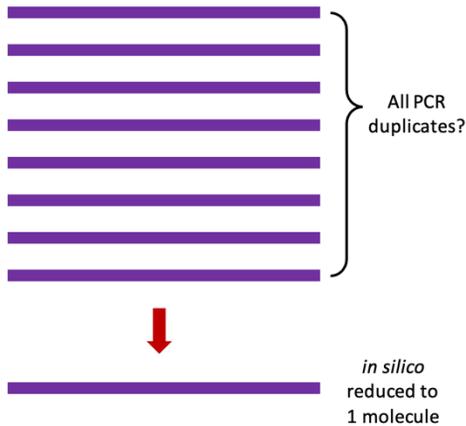
UMI – UNIQUE MOLECULAR IDENTIFIERS

After PCR enrichment, without UMIs, one can not distinguish if multiple copies of a fragment are caused by PCR clones or if they are real biological duplicated.

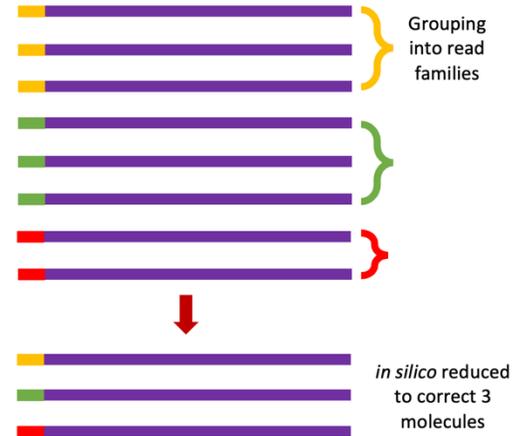
By using UMIs, PCR clones can be found by searching for non-unique fragment-UMI combinations, which can only be explained by PCR clones.

When performing variant analyses, these falsely overrepresented fragments can result in incorrect calls and thus wrong diagnostic findings

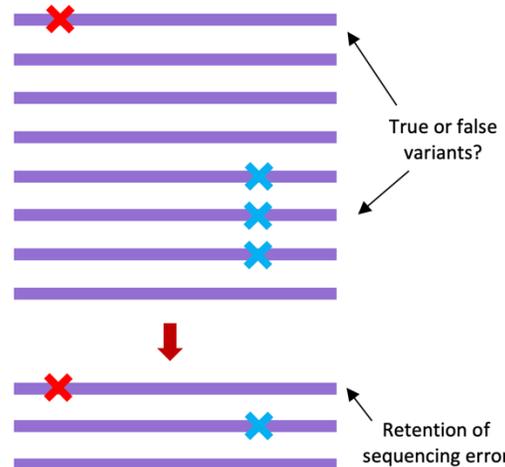
PCR duplicate removal **without** UMIs



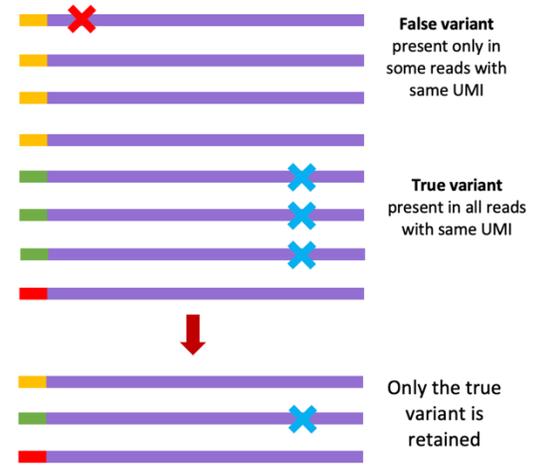
PCR duplicate removal **with** UMIs



Variant calling **without** UMIs

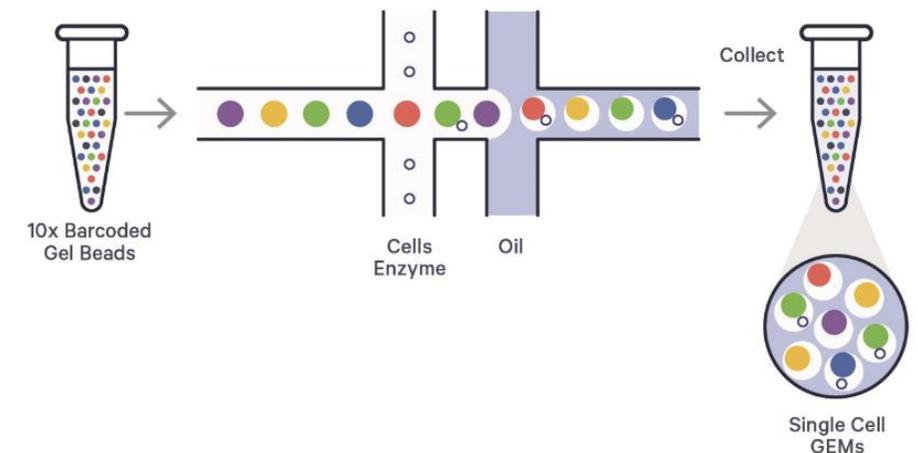
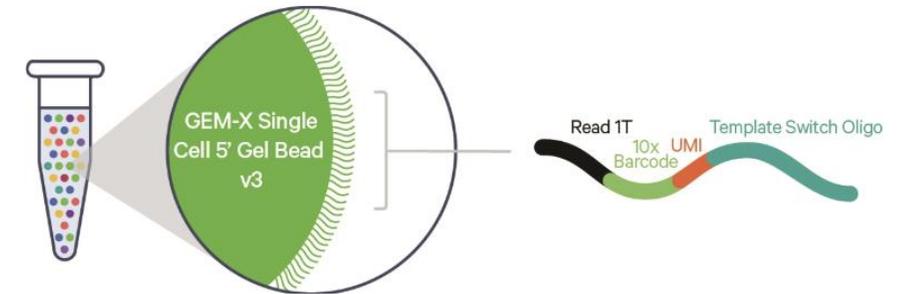
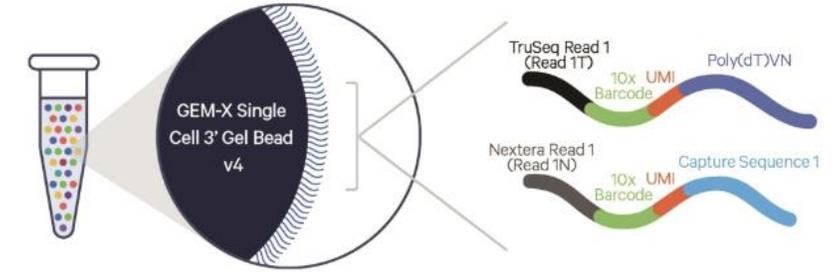


Variant calling **with** UMIs



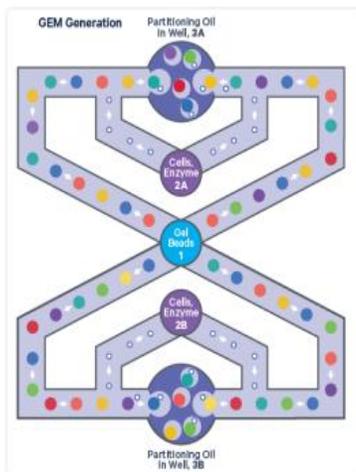
10X GENOMICS OVERVIEW

- Droplet-based similar to Drop-Seq, 3' or 5' mRNA
- In contrast to Drop-seq, where solid beads are used for RNA capture, 10X uses soft hydrogels containing oligos. These enable “single Poisson loading” leading to capture of >60% of input cells.
- Standardized instrumentation and reagents (unhackable so no customisation or control)
- Very easy to use and less processing time
- More high-throughput scaling - 16 samples can be processed simultaneously with up to 20000 cells captured per sample
- The doublet rate increases with number of cells loaded
- CellRanger and CellLoupe software are available and user friendly
- A lot of additional addons possible (CRISPR, TCR & BCR profiling etc.)



Source: 10x Genomics

10X GENOMICS OVERVIEW

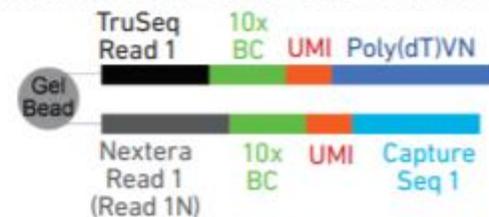


Chromium X



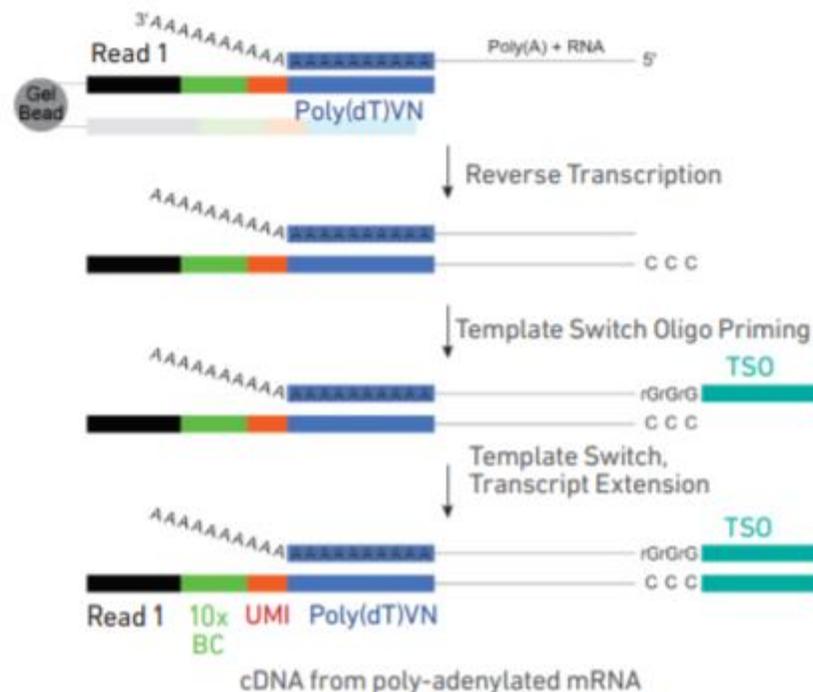
Making 1 million cell experiments routine

Inside individual GEMs



+
Master Mix
+
Cell Lysate

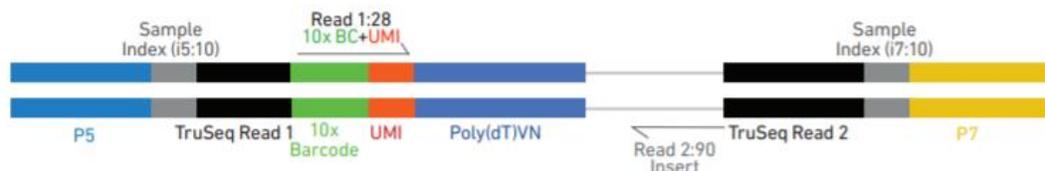
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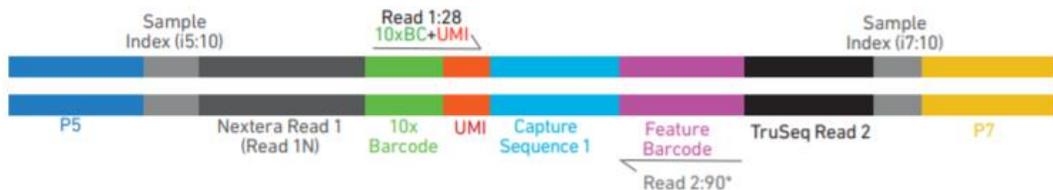
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10X GENOMICS LIBRARIES

Chromium Single Cell 3' Gene Expression Dual Index Library



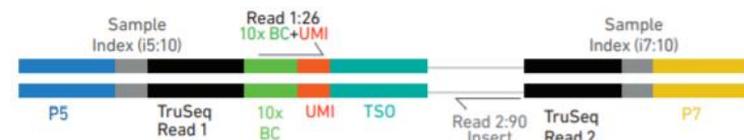
Chromium Single Cell 3' Cell Surface Protein Dual Index Library



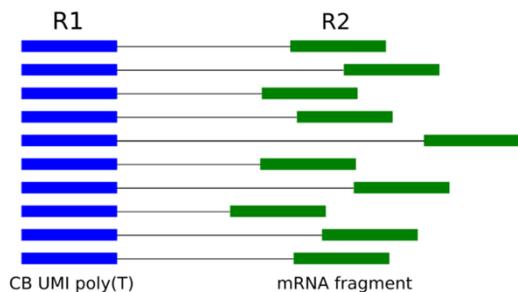
Chromium Single Cell V(D)J Dual Index Library



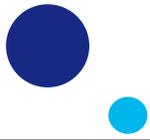
Chromium Single Cell 5' Gene Expression Dual Index Library



Source: 10x Genomics

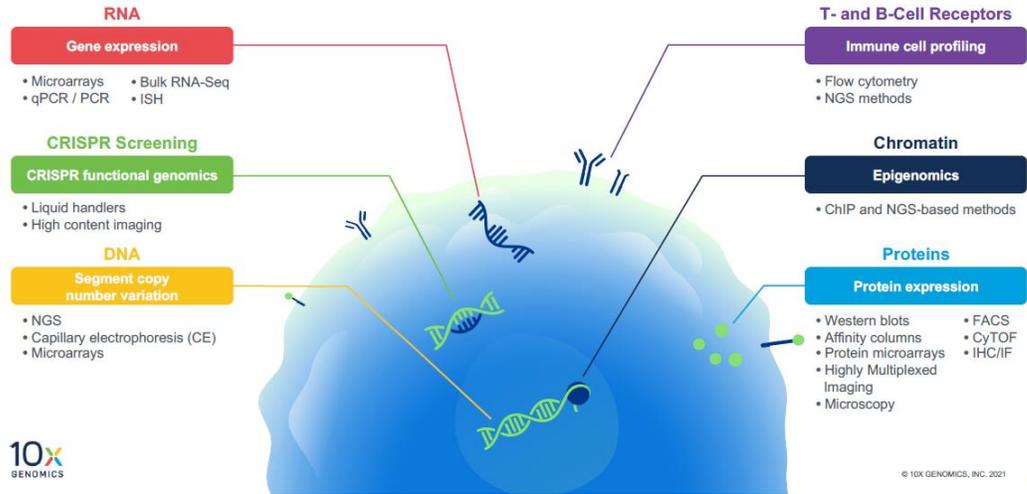


Sequencing Read	Description	Number of cycles
Read1	10x Barcode Read (Cell) + Randomer Read (UMI)	28bp
i7 index	Sample index read	10bp
i5 index	Sample index read	10bp
Read2	Insert Read (Transcript)	90bp



MULTIOMICS AGE

Replacing the Legacy Toolkit Across Biology



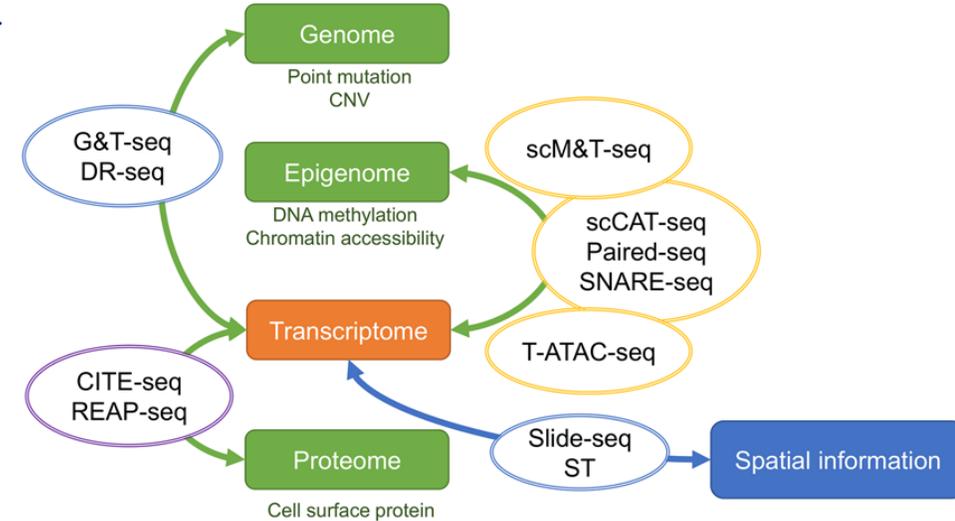
Source: 10x Genomics

TECHNOLOGY FEATURE | 19 July 2021 | Correction 21 July 2021

Single-cell analysis enters the multiomics age

A rapidly growing collection of software tools is helping researchers to analyse multiple huge ‘-omics’ data sets.

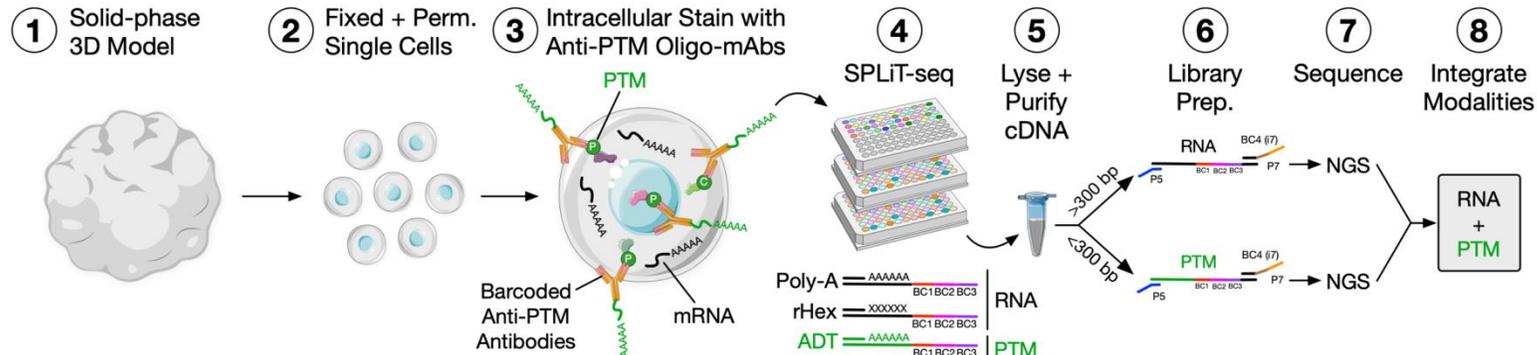
Jeffrey M. Perkel



Kashima Y et al. Exp Mol Med 52, 1419–1427 (2020)

SIGNAL-seq: a multiplexed split-pool combinatorial barcoding method that simultaneously measures RNA and post-translational modifications (PTMs) in fixed single cells.

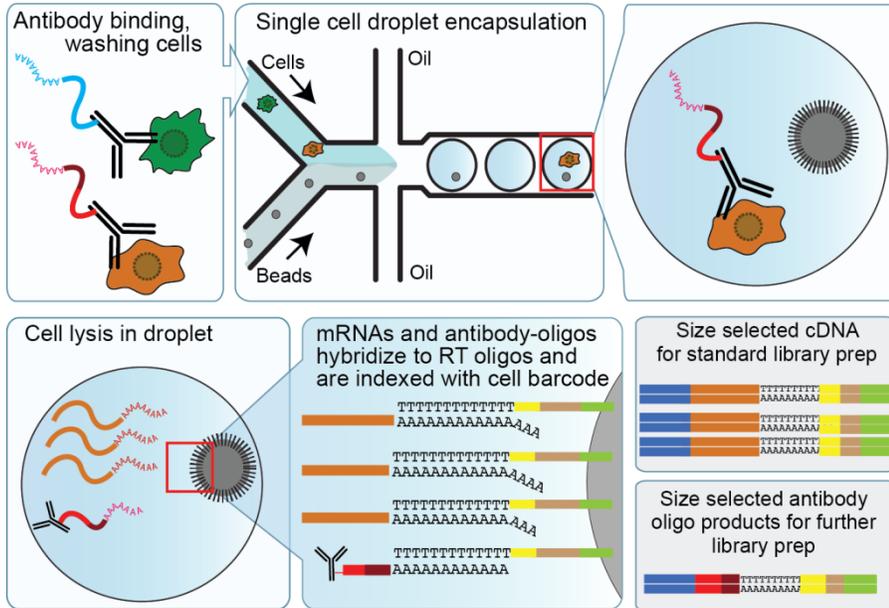
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CITE-SEQ

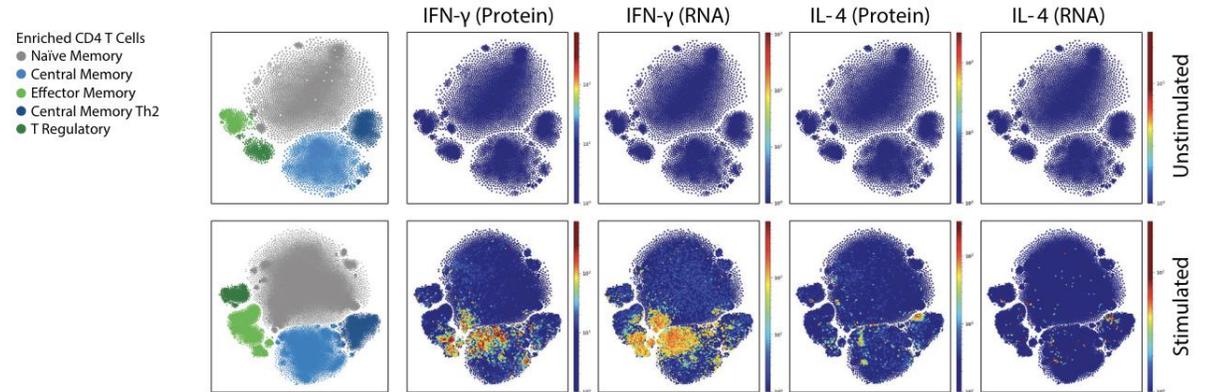
- Cellular Indexing of Transcriptomes and Epitopes by Sequencing
- CITE-seq uses DNA-barcoded antibodies to convert detection of proteins into a quantitative, sequenceable readout



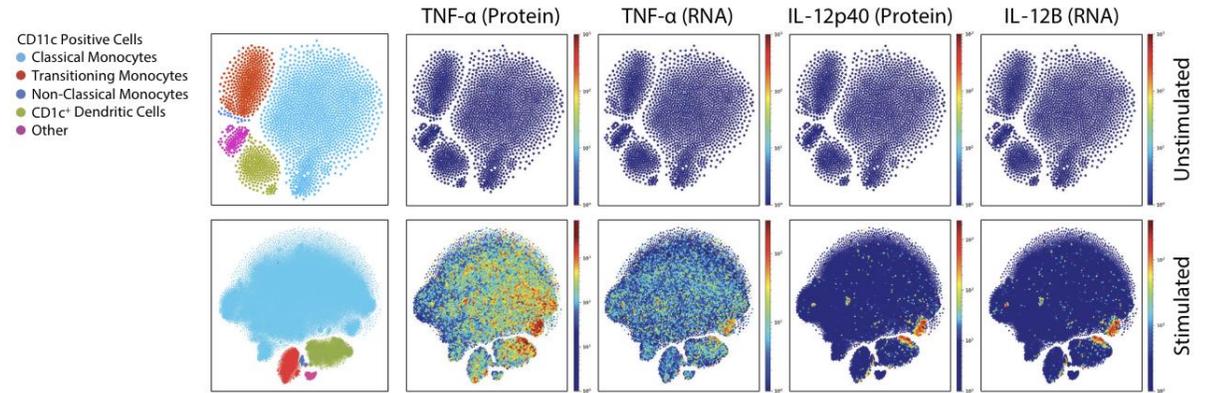
Source: cite-seq.com

Characterize RNA, Surface, and Intracellular Proteins in the Same Cell

Our TotalSeq-B conjugates offer a wide selection of targets against important secreted and intracellular proteins and complement your single-cell multiomics experiment for comprehensive cellular characterization.



CD4⁺ T cells were magnetically enriched from PBMCs and stimulated with PMA and Ionomycin for six hours in the presence of BFA.



CD14⁺ cells were magnetically enriched from PBMCs and stimulated with GM-CSF overnight in the presence of LPS and R848, and for the final six hours with BFA.

Source: BioLegend, TotalSeq-Intracellular-Targets-Info-Sheet



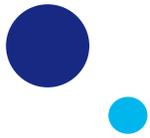
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BioLegend solutions:

TotalSeq-A – Poly(dT) based system

TotalSeq-B – 3' v3.1 Feature barcode

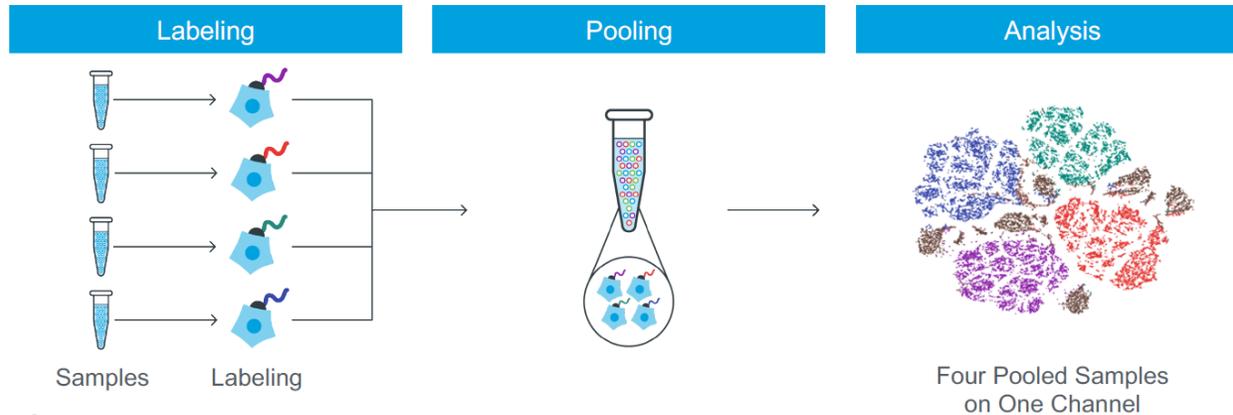
TotalSeq-C – 5' v2.0



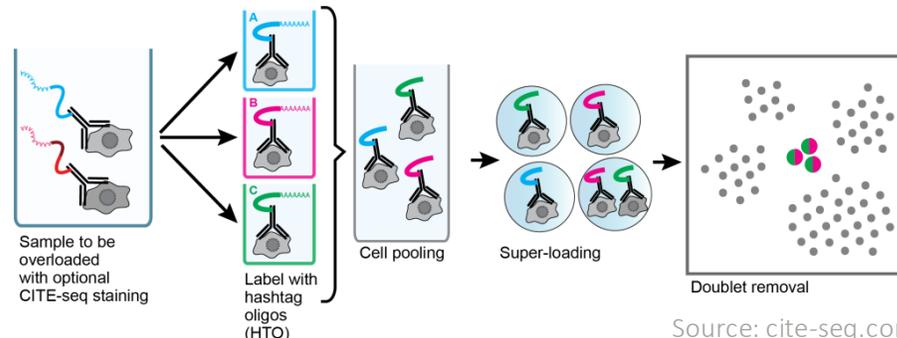
CELL HASHING



Reduces cost of running multiple samples by adding hashtag oligos and pooling into single channel of 10x chip (10x CellPlex or TotalSeq antibodies)



Allows overloading as by sequencing tags alongside the cellular transcriptome, we can assign each cell to its sample of origin, and robustly identify doublets originating from multiple samples



Genotype-free demultiplexing of pooled single-cell RNA-Seq

Jun Xu^a, Caitlin Falconer^b, Quan Nguyen^b, Joanna Crawford^b, Brett D. McKinnon^{b,e}, Sally Mortlock^b, Alice Pébay^{f,g,h,i}, Alex W. Hewitt^{f,g,h,i}, Anne Senabouth^d, Nathan Palpant^{a,b}, Han Chiu^b, Stacey Andersen^{a,b}, Grant W. Montgomery^{a,b}, Joseph Powell^{c,d}, Lachlan Coin^{a,b,*}



Article | Published: 17 June 2019

MULTI-seq: sample multiplexing for single-cell RNA sequencing using lipid-tagged indices

Christopher S. McGinnis, David M. Patterson, Juliane Winkler, Daniel N. Conrad, Marco Y. Hein, Vasudha Srivastava, Jennifer L. Hu, Lyndsay M. Murrow, Jonathan S. Weissman, Zena Werb, Eric D. Chow & Zev J. Gartner

Nature Methods 16, 619–626(2019) | [Cite this article](#)

15k Accesses | 27 Citations | 85 Altmetric | [Metrics](#)



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TISSUE PRESERVATION/CELLS FIXATION

Research | [Open Access](#) | Published: 02 June 2020

Systematic assessment of tissue dissociation and storage biases in single-cell and single-nucleus RNA-seq workflows

[Elena Denisenko](#), [Belinda B. Guo](#), [Matthew Jones](#), [Rui Hou](#), [Leanne de Kock](#), [Timo Lassmann](#), [Daniel Poppe](#), [Olivier Clément](#), [Rebecca K. Simmons](#), [Ryan Lister](#) & [Alistair R. R. Forrest](#) ✉

Genome Biology 21, Article number: 130 (2020) | [Cite this article](#)

14k Accesses | 39 Citations | 40 Altmetric | [Metrics](#)

Genome Biol. 2020; 21: 1.

Published online 2019 Dec 31. doi: [10.1186/s13059-019-1906-x](https://doi.org/10.1186/s13059-019-1906-x)

PMCID: PMC6937944

PMID: [31892341](https://pubmed.ncbi.nlm.nih.gov/31892341/)

scRNA-seq assessment of the human lung, spleen, and esophagus tissue stability after cold preservation

[E. Madisson](#),^{#1,2} [A. Wilbrey-Clark](#),^{#1} [R. J. Miragaia](#),¹ [K. Saeb-Parsy](#),³ [K. T. Mahbubani](#),³ [N. Georgakopoulos](#),³ [P. Harding](#),¹ [K. Polanski](#),¹ [N. Huang](#),¹ [K. Nowicki-Osuch](#),⁴ [R. C. Fitzgerald](#),⁴ [K. W. Loudon](#),⁵ [J. R. Ferdinand](#),⁵ [M. R. Clatworthy](#),⁵ [A. Tsingene](#),¹ [S. van Dongen](#),¹ [M. Dabrowska](#),¹ [M. Patel](#),¹ [M. J. T. Stubbington](#),^{1,6} [S. A. Teichmann](#),¹ [O. Stegle](#),² and [K. B. Meyer](#)^{#1}

Research | [Open Access](#) | Published: 10 May 2021

Cryopreservation of human cancers conserves tumour heterogeneity for single-cell multi-omics analysis

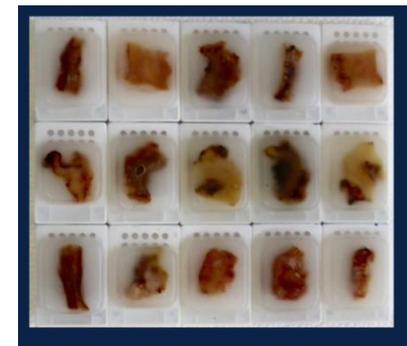
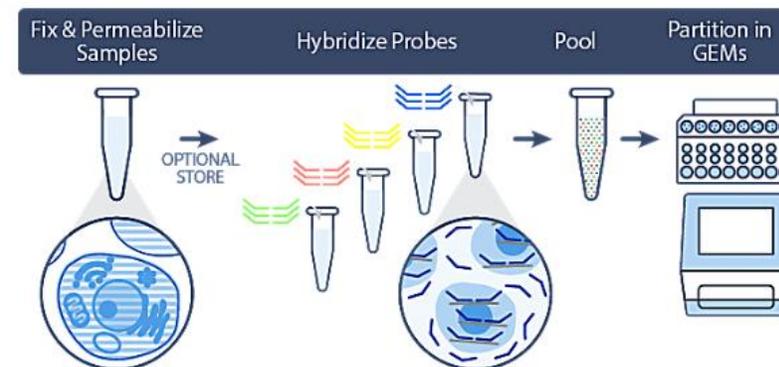
[Sunny Z. Wu](#), [Daniel L. Roden](#), [...] [Alexander Swarbrick](#) ✉

Genome Medicine 13, Article number: 81 (2021) | [Cite this article](#)

1757 Accesses | 34 Altmetric | [Metrics](#)

10x Fixed RNA Profiling

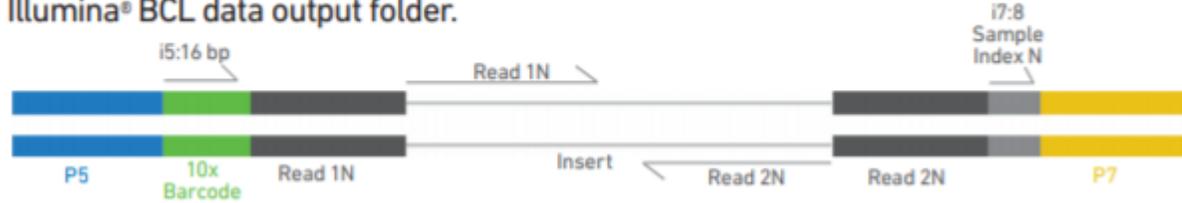
- new chemistry, compatible with formaldehyde fixed samples
- RNA is captured using probes, not poly(d)T like in 3' solution
- Available for human (~18k genes) and mouse (~20k genes) only
- Probes contain barcodes so no additional staining needed for cell hashing
- Kit potentially opens the door to archival material (FFPE blocks)
- pre-print: snPATHO-seq: unlocking the FFPE archives for single nucleus RNA profiling



SplitSeq (Parse Bioscience) requires fixation as well

10X ATAC

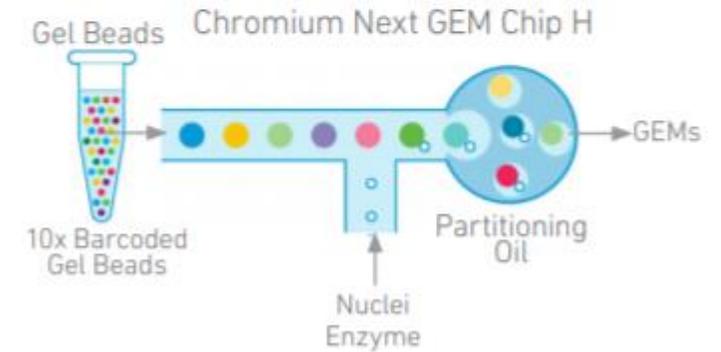
Chromium Single Cell ATAC libraries comprise double stranded DNA fragments which begin with P5 and end with P7. Sequencing these libraries produces a standard Illumina® BCL data output folder.



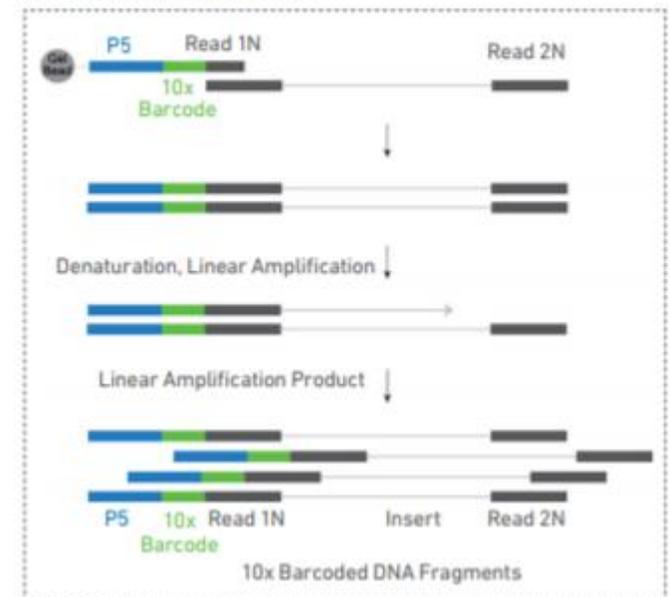
Sequencing Read	Description	Number of cycles
Read1	Insert Sequence 1N	50bp
i7 index	Sample index read	8bp
i5 index	10x Barcode Read (Cell)	16bp
Read2	Insert Sequence 2N (opposite end)	50bp

- ASAP-seq is to scATAC-seq what CITE-seq is to scRNA-seq.

- Scale Biosciences – ‘pre-indexing of nuclei through tagmentation’ = 100k nuclei per 10x channel with low number of doublets



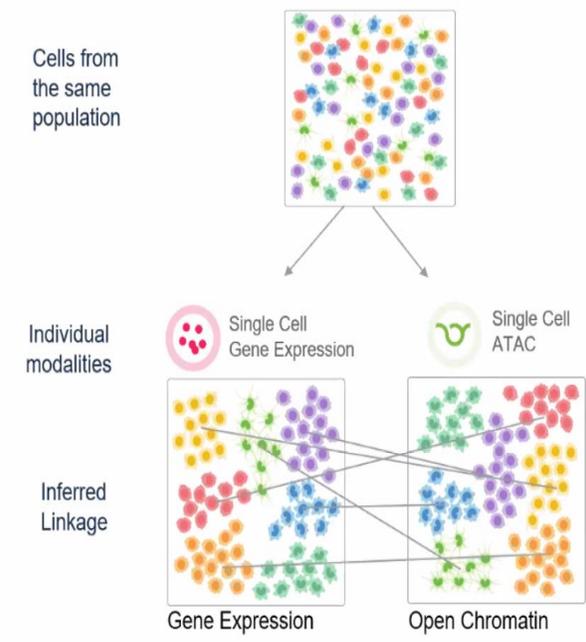
Inside Individual GEMs



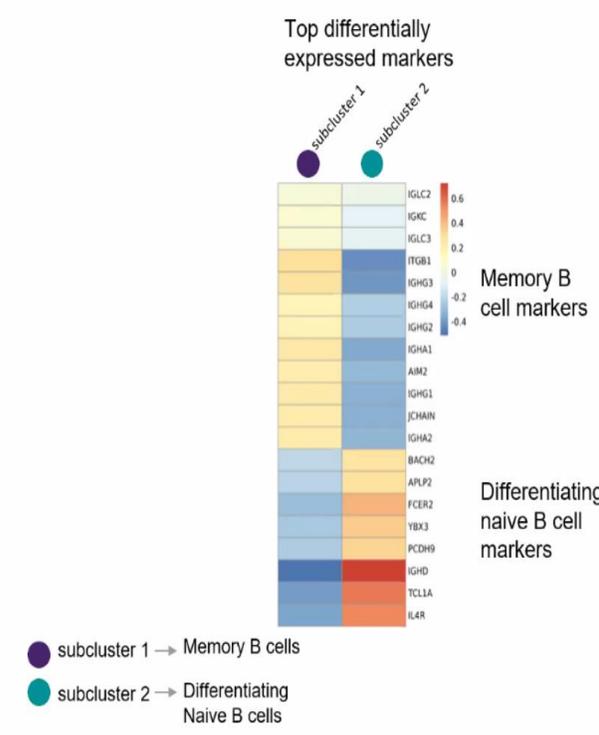
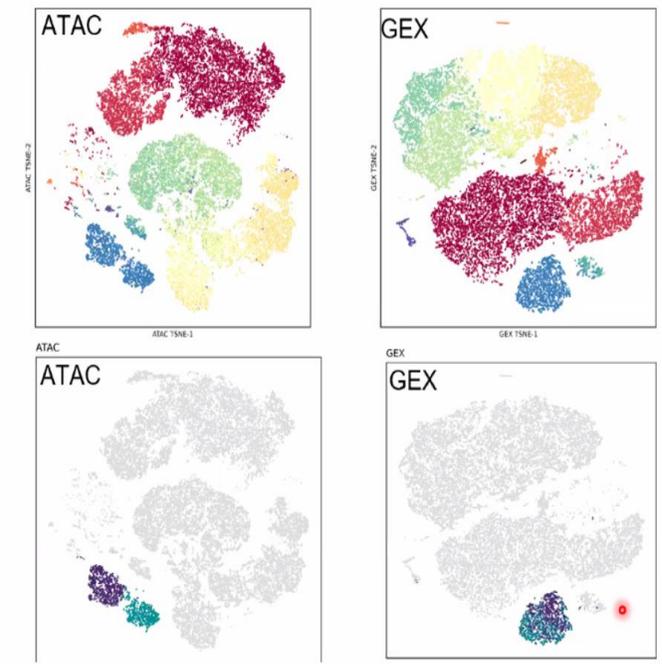
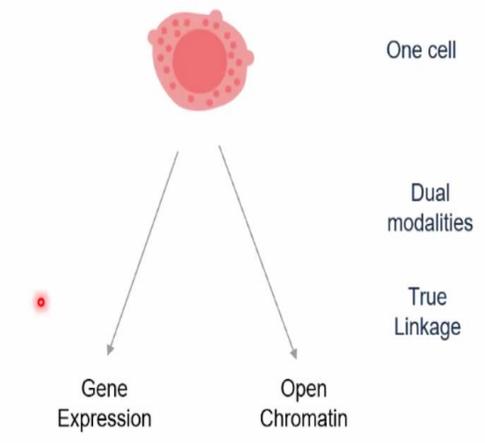
Source: 10x Genomics

10X MULTIOME (RNA+ATAC)

Profiling Different Modalities To Gain Deeper Insights



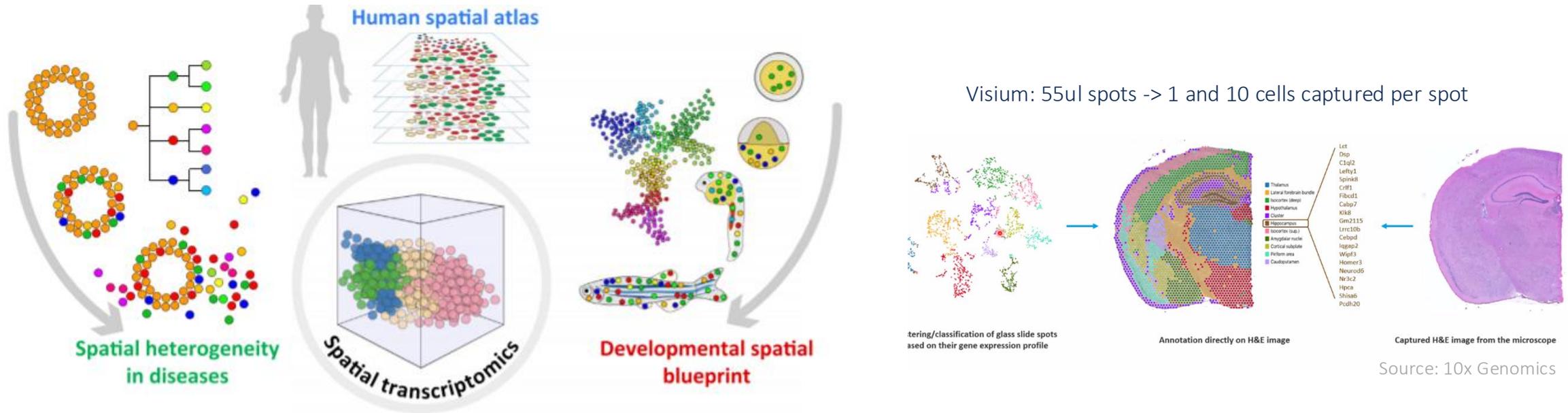
Dive Deep Where It Matters



Source: 10x Genomics

-TEA-seq (Transcription, Epitopes, and Accessibility) = Multiome with permabilised cells & CITEseq

SPATIAL TRANSCRIPTOMICS



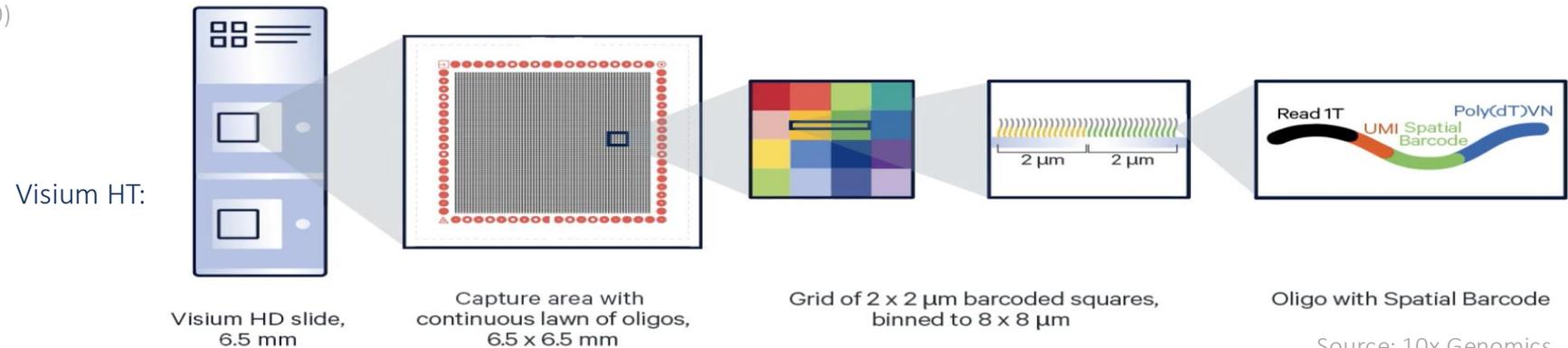
Visium: 55ul spots -> 1 and 10 cells captured per spot

Source: 10x Genomics

Trends in Biotechnology

Figure 3. Applications for Spatially Resolved Transcriptomics. Three primary kinds of hot issues can be resolved by spatially resolved transcriptomics: left, discovering spatial heterogeneity of diseases; middle, establishing spatial transcriptome atlases for the human body; and right, delineating an embryonic developmental and spatial blueprint.

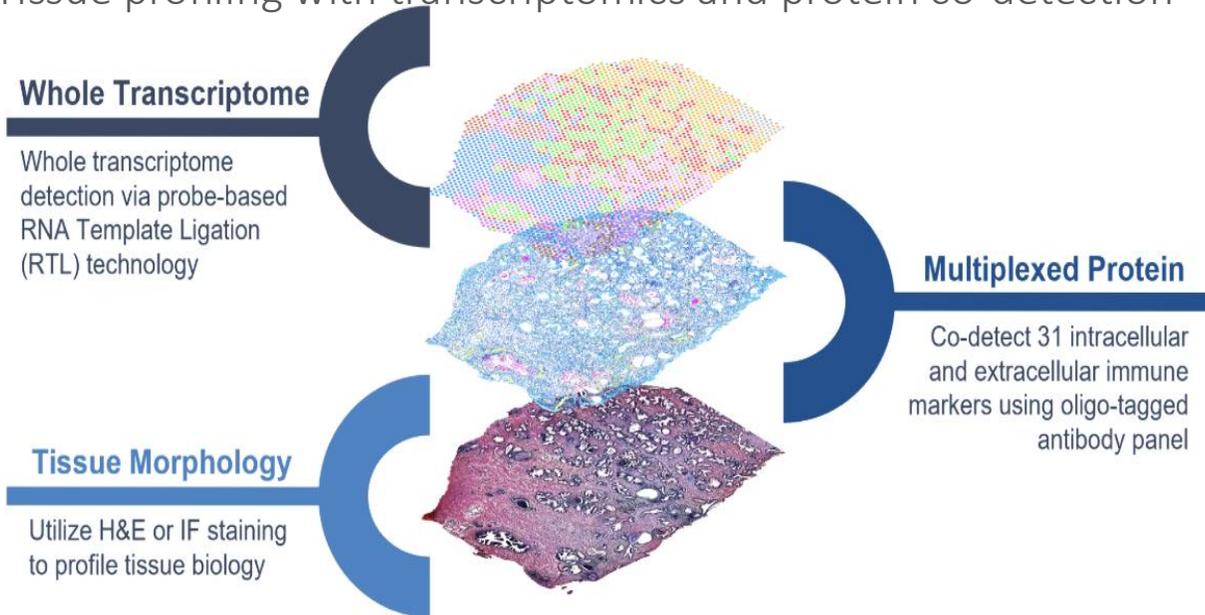
Source: Liao et al. Trends in Biotechnology. (2020)



Source: 10x Genomics

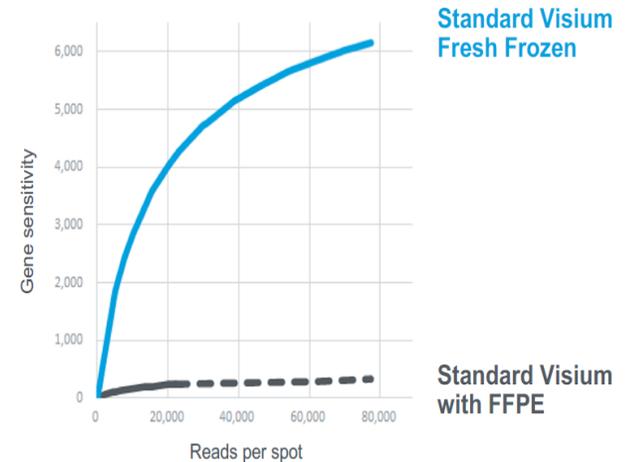
SPATIAL TRANSCRIPTOMICS

Tissue profiling with transcriptomics and protein co-detection



The Challenge of FFPE Samples

FFPE tissue preservation degrades RNA



- Visium FFPE uses probe based chemistry similarly to Fixed RNA profiling for single cell

ELEMENT BIOSCIENCES

5D Multiomic Insights

Explore biology at unprecedented depths with AVITI24™. Traditional multiomics approaches require multiple platforms, samples, and integration of disparate datasets. AVITI24 changes that with high-dimensional co-detection of RNA, morphology, protein, and their dynamic response and spatial relationships—all with no library prep. It's the platform designed for discovery.



Spatial Resolution

With precise single cell imaging, each analyte is spatially captured, revealing the exact intracellular locations of RNA and protein expression, as well as the intricate organization of structures within a sample.

RNA

Profile 350 transcripts with enhanced sensitivity and dynamic range. With automated onboard probe hybridization, and amplification, you can detect 1000s of transcripts per cell.

Morphology

Visualize nucleus, cell membrane, actin, endoplasmic reticulum, Golgi apparatus, and mitochondria. Reversible cell labels multiplex feature detection and onboard primary analysis performs real-time segmentation and feature extraction.

Protein

Map up to 138 cell surface, intracellular, and phospho proteins at subcellular resolution with automated onboard antibody binding, amplification, and detection by ABC sequencing.

Dynamic Response

Understand cellular responses over time, screen different compounds and dosages, or profile gene perturbations within a single run. Scale your throughput with 1-well, 12-well, and 48-well* formats for profiling of up to 96 samples in a dual sided run.

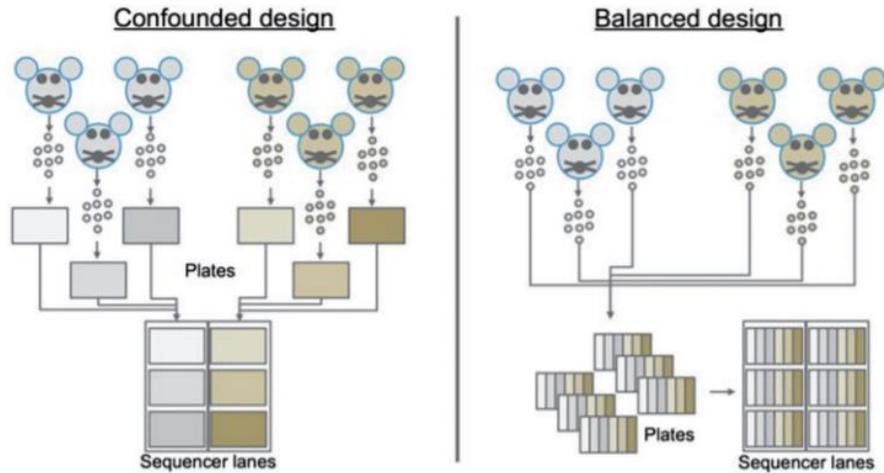
*48-well coming in 2025



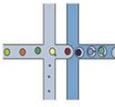
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EXPERIMENTAL DESIGN



Source: Baran-Gale et al. *Brief Func Genomics*. 17 (4):233–239. (2018)

I. Tissue Procurement			
	Source: <ul style="list-style-type: none"> - Primary human - Model organism - Cell culture 	Key considerations: <ul style="list-style-type: none"> - Biological variation - Sampling/handling variation - Duration of sourcing 	Study design: <ul style="list-style-type: none"> - Biological replicates - Technical replicates - Cell number calculation - Workflow optimization
II. Tissue Dissociation			
	Method: <ul style="list-style-type: none"> - Mechanical mincing - Enzymatic digestion - Automated blending - Microfluidics devices 	Key considerations: <ul style="list-style-type: none"> - Experimental consistency - Shortest duration - Highest cell/nucleus quality - Representation of all cell types 	Quality control: <ul style="list-style-type: none"> - FACS analysis - qPCR for marker genes - Imaging of cell integrity - RNA quality (RIN)
III. Cell Enrichment (optional)			
	Method: <ul style="list-style-type: none"> - Differential centrifugation, sedimentation, filtration - Antibody labeling for positive/negative selection - Flow cytometry or bead-based enrichment - Dead cell removal 	Key considerations: <ul style="list-style-type: none"> - Additional handling - Longer duration - Loss of RNA quality - Transcriptome changes 	
IV. Single Cell RNAseq Platform			
	Method: <ul style="list-style-type: none"> - Droplet-based - Tube-based after FACS - Microwell-based - Microfluidics-enabled 	Key considerations: <ul style="list-style-type: none"> - Cell throughput and handling time - Gene coverage and cell type detection - Whole transcript versus 3' end counting - Imaging capability for doublet detection 	
V. Library Sequencing			
	Method: <ul style="list-style-type: none"> - Illumina NGS - Compatible with cDNA library 	Sequencing depth considerations: <ul style="list-style-type: none"> - 3' end counting: low depth ~50K RPC - Whole transcript: high depth ~1M RPC - Alternative splicing: ~20-30M RPC - Iterative optimization for biological system 	
VI. Computational Analysis			
	Key considerations: <ul style="list-style-type: none"> - Separation of batch and condition - Technical vs. biological variation 	Sample Batch correction approaches: <ul style="list-style-type: none"> - Cell Hashing - Demuxlet - Canonical correlation analysis (CCA) - MAST 	

Source: Nguyen QH et al. *Front Cell Dev Biol* 6:108. (2018)

● WHAT PLATFORM SHOULD I USE?

Choose protocol based on:

- Throughput (number of cells per reaction)
- Sample of origin
- Cost / Labour / Time limitations
- Gene body coverage: 5' / 3' biased or full-length?
- UMI vs no-UMI
- Sequencing depth per cell

Examples:

- If your sample is fairly homogeneous – bulk RNAseq
- If your sample is limited in cell number – plate-based method
- If you want re-annotate the transcriptome and discover new isoforms – full-length coverage (SMART-seq2, seqWell)
- If you are looking to classify all cell types in a diverse tissue - high throughput
- If you have only archival human samples – nuclei isolation or 10x fixed RNA profiling

LITERATURE:

- https://hbctraining.github.io/scRNA-seq/slides/Single_Cell_2_27_20.pdf.
- <https://www.slideshare.net/TimothyTickle/introduction-to-singlecell-rnaseq>
- <https://www.decibio.com/insights/10x-genomics-single-cell-dominance-is-it-sustainable>
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- Wang et al. Direct Comparative Analyses of 10X Genomics Chromium and Smart-seq2. *Genom Proteom Bioinform* Apr;19(2):253-266 (2021).
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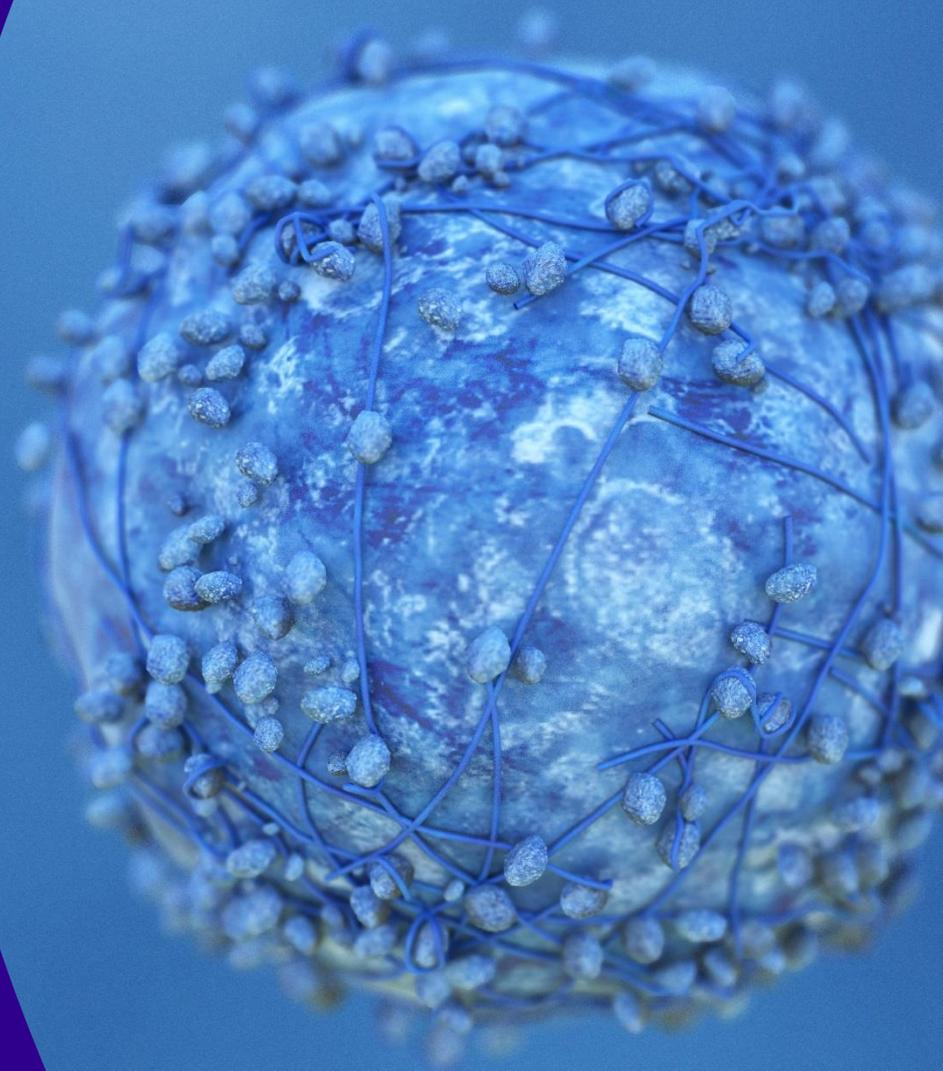
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USEFUL RESOURCES:

- A Single-Cell Sequencing Guide for Immunologists. *Frontiers in immunology*, 9, 2425. (2018).
- Single-cell immune profiling of γ -delta T-cells (<https://shorturl.at/IXEjc>)
- Van de Sande *et al.* Applications of single-cell RNA sequencing in drug discovery and development. *Nat Rev Drug Discov* (2023).
- Haque *et al.* A practical guide to single-cell RNA-sequencing for biomedical research and clinical applications. *Genome Med.* 2017;9(1):75.
- Single cell course by Hemberg Lab, Wellcome Sanger Institute (<http://hemberg-lab.github.io/scRNA.seq.course/index.html>)
- Tabula Muris (<https://tabula-muris.ds.czbiohub.org/>)
- Human Cell Atlas (<https://www.humancellatlas.org/>)
- Worthington Tissue Dissociation Guide (<http://www.worthington-biochem.com/tissuedissociation/default.html>)
- Broad Institute Single Cell Portal (https://singlecell.broadinstitute.org/single_cell)
- List of software packages for single cell data analysis (<https://github.com/seandavi/awesome-single-cell>)
- SPLIT-seq (<https://www.youtube.com/watch?v=WqaeZe7mKUc>)
- CITE-seq (<https://cite-seq.com/>)
- Biolegend TotalSeq (<https://www.biolegend.com/en-us/totalseq>)
- Can chatGPT do single-cell bioinformatic analysis? <https://www.youtube.com/watch?v=fkuLFIC2ZWk>



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