

Integrative Genomics Viewer IGV

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- Introduction to IGV.
 - What is IGV.
 - How to run IGV.
- Navigating IGV.
 - The IGV user interface.
 - Moving around genomes.
- Loading and visualising data.
 - Genome information and annotation.
 - User supplied data.
 - Sample information.
 - External data.
- Displaying genomics data
 - Basic visualisation.
 - Data dependent visualisation.

What is IGV?

- Created by the Broad institute.
- Genome browser.
 - Visualises genomic data (expression, ChIP, resequencing, multiple alignment, shRNA)
 - Handles most common genomic data types.
- Java Desktop application
 - No dependence on server
 - Loads data locally or from URL, consumes memory and CPU.

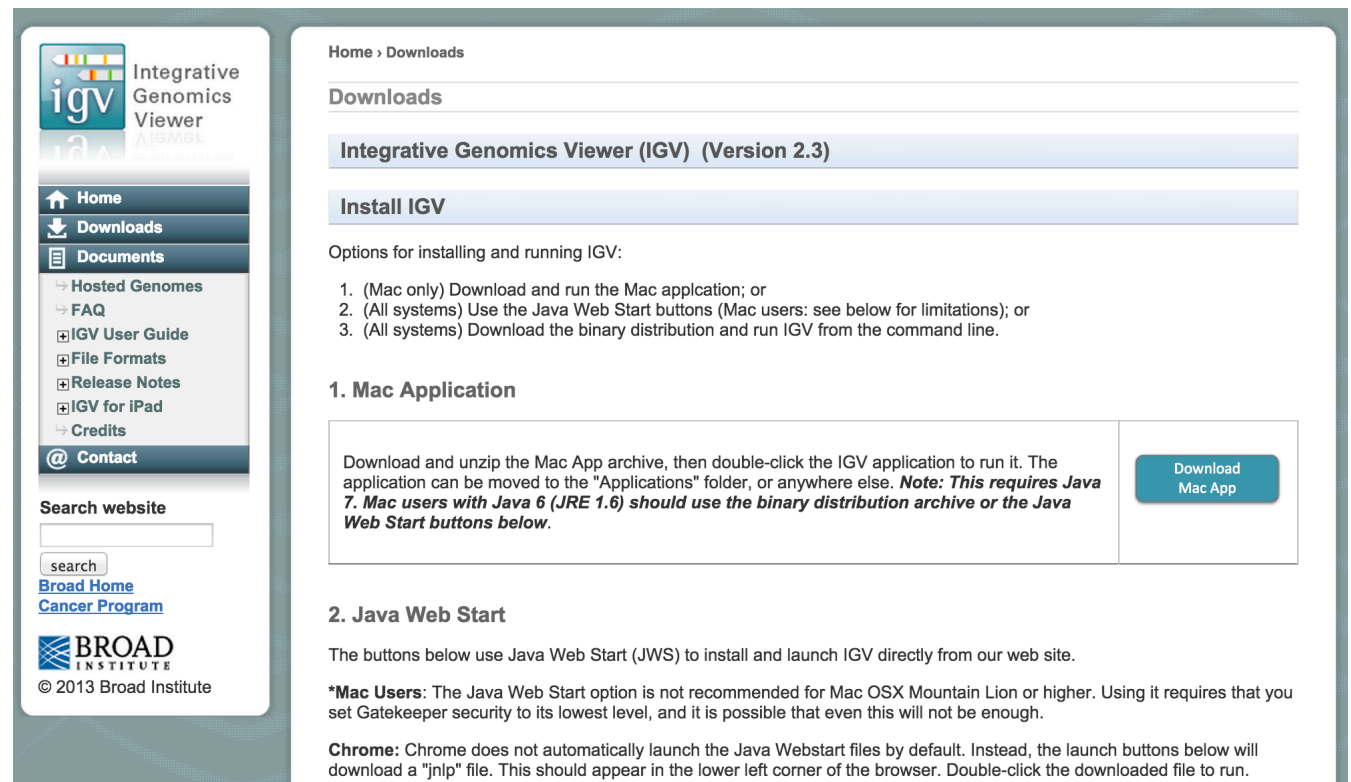
How to run IGV?

- Requires Java



- IGV available from Broad

<http://www.broadinstitute.org/software/igv/download>

A screenshot of the Integrative Genomics Viewer (IGV) download page. The page has a sidebar on the left with navigation links: Home, Downloads, Documents, Hosted Genomes, FAQ, IGV User Guide, File Formats, Release Notes, IGV for iPad, Credits, and Contact. The main content area is titled "Home > Downloads" and "Downloads". It features a section for "Integrative Genomics Viewer (IGV) (Version 2.3)" with an "Install IGV" button. Below this, it lists "Options for installing and running IGV:" with three numbered steps: 1. (Mac only) Download and run the Mac application; or 2. (All systems) Use the Java Web Start buttons (Mac users: see below for limitations); or 3. (All systems) Download the binary distribution and run IGV from the command line. Under "1. Mac Application", there is a text box describing the installation process and a "Download Mac App" button. Under "2. Java Web Start", it explains that the buttons use Java Web Start (JWS) to install and launch IGV directly from the website. It also includes a note for Mac users about Gatekeeper security and a note for Chrome users about downloading a ".jnlp" file.

- Download to computer.
- Runs locally.
- Archived versions available

3. Binary Distribution

Download and unzip the binary distribution archive in a folder of your choosing. IGV is launched from a command prompt -- follow instructions in the "readme" file. To launch igv on Mac or Linux platforms use the shell script "igv.sh". On Windows use "igv.bat".

Download
Binary Distribution

igvtools

Utilities for preprocessing data files.

- [igvtools_2.3.40.zip](#)

Other IGV Versions

[Development Snapshot Build](#) *Latest development snapshot; built at least nightly.*

[Archived Versions](#)

- Runs from webstart.
- Always runs latest version of IGV.

2. Java Web Start

The buttons below use Java Web Start (JWS) to install and launch IGV directly from our web site.

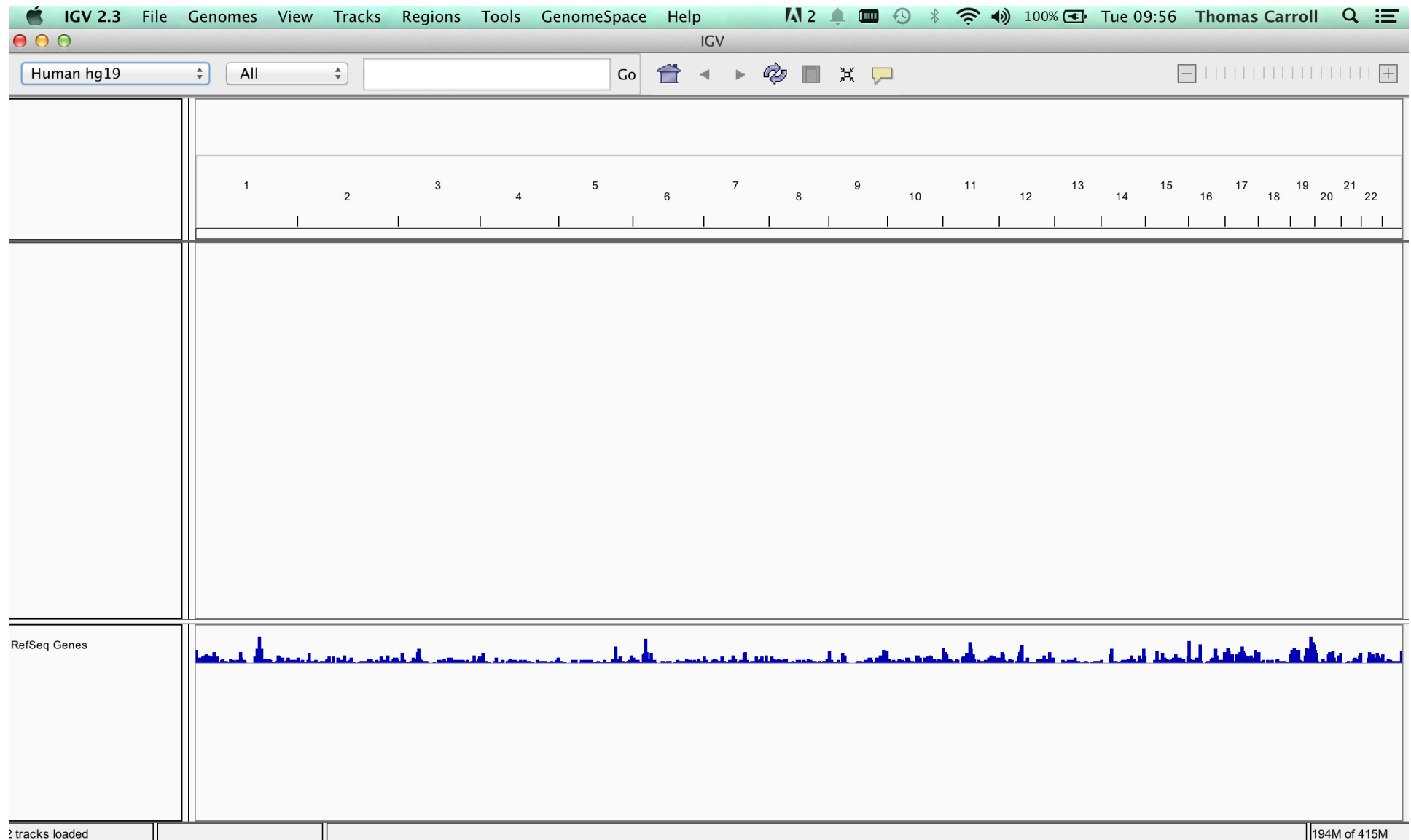
***Mac Users:** The Java Web Start option is not recommended for Mac OSX Mountain Lion or higher. Using it requires that you set Gatekeeper security to its lowest level, and it is possible that even this will not be enough.

Chrome: Chrome does not automatically launch the Java Webstart files by default. Instead, the launch buttons below will download a "jnlp" file. This should appear in the lower left corner of the browser. Double-click the downloaded file to run.

Windows users: To run with more than 1.2 GB of memory you must install 64-bit Java. ***Most Windows installs do not include 64-bit Java by default, even if the operating system is 64-bit.*** Attempting to use the 2GB or greater launch options with 32-bit Java will result in the error "*could not create virtual machine*".

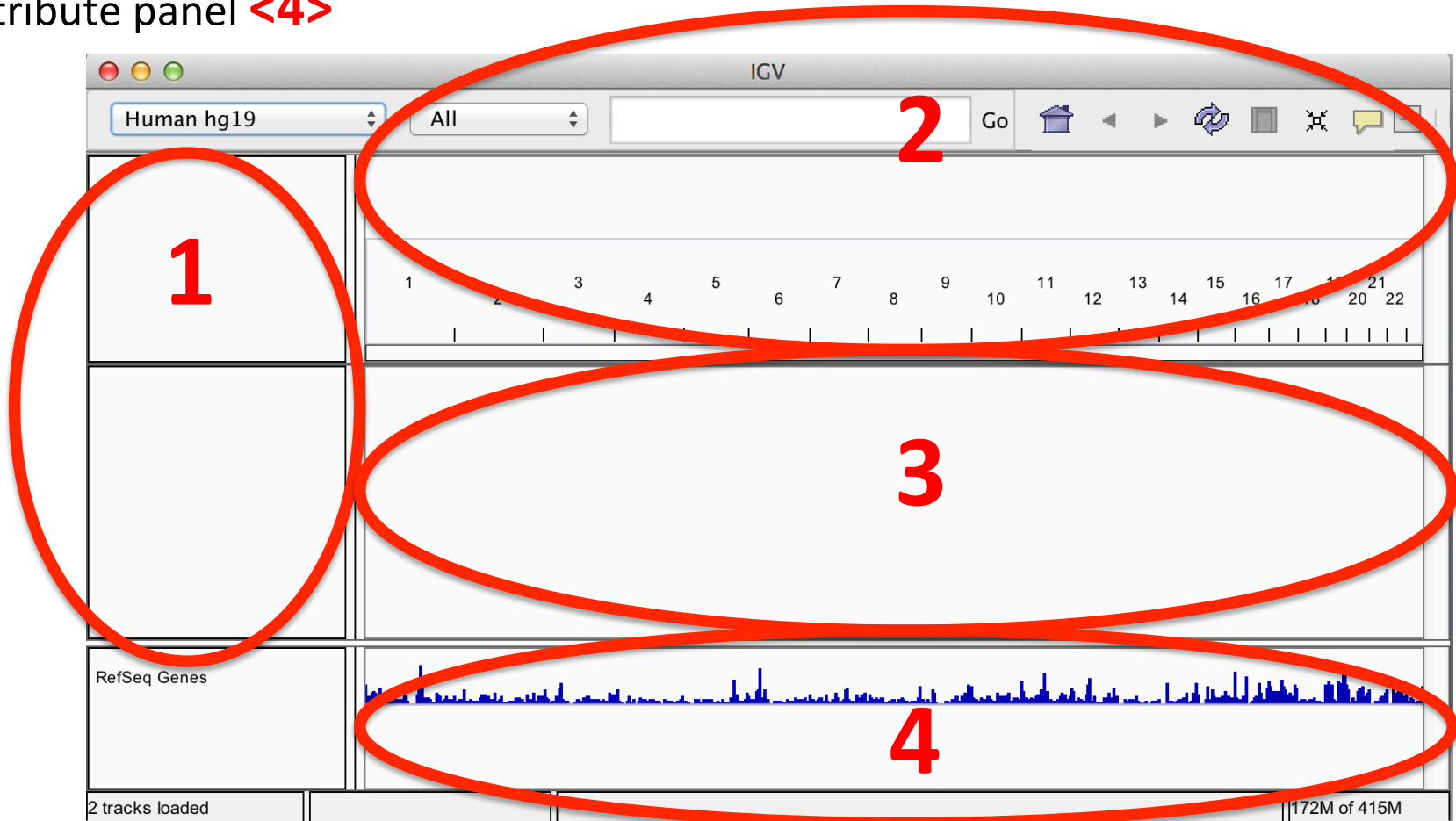
 Launch with 750 MB	 Launch with 1.2 GB Maximum usable memory for Windows OS with 32-bit Java.	 Launch with 2 GB Maximum usable memory for 32-bit MacOS.	 Launch with 10 GB For large memory machines with 64-bit Java.
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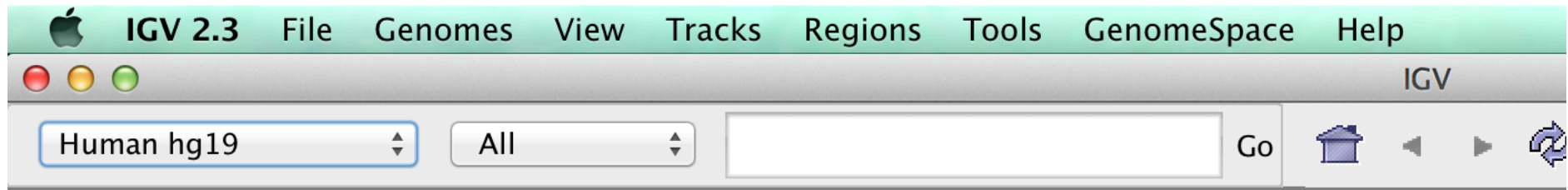
IGV GUI



- IGV GUI:-

- Sample information panel <1>
- Genome Navigation panel <2>
- Data panel <3>
- Attribute panel <4>

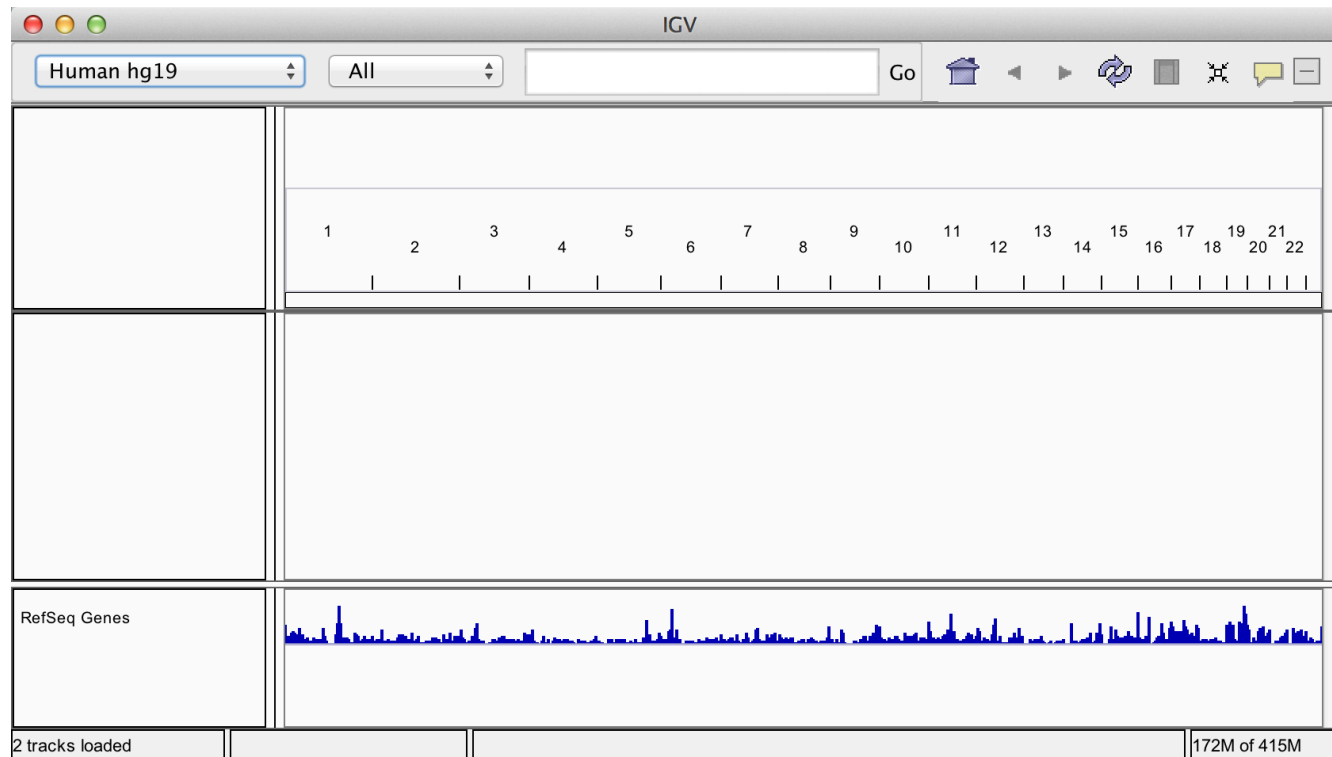




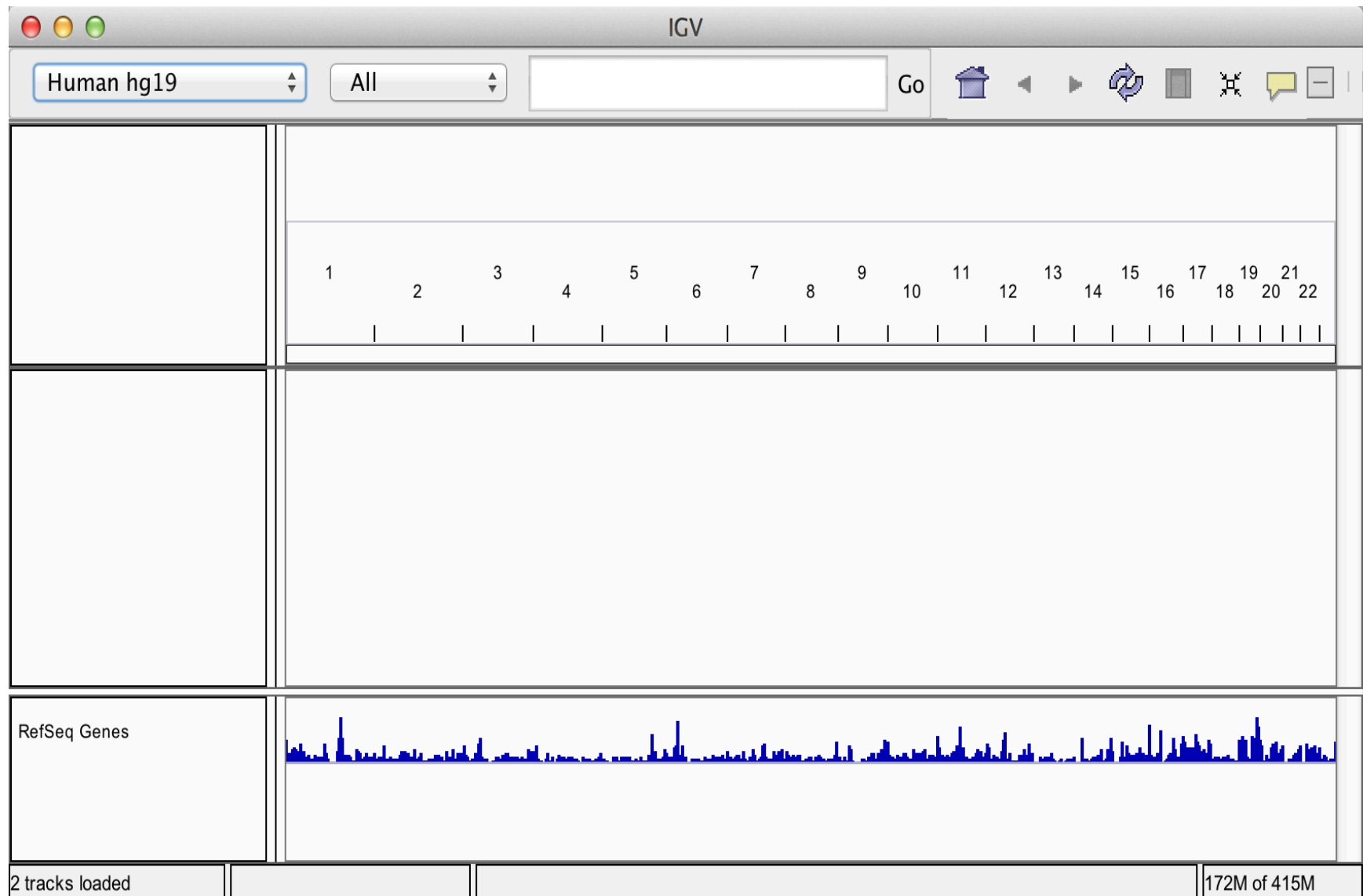
- Menu bar
 - **File**, Load data/sample information.
 - **Genome**, Load and manage genomes.
 - **View**, Display preferences.
 - **Tracks**, Group/sort/filter data tracks.
 - **Regions**, Create region/gene lists.
 - **Tools**, Access to Integrated tools (IGVtools/Bedtools).
 - **GenomeSpace**, Export/import from Genomespace

Moving around genomes

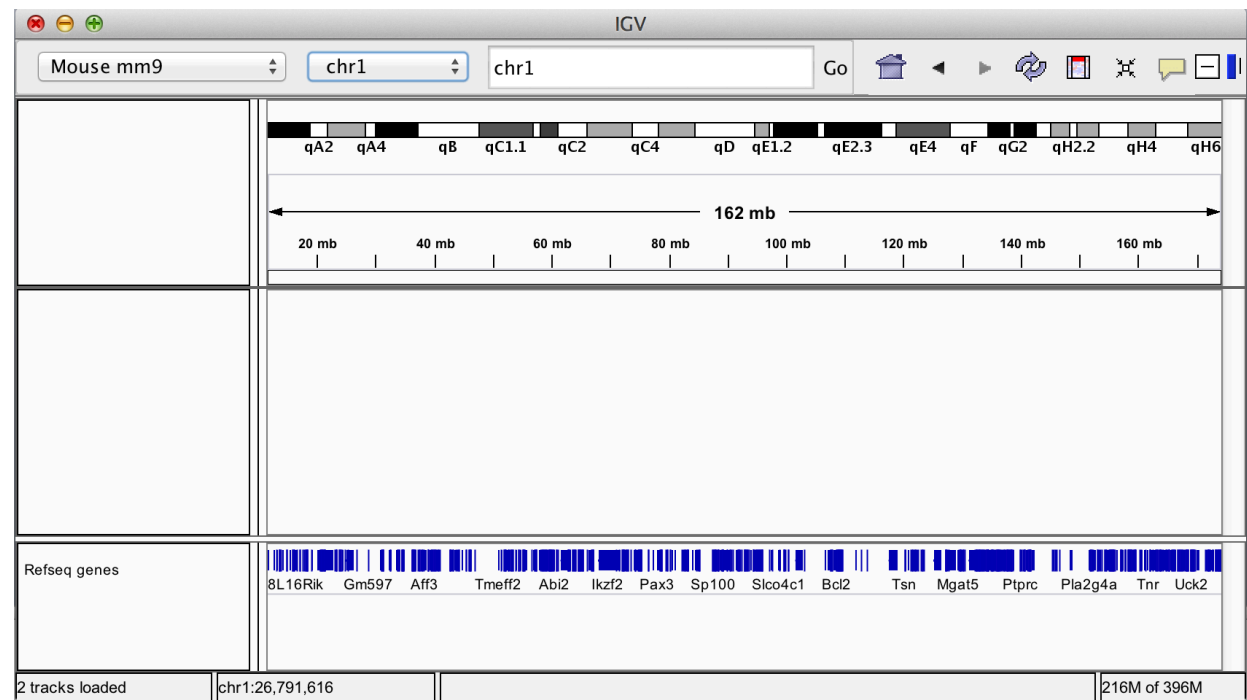
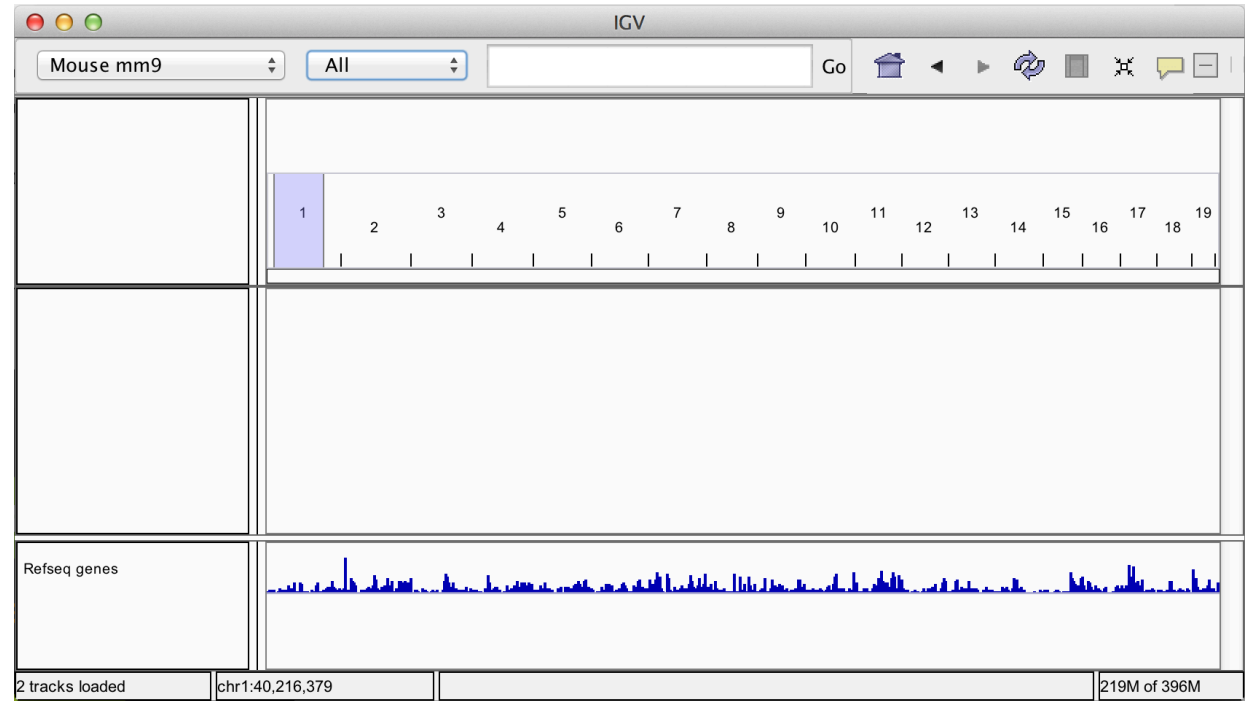
- Cytoband selection and zooming
- Scrolling
- Selection of region of interest



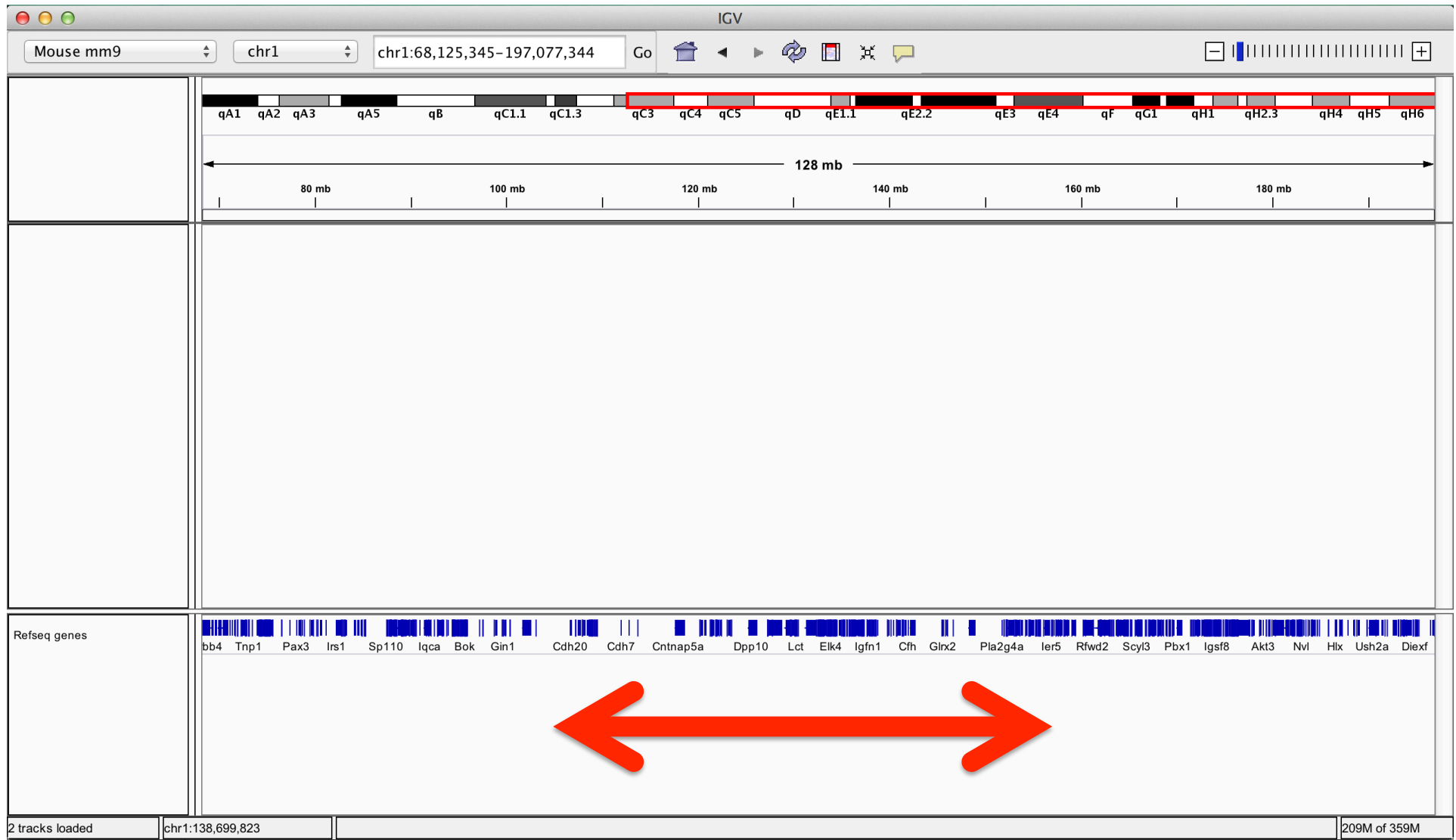
Whole genome view



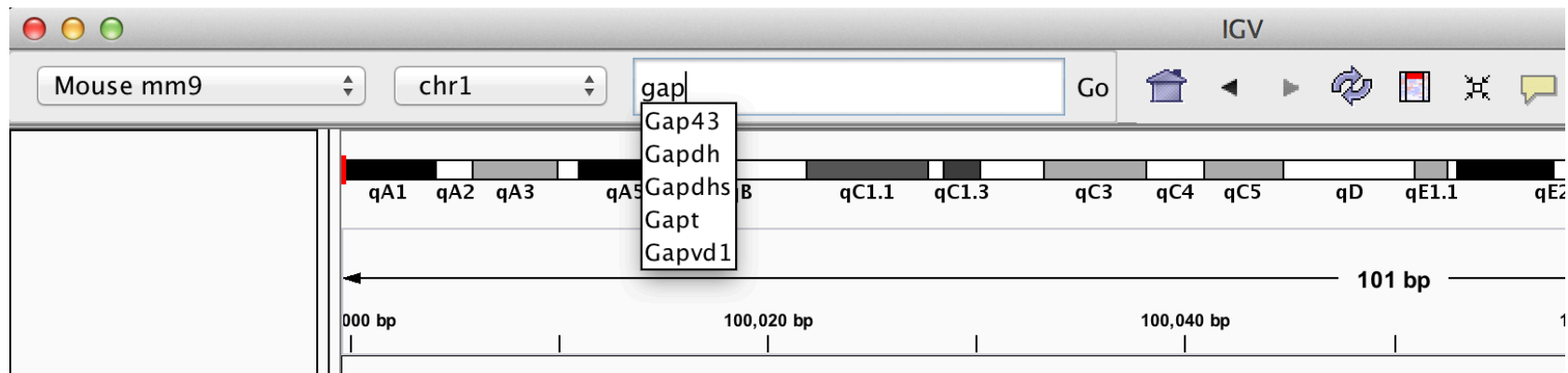
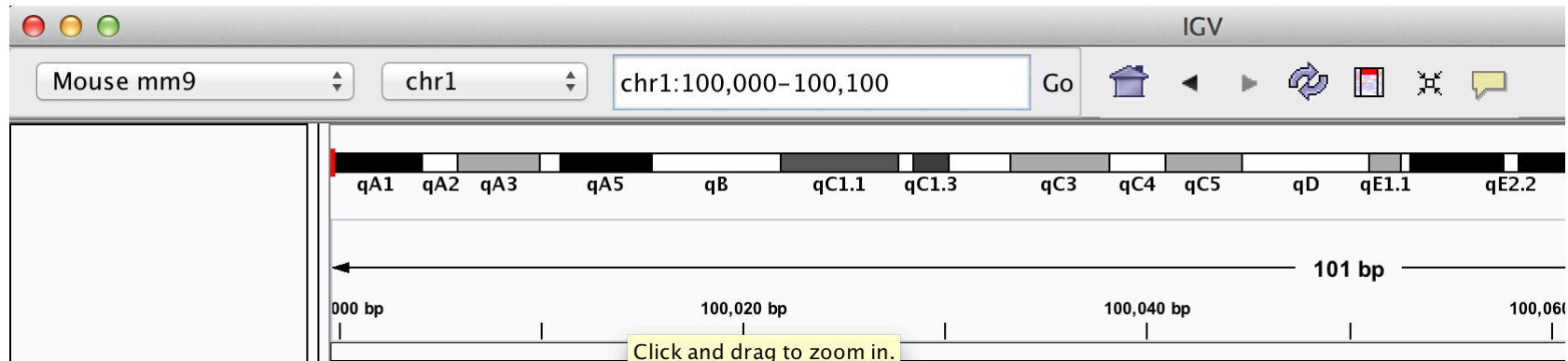
Zooming



Scrolling

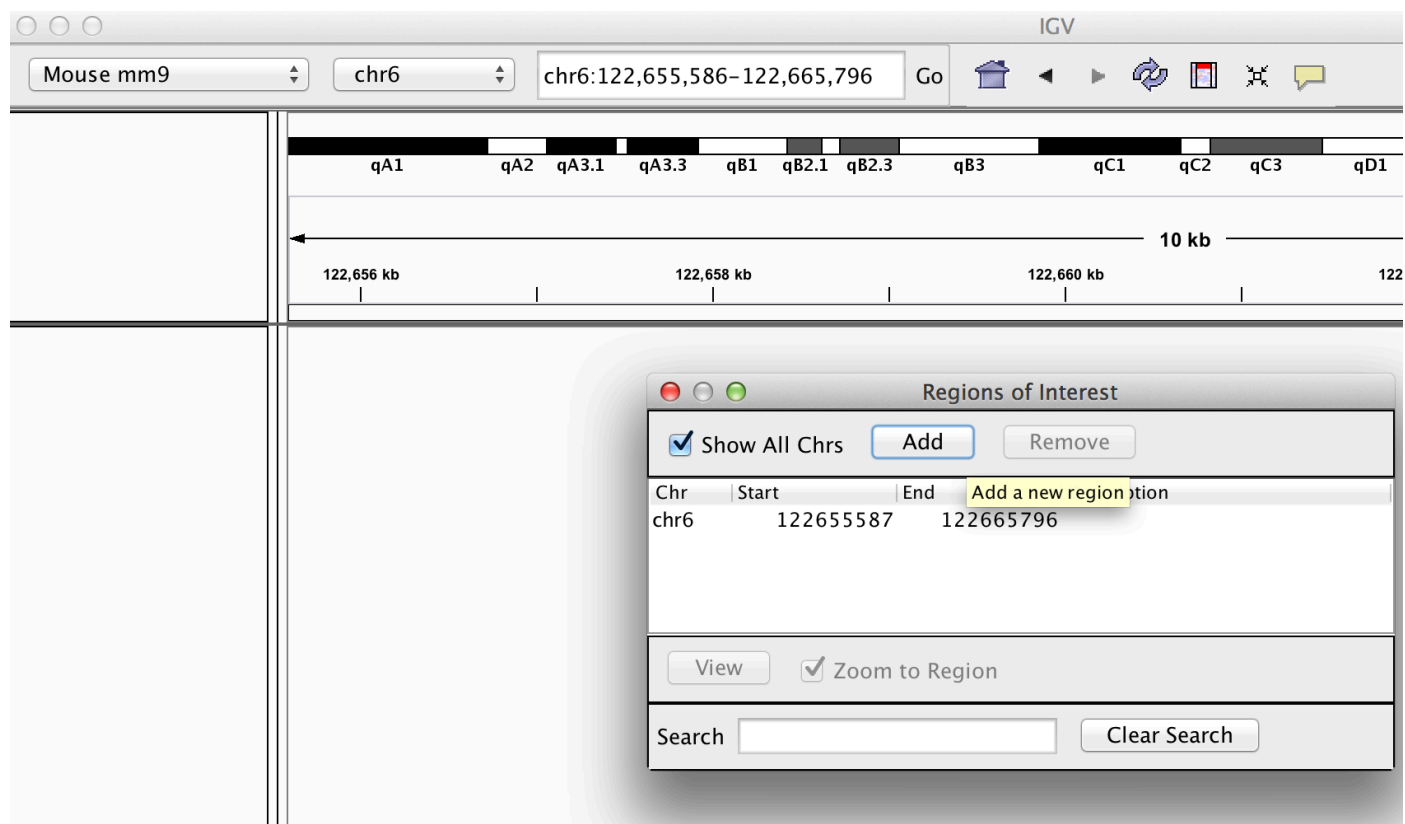
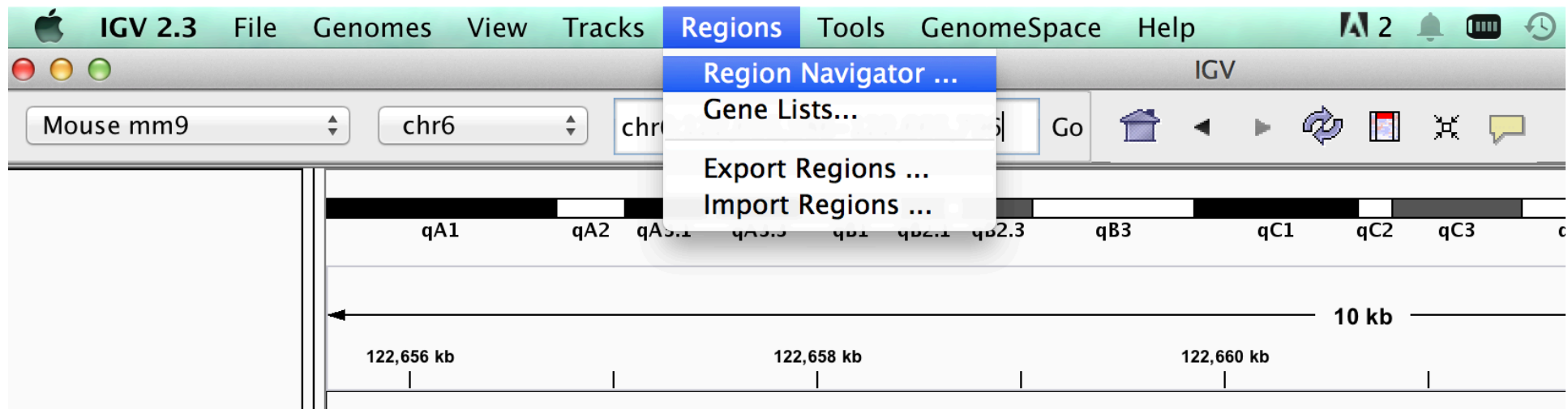


Jump to Region



“Bookmarking” regions of interest

- Regions may be added to “Regions of interest”
- These act as bookmarks for areas of particular interest



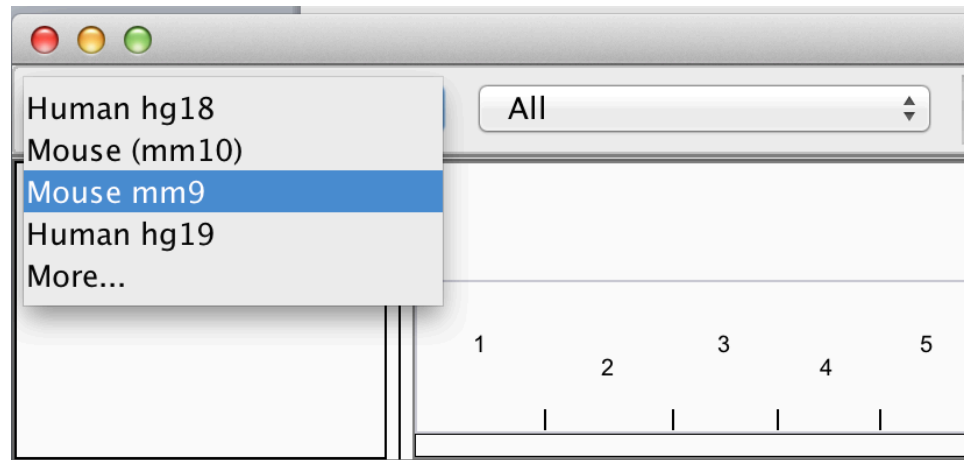
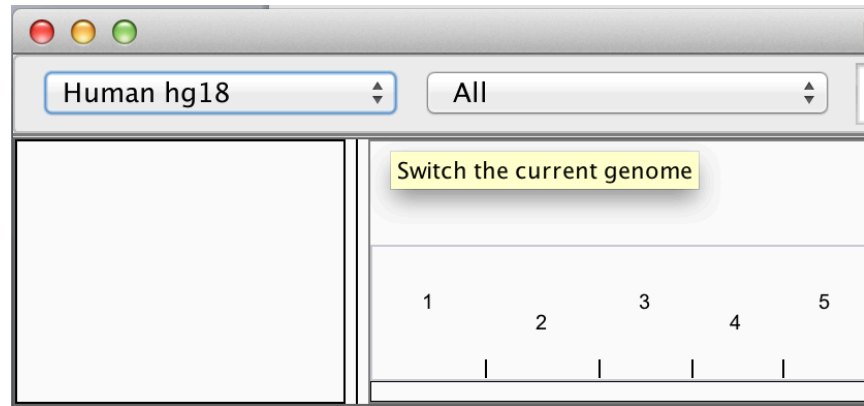
“Book marked” Region Of Interest



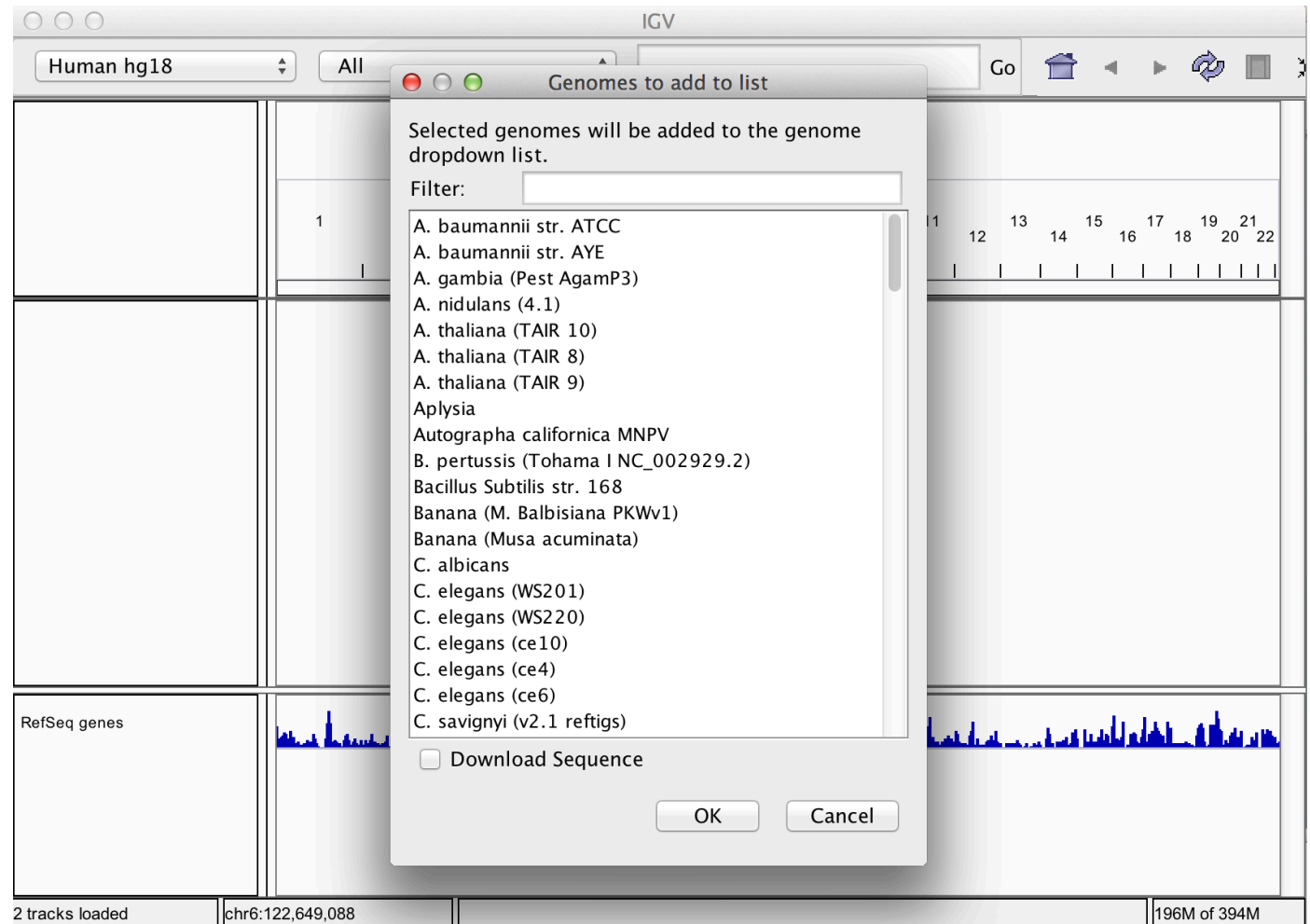
Loading data in IGV

Loading Genome Information

- Most genomes can be selected from dropdown.

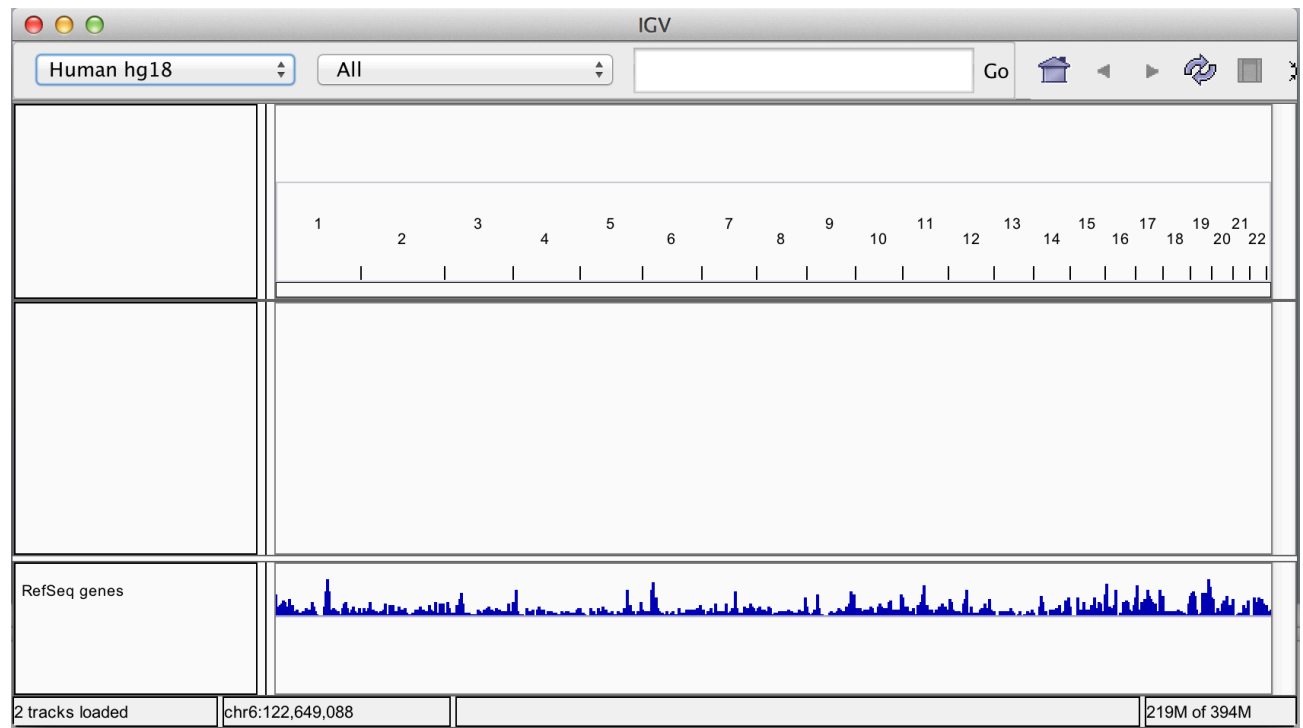


- Genomes not included may be downloaded from repository



Loading genome annotation

- For supported genomes, gene positions are automatically included in “feature” panel.
- Additional gene positions can be loaded into IGV in gff format.

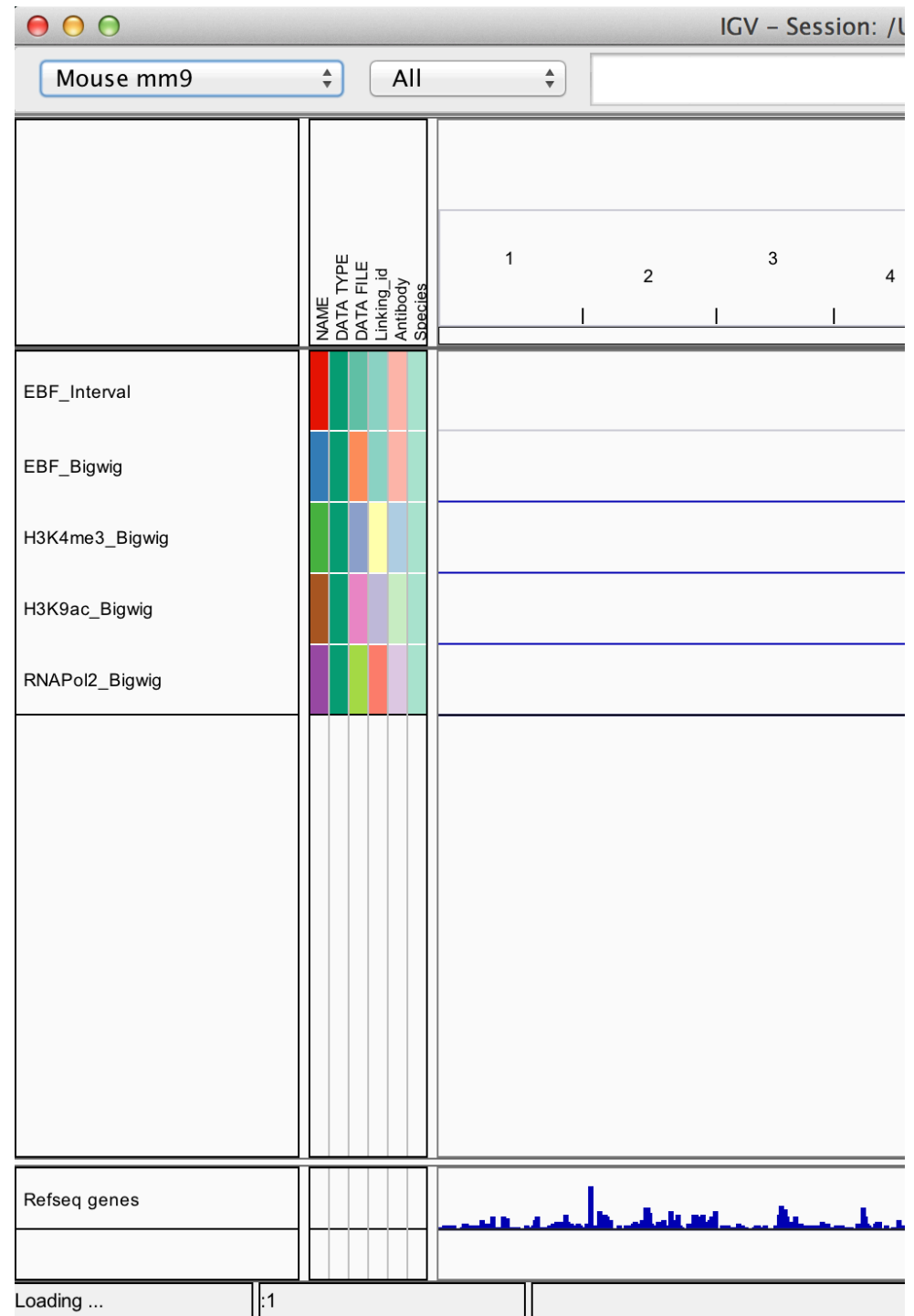


Loading sample data.

- Most common formats can be loaded into IGV through file menu
 - Acceptable data formats include:-
 - BED (.bed)
 - BAM and index (.bam with .bai/.bam.bai)
 - BigWig (.bw)
 - BedGraph/Wig (.bedGraph, .wig)
- And many more...

Loading sample metadata

- IGV allows the inclusion of information on samples.
- Sample information is then included in sample information panel.



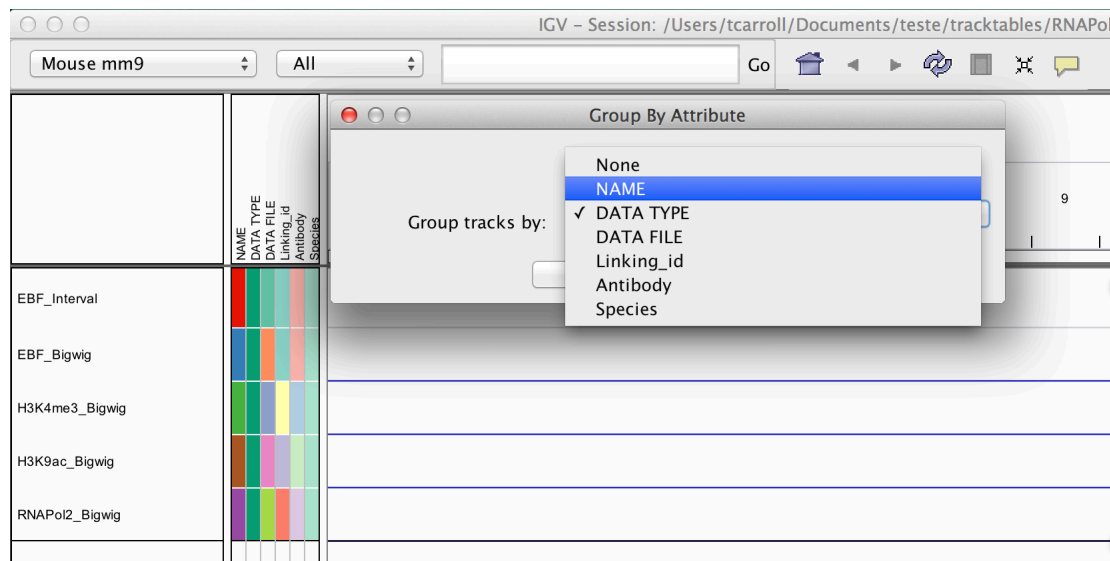
- Example Sample Information file

<http://www.broadinstitute.org/igvdata/exampleFiles/exampleSampleInfo.txt>

TRACK_ID	Data_Type	PARTICIPANT_ID	SAMPLE_ID	GENDER	T/N	Tumor_type	Treated	Primary/ Secondary	Hypermuted
EX-01-001	Expression	P-01-P001	P-01-S001	M	Tumor	GBM	Y	Primary	Y
CN-01-002	CopyNumber	P-01-P001	P-01-S001	M	Tumor	GBM	Y	Primary	Y
MU-01-003	Mutation	P-01-P001	P-01-S002	M	Tumor	GBM	Y	Primary	Y
EX-01-004	Expression	P-01-P002	P-01-S003	M	Normal	GBM	Y	Secondary	Y
CN-01-005	CopyNumber	P-01-P002	P-01-S004	M	Tumor	GBM	Y	Secondary	N
EX-01-006	Expression	P-01-P002	P-01-S004	M	Tumor	GBM	Y	Secondary	N
ME-01-007	Methylation	P-01-P002	P-01-S004	M	Tumor	GBM	Y	Secondary	N
EX-01-008	Expression	P-01-P003	P-01-S006	F	Tumor	GBM	N	Primary	Y
EX-01-009	Expression	P-01-P004	P-01-S009	F	Tumor	GBM	N	Primary	Y
EX-01-0010	Expression	P-01-P005	P-01-S0011	M	Control				

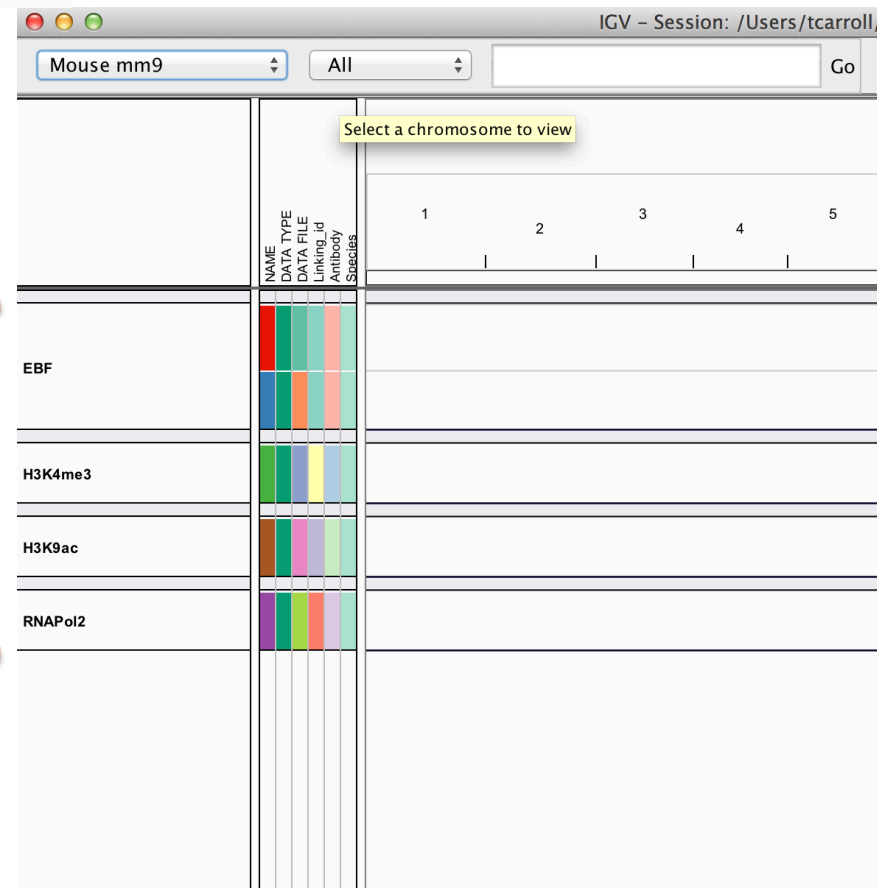
Using sample information.

- Sample information can include discrete and continuous.
- Can be used to “sort” and “filter” tracks.
- Can split tracks across panels by “group”



**Grouped by
unique ID**

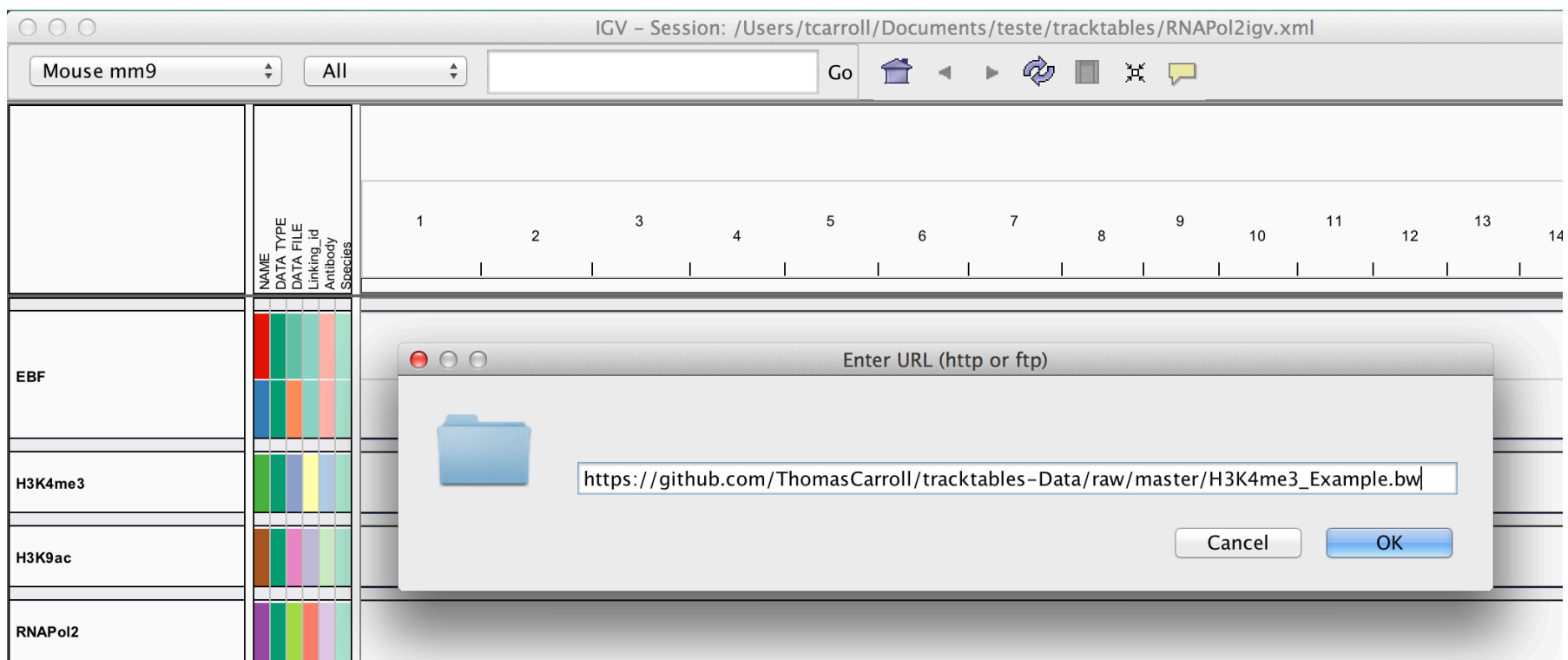
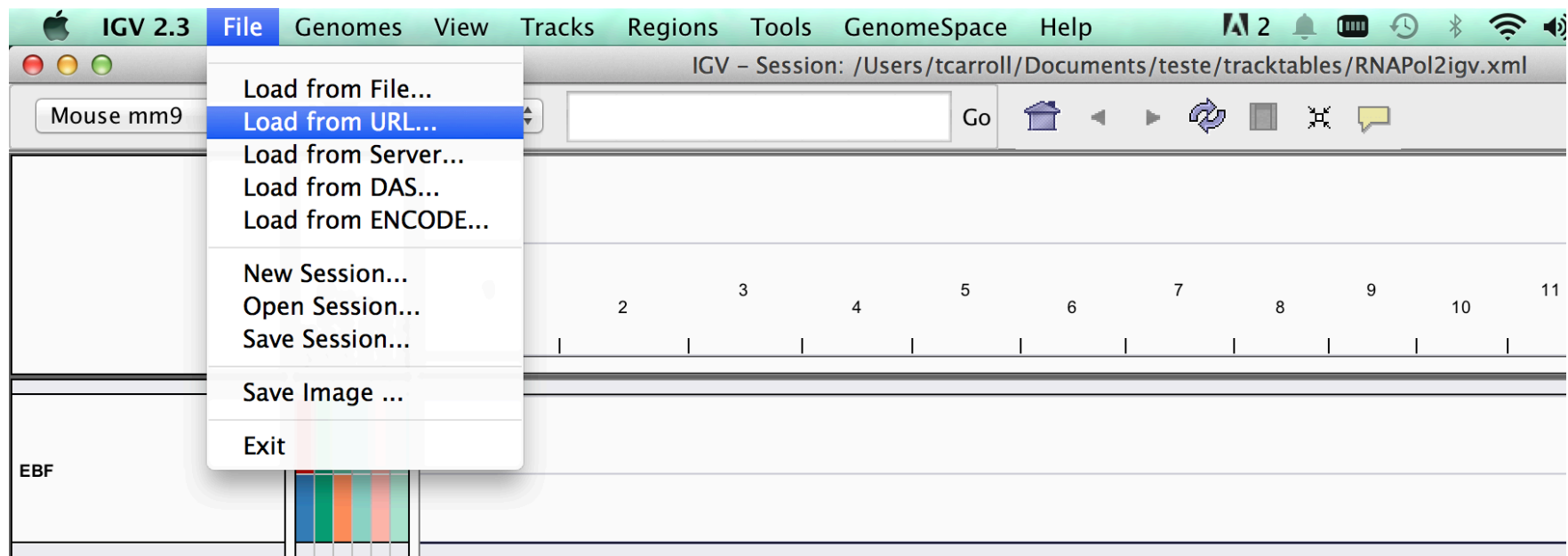
**Grouped related files
by name**



Loading external data and annotation

Load data from a URL.

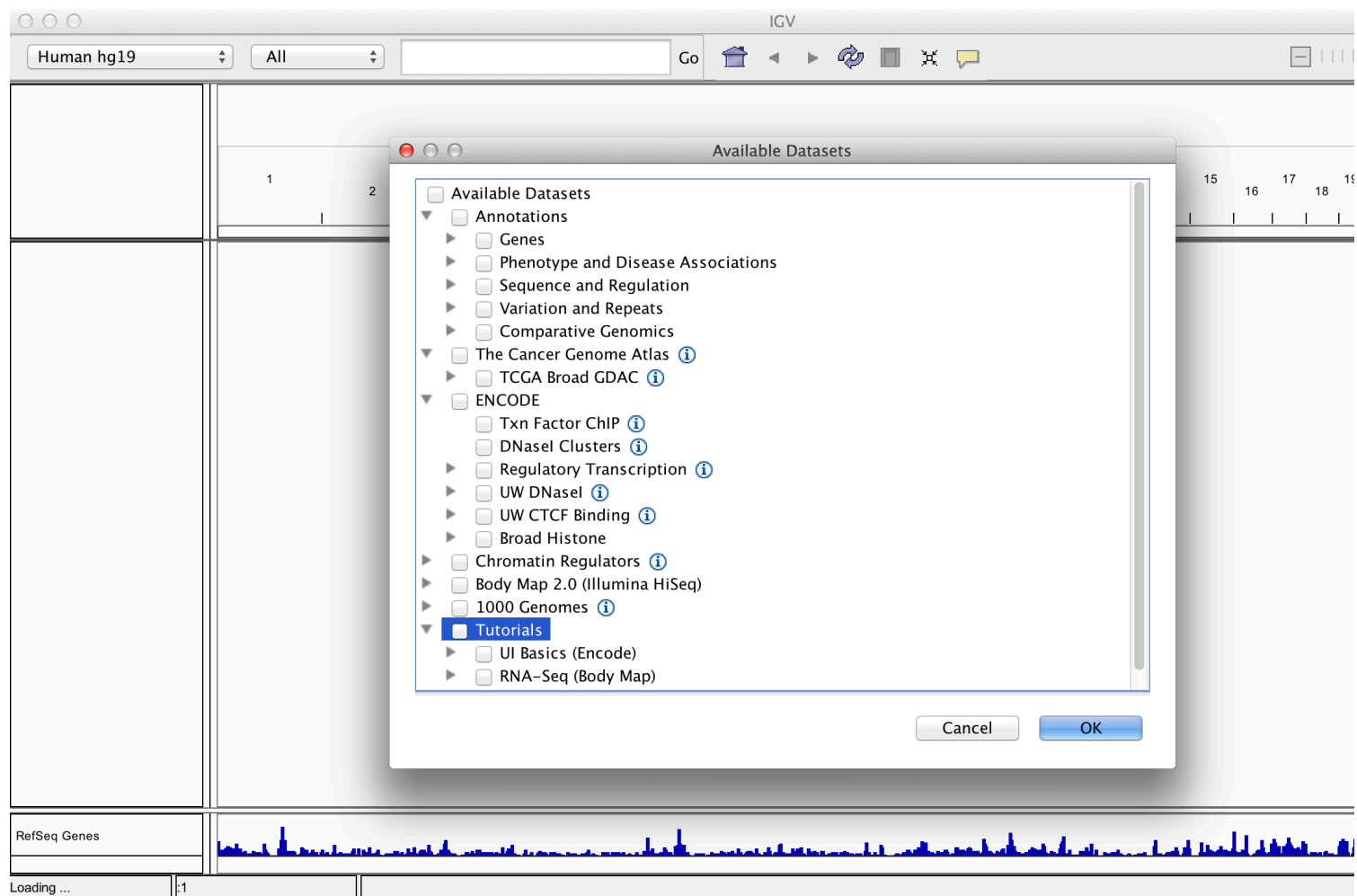
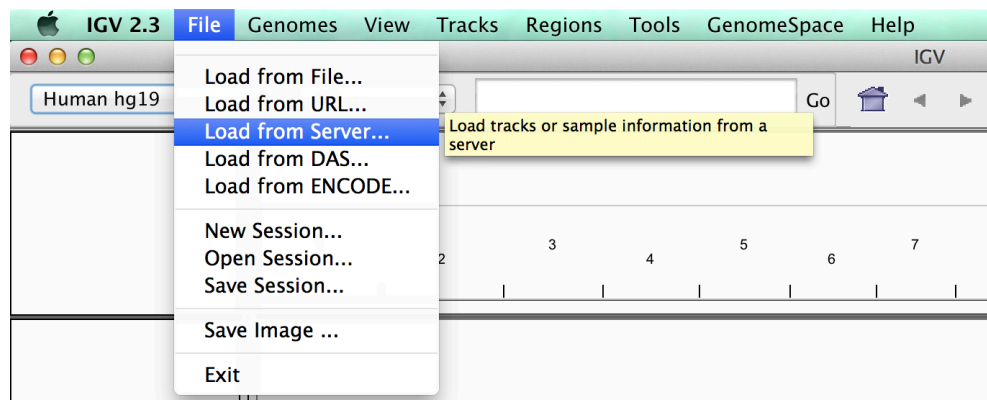
- As with UCSC, IGV supports data hosted on external servers.
- Data accessible from a URL such as HTTP and FTP can be loaded using the **“Load from URL”**.



Loading external data and annotation

Load data from a server.

- Unlike UCSC, IGV comes with few external tracks.
- External tracks (relevant to the genome) can be loaded from the IGV server or Encode-IGV server.



Viewing data

- IGV associates common file formats with default display methods.
- Most of the time IGV will make a sensible choice how we wish to display data.

Accepted formats and default display.

Information on accepted file formats and default display can be found at <http://www.broadinstitute.org/software/igv/RecommendedFileFormats>

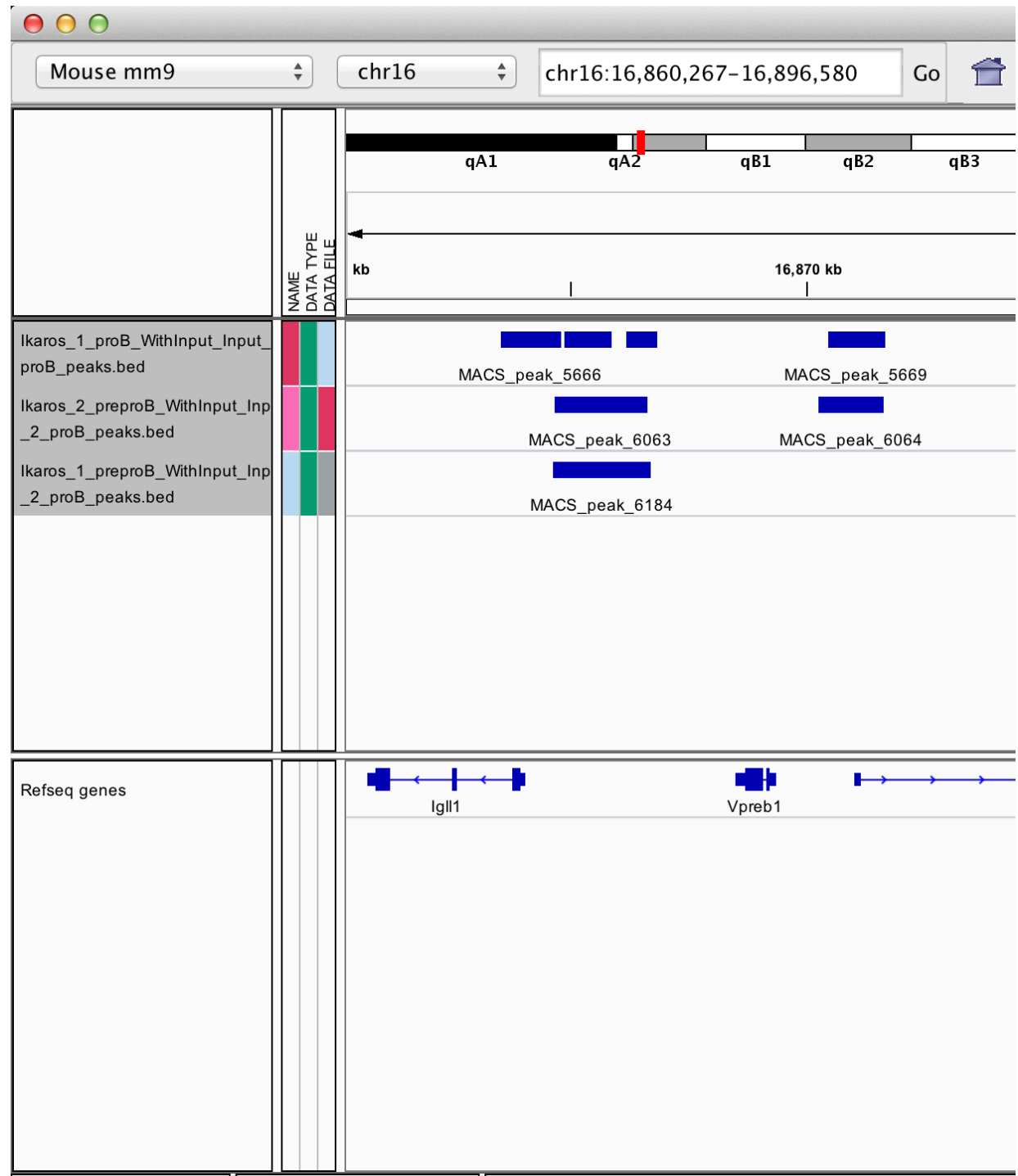
Recommended File Formats

The following table lists recommended file formats for data types. **Tip:** Using [igvtools](#) to convert a source data file to a binary TDF file can reduce loading time and improve performance.

Source Data	Recommended File Formats
ChIP-Seq, RNA-Seq	TDF format. Use the igvtools package (count command) to generate a binary read count density file in TDF format. Load the resulting TDF file into IGV.
Copy number	CN format, SNP format Log values: If copy number data contains negative values, IGV assumes they are scaled log values and displays them without modification. If copy number data contains all positive values, IGV assumes they are unscaled. It centers the unscaled values around 2 (1 for allele specific files), log transforms them ($\text{logValue} = \log_2(\text{copyNumber} / 2)$), and displays the log values.
Gene expression data	GCT format. RES format

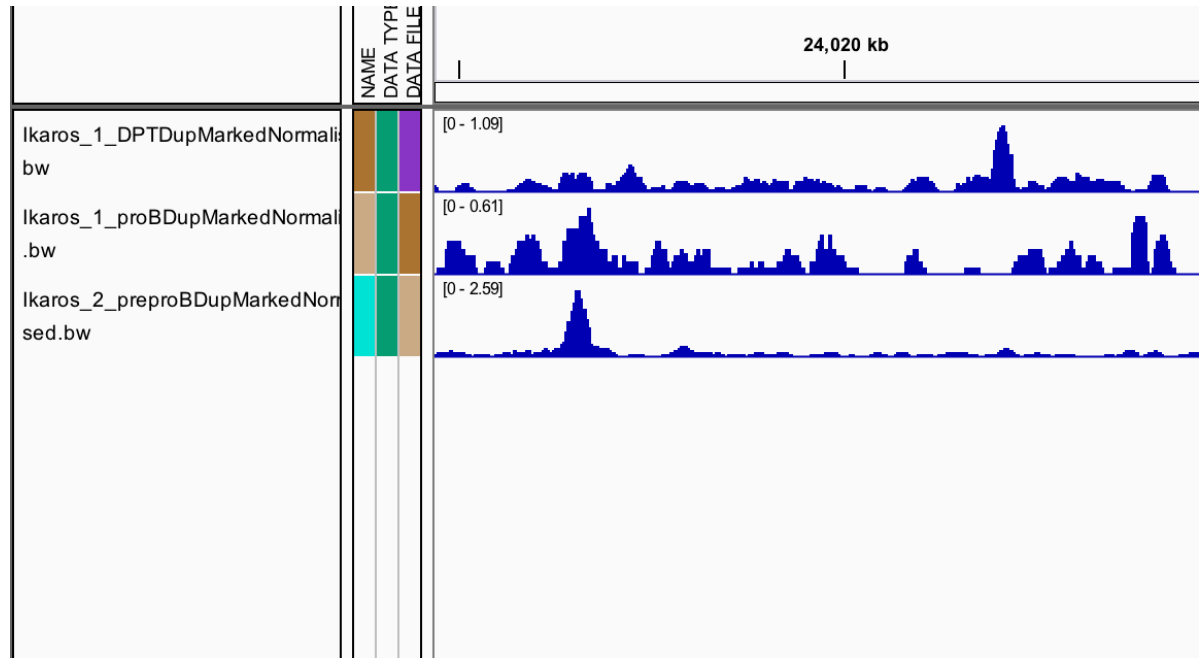
Bed/bigBed

- Basic
 - Tab-delimited
Chrom,Start,End
- Bed6
- bigBed
(recommended)



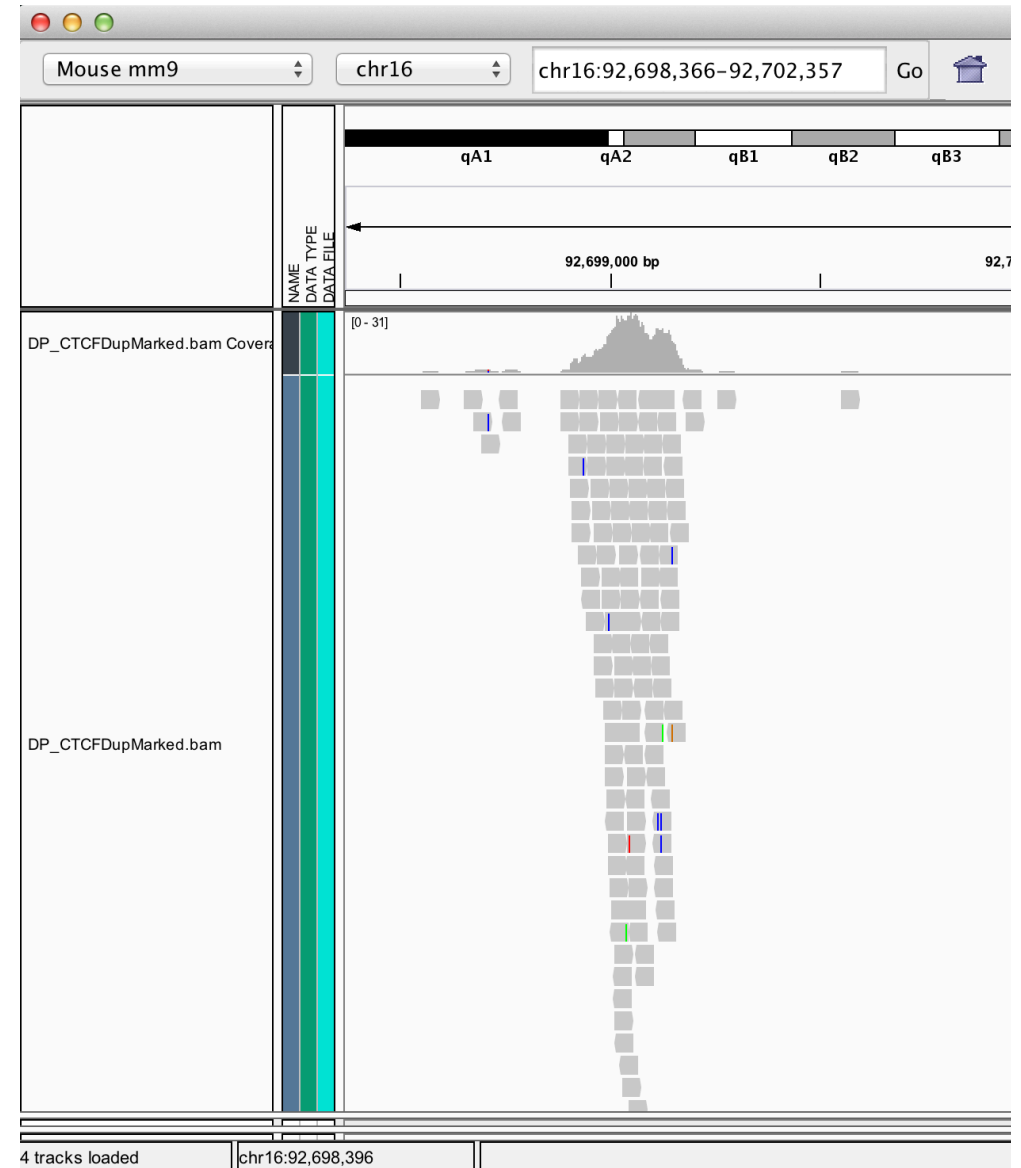
Wig, BedGraph and BigWig

- Wig/bedGraph require high memory load
- Recommended format is bigWig



BAM alignment files

- BAM files contain alignment information.
- Require an accompanying .bai index file for display



Finer control of display

- IGV allows for customization of track display.
 - Menu bar -> View -> Preferences
 - Select track (right click)

Display preferences

General

The screenshot shows a 'Display preferences' dialog box with a 'General' tab selected. The dialog has a title bar with standard macOS window controls. The 'General' tab is active, showing various settings. The 'Distinguish missing data' checkbox is unchecked, with a description below it. The 'Display all tracks in a single panel' checkbox is unchecked. The 'Show attribute panel' and 'Show default track attributes' checkboxes are checked. The 'Show region boundaries' checkbox is unchecked. The 'Zoom to features' checkbox is checked, with a description below it. There are two text input fields: 'Sequence resolution threshold (bp/pixel)' with the value '2' and 'Feature flanking region (bp or %)' with the value '2000'. At the bottom, there is a 'Background color click to change)' label with a color swatch and a 'Reset to default' button, and a 'Default font:' label with a text field containing 'Arial 10' and a 'Change...' button. 'OK' and 'Cancel' buttons are at the bottom right.

General Tracks Mutations Charts Alignments Probes Proxy Advanced IonTorrent

☐ Distinguish missing data
Distinguish regions with zero values from regions with no data on plots (e.g. bar charts). Regions with no data are indicated with a gray background.

☐ Display all tracks in a single panel

☒ Show attribute panel

☒ Show default track attributes
Display default track attributes (NAME, DATA_TYPE, and DATA_FILE) in the attribute panel.

☐ Show region boundaries

☒ Zoom to features
This option controls the behavior of feature searches. If true, the zoom level is changed as required to size the view to the feature size. If false the zoom level is unchanged.

Sequence resolution threshold (bp/pixel):
Resolution in base-pairs per pixel at which sequence track becomes visible.

Feature flanking region (bp or %):
Added before and after feature locus when zooming to a feature. Also used when defining panel extents in gene/loci list views. A negative number is interpreted as a percentage.

Background color click to change):

Default font:

Tracks

General Tracks Mutations Charts Alignments Probes Proxy Advanced IonTorrent

Default Track Height, Charts (Pixels) *Default height of chart tracks (barcharts, scatterplots, etc)*

Default Track Height, Other (Pixels) *Default height of all other tracks*

Track Name Attribute

Name of an attribute to be used to label tracks. If provided tracks will be labeled with the corresponding attribute values from the sample information file

☐ Expand Feature Tracks

☐ Show Expand Icon

☐ Normalize Coverage Data

Applies to coverage tracks computed with igvtools (.tdf files). If selected coverage values are scaled by (1,000,000 / totalCount), where totalCount is the total number of features or alignments.

OK Cancel

Alignments

The screenshot shows a software window with multiple tabs: General, Tracks, Mutations, Charts, **Alignments**, Probes, Proxy, Advanced, and IonTorrent. The 'Alignments' tab is active and contains the following settings:

- Visibility range threshold (kb):** 30. *Nominal window size at which alignments become visible*
- ☒ **Downsample reads** Max read count: 100 per window size (bases): 50
- Filter and shading options**
 - Coverage allele-freq threshold: 0.2 Mapping quality threshold: 0
 - ☒ Filter duplicate reads ☒ Show center line
 - ☒ Filter vendor failed reads ☒ Show coverage track
 - ☐ Filter secondary alignments ☐ Show soft-clipped bases
 - ☐ Filter supplementary alignments ☐ Flag unmapped pairs
 - ☒ Shade mismatched bases by quality: 5 to 20
 - ☐ Flag insertions larger than: [] bases
 - ☐ Filter alignments by read group: URL or path to filter file
- Splice Junction Track Options**
 - ☐ Show junction track Min flanking width: 0 Min junction coverage: 1
 - ☒ Show flanking regions
- Insert Size Options**

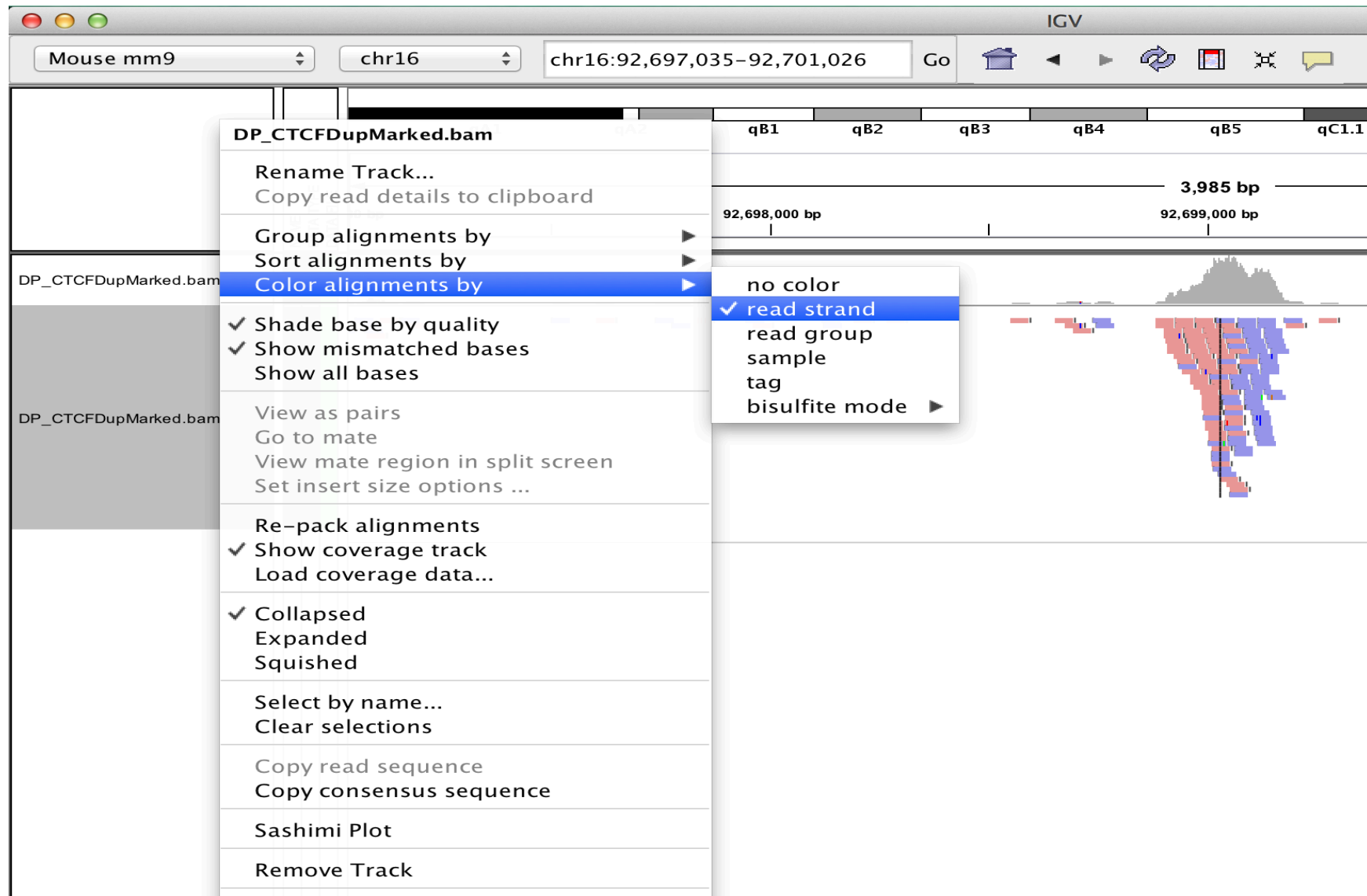
These options control the color coding of paired alignments by inferred insert size. Base pair values set default values. If "compute" is selected values are computed from the actual size distribution of each library.

Defaults	Minimum (bp): 50	☒ Compute	Minimum (percentile): 0.5
	Maximum (bp): 1000		Maximum (percentile): 99.5

Buttons: OK, Cancel

Track display options

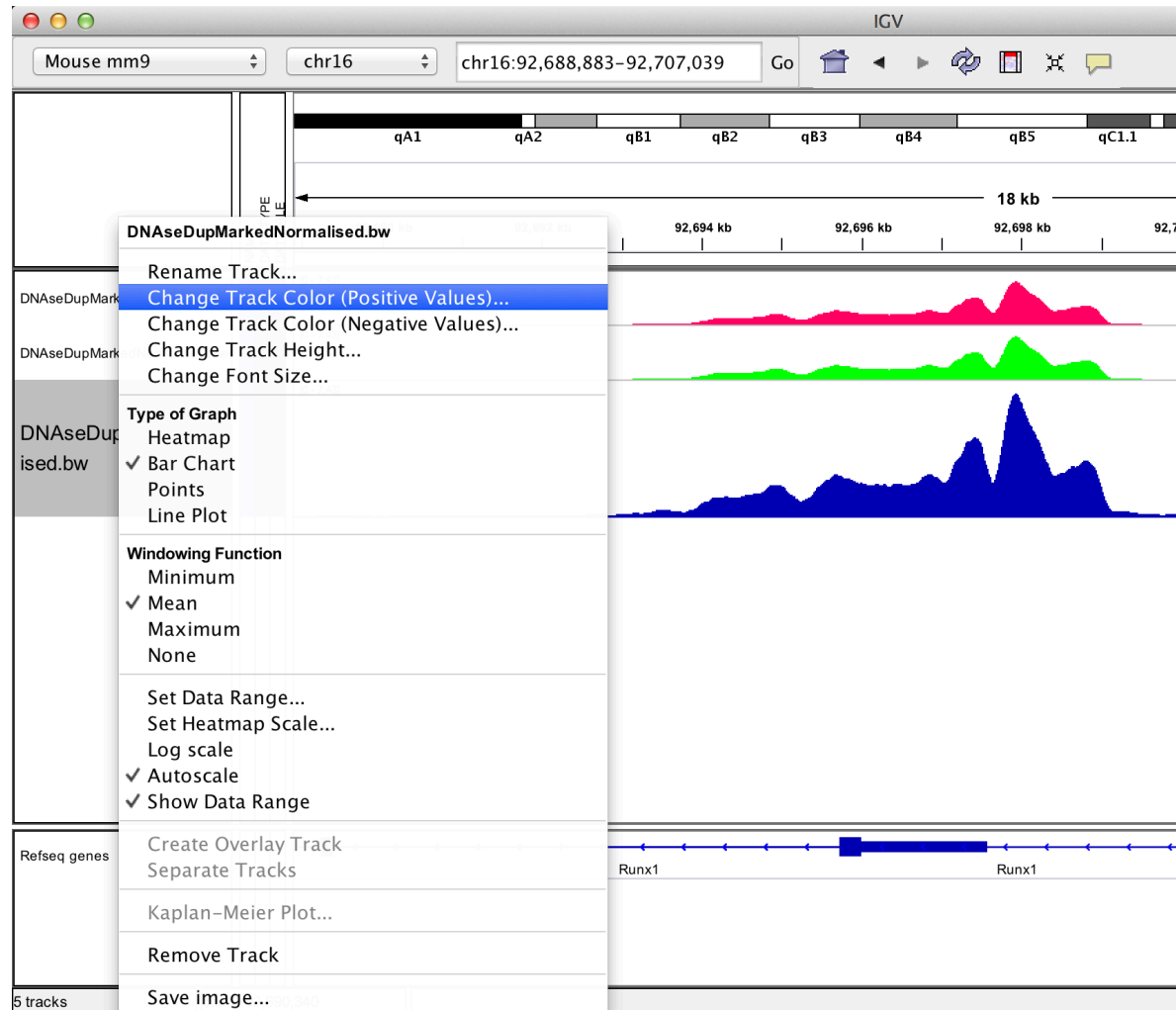
- Read Packing, grouping, sorting, colouring options.



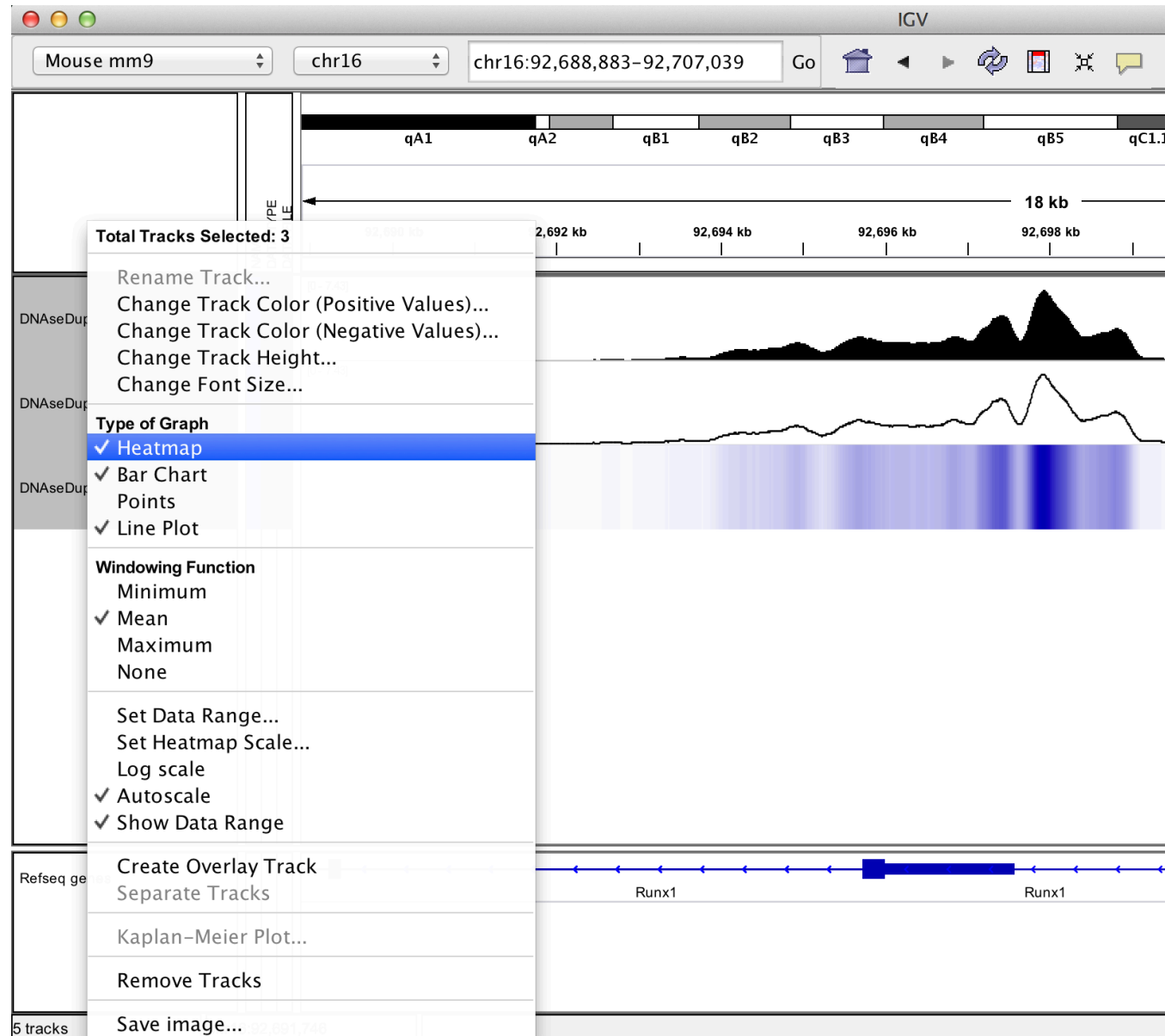
Track display options

Graph/interval files

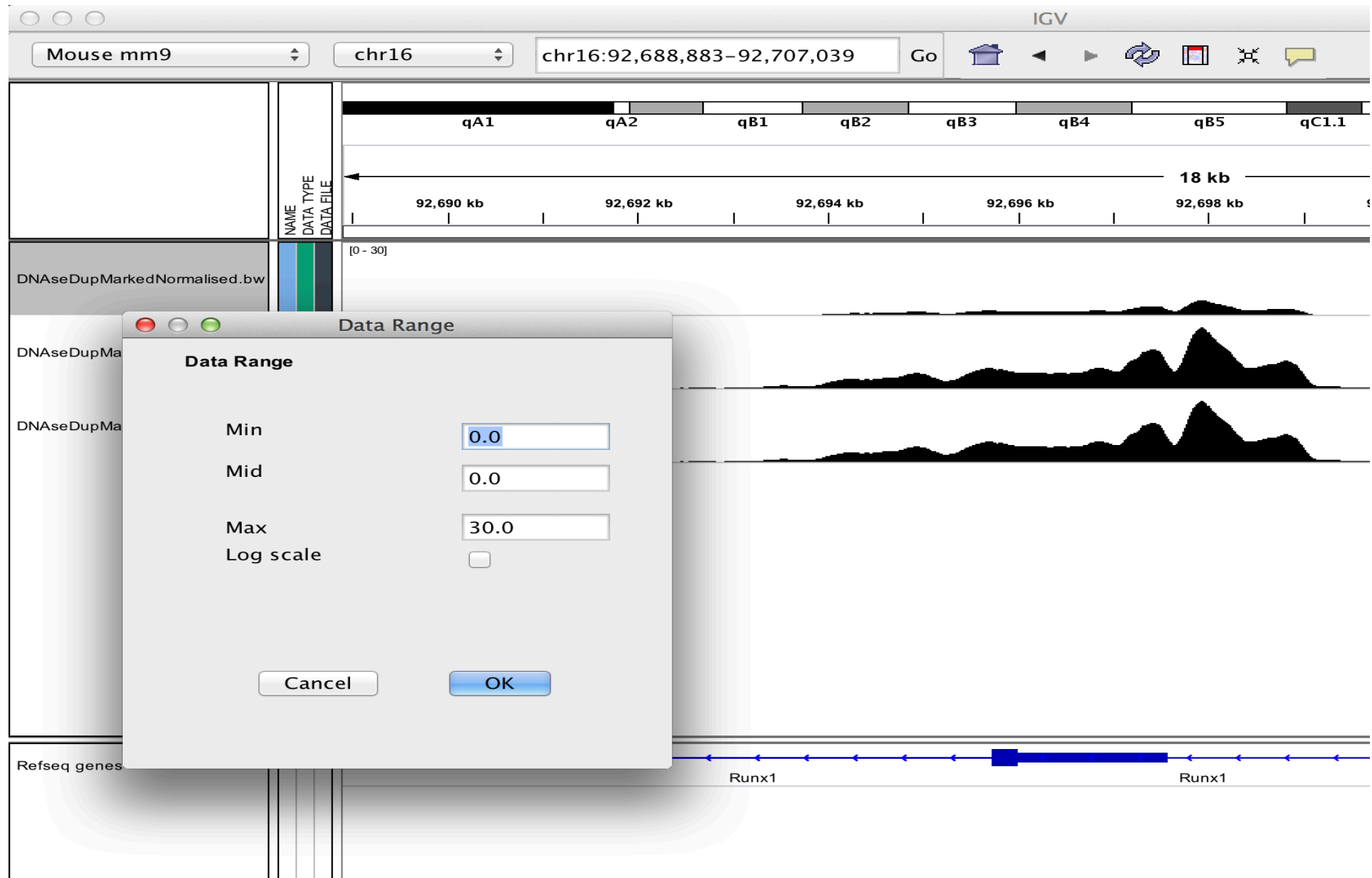
- Track colour/appearance



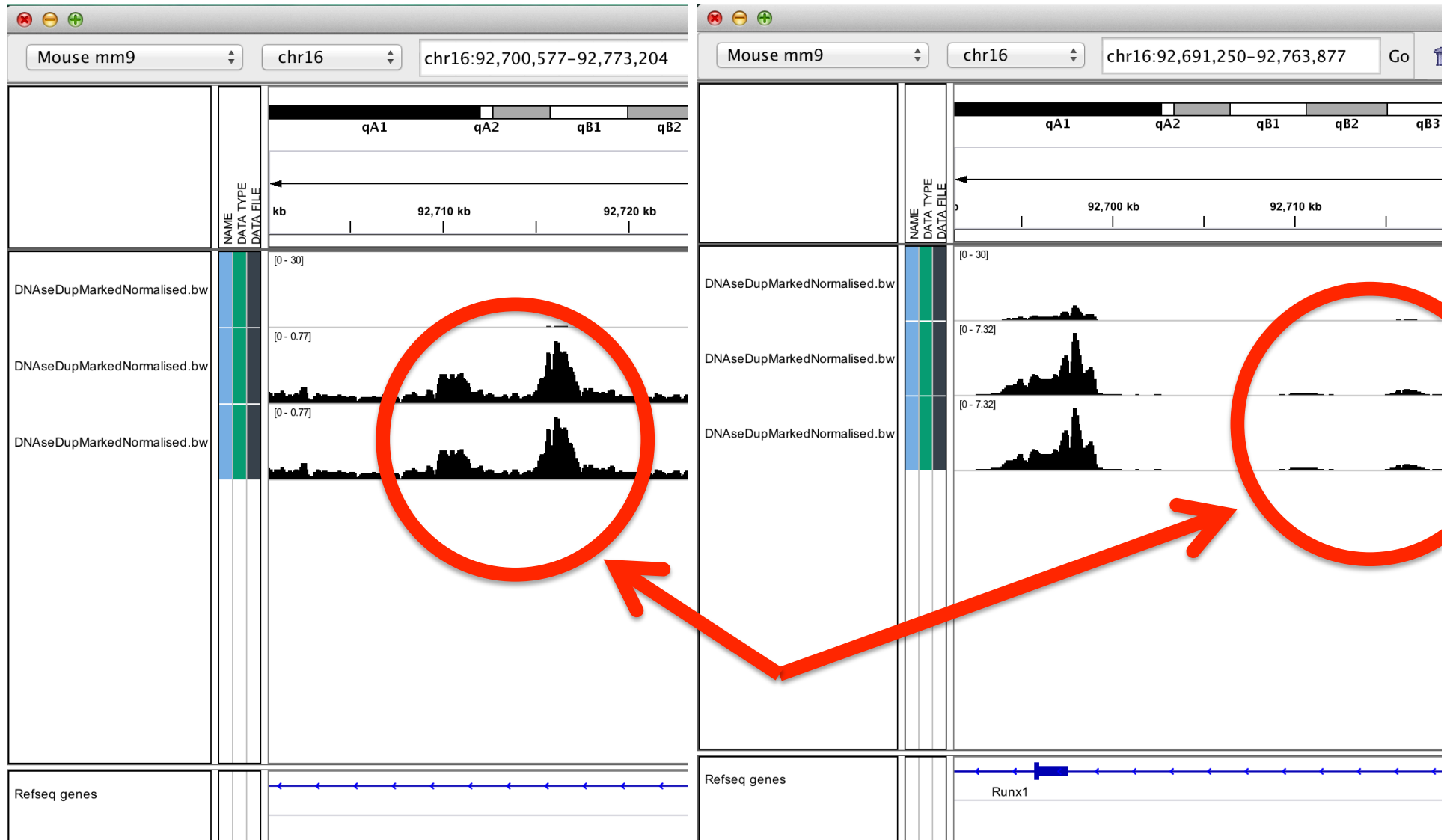
■ Graph type



Data Scaling.

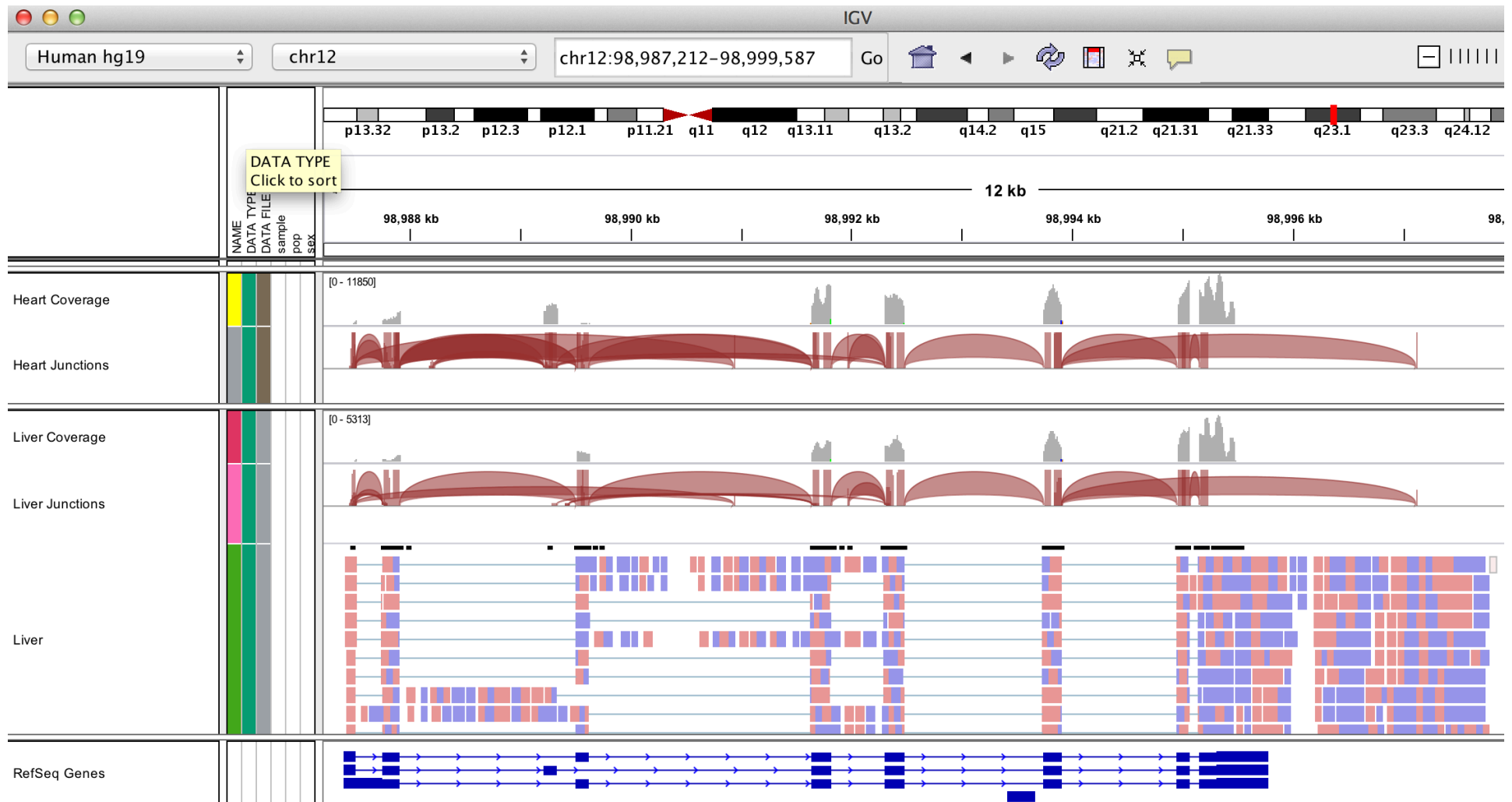


- Autoscaling adjusts to track's visible signal maximum



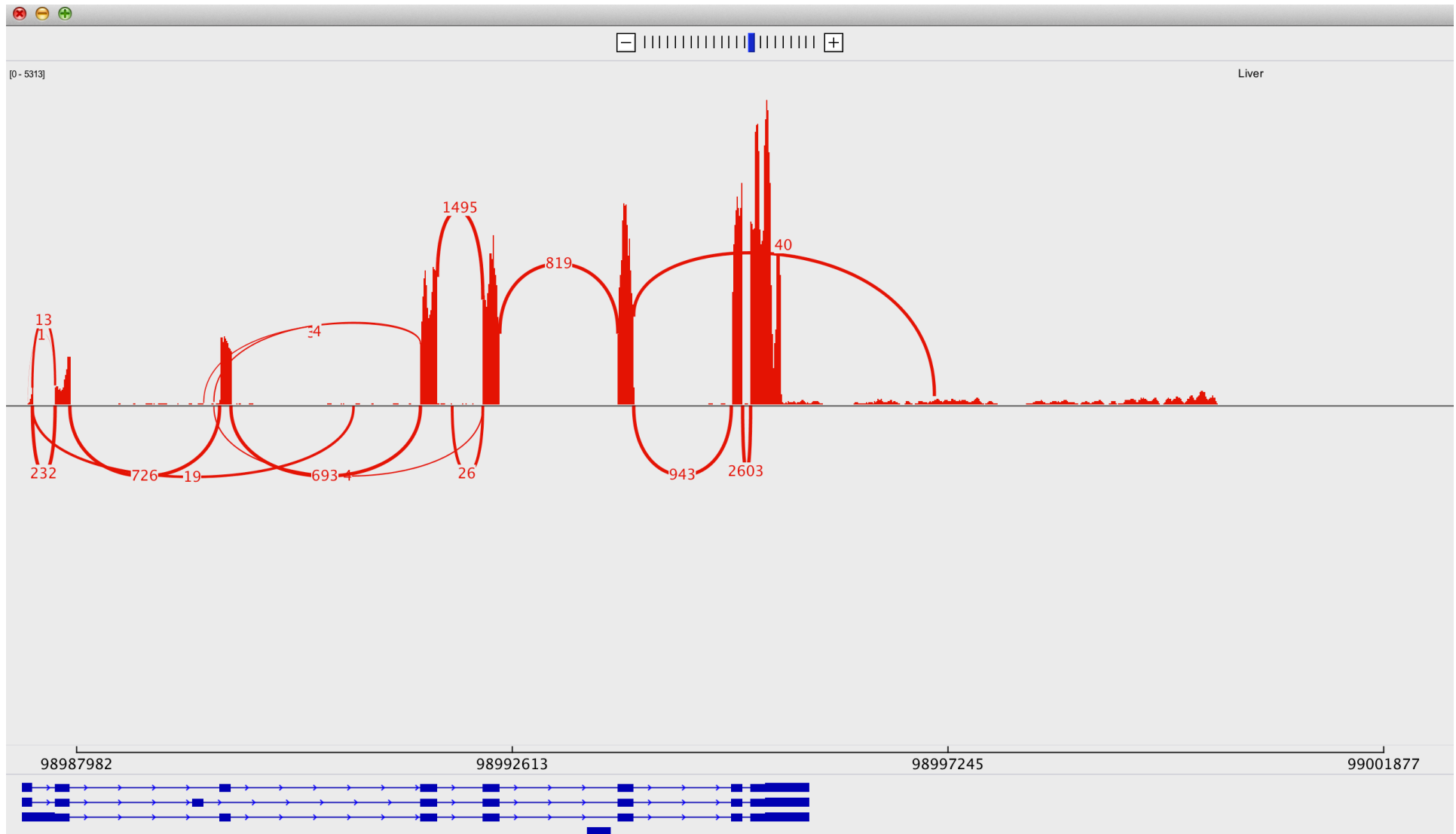
Some cool features.

IGV can display splicing information

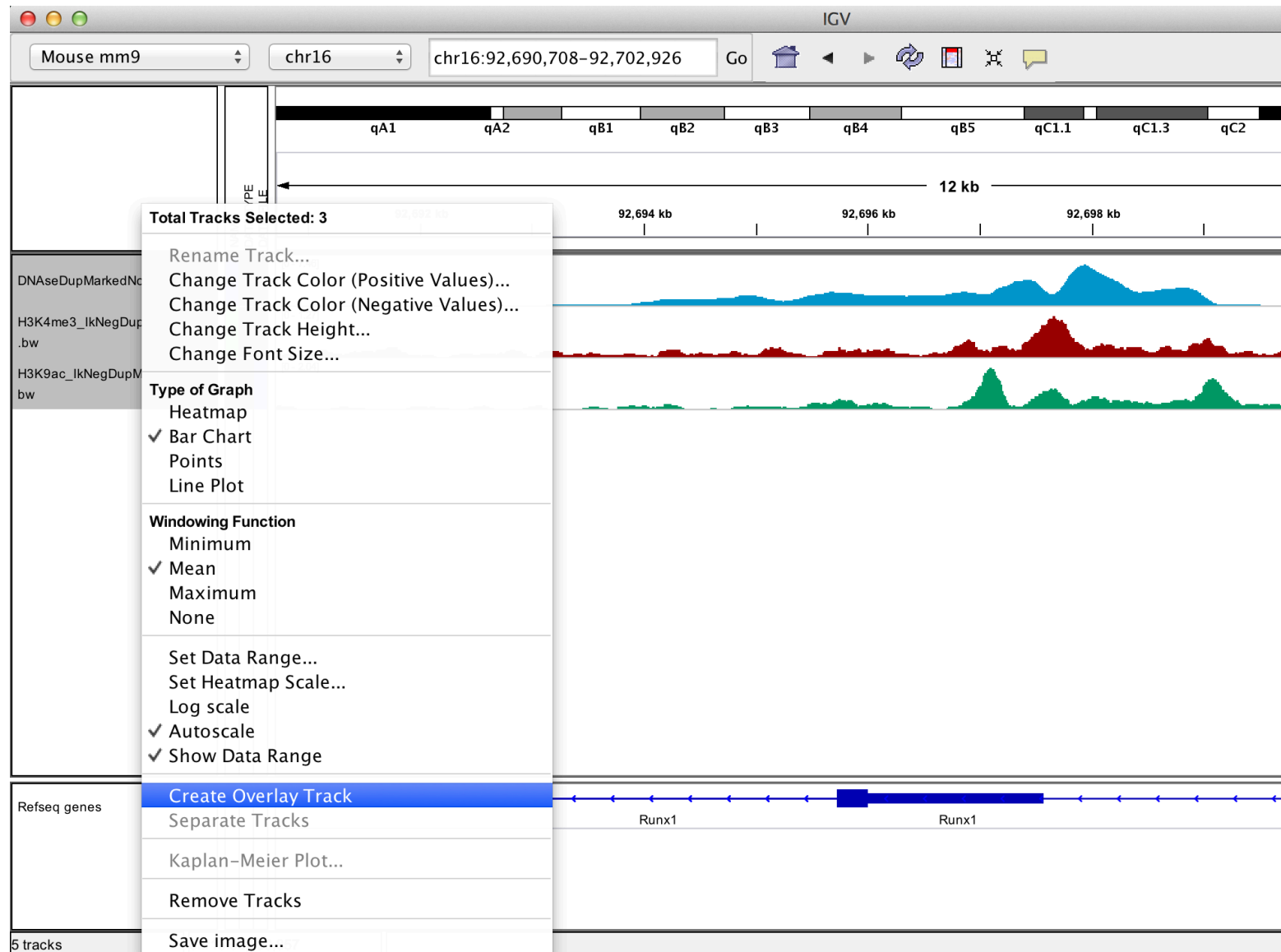


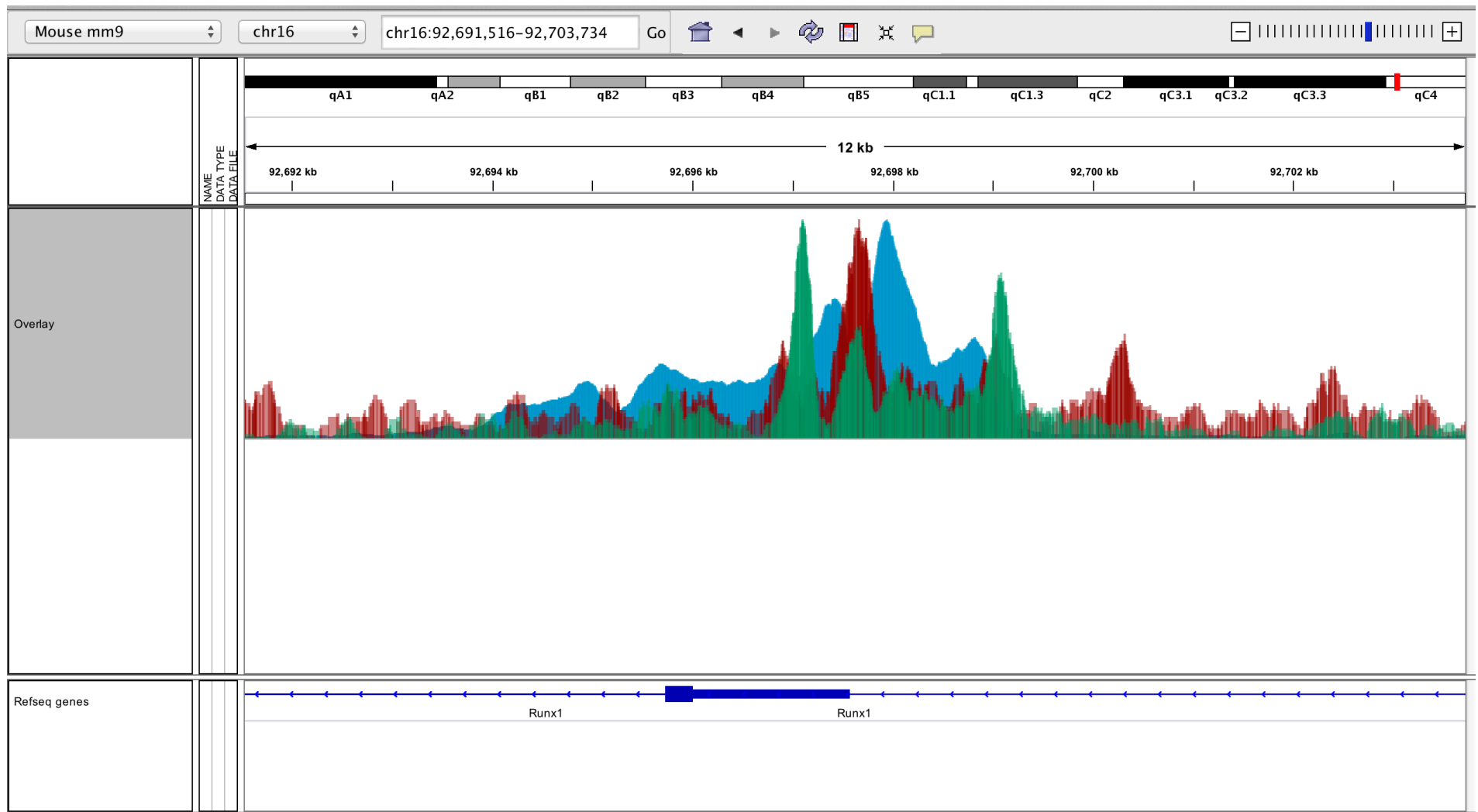
..and Sashimi plots

(<http://www.broadinstitute.org/igv/Sashimi>)



IGV can overlay tracks too.

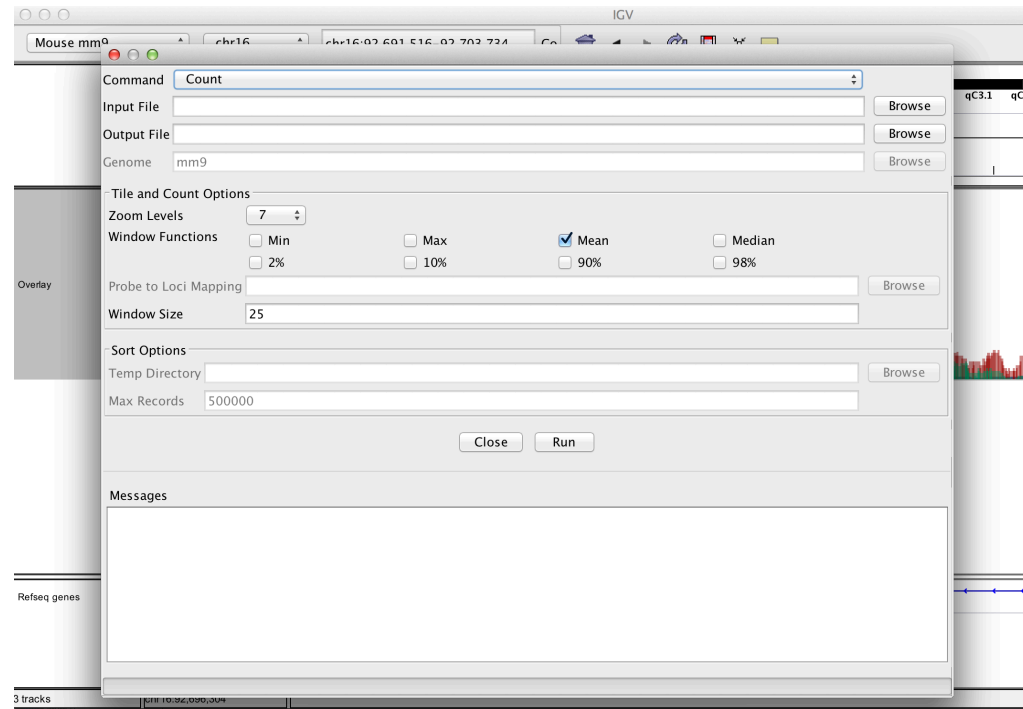




IGV includes a simple toolset

IGVTools

- IGVTools can be used to post-process genomics data.
- Includes indexing, sorting and genome graph creation.



Where to get help?

- <http://www.broadinstitute.org/igv/UserGuide>
- <https://groups.google.com/forum/#!forum/igv-help>